

Original Research

View Article online

 Check for updates

Received 25 July 2025

Revised 22 August 2025

Accepted 30 September 2025

Available Online: 17 January 2026

Edited by Kannan RR Rengasamy

KEYWORDS:

Antioxidant capacity

Antibacterial activity

Citrus macrocarpa

Essential oil

Natr Resour Human Health 2026; 6 (1): 280–286

<https://doi.org/10.53365/nrfhh/211518>

eISSN: 2583-1194

Copyright © 2026 Visagaa Publishing House

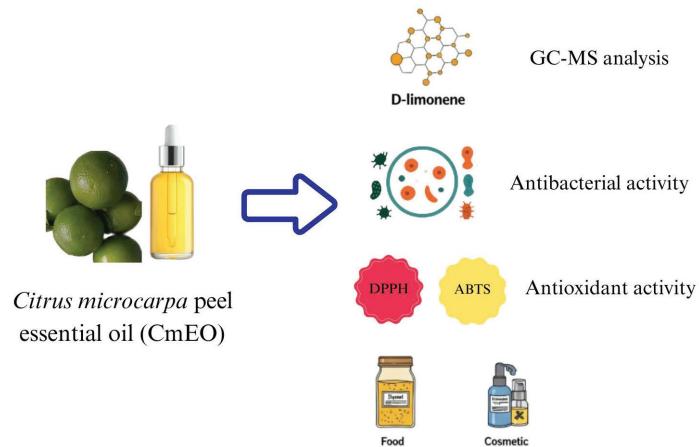
Calamondin (*Citrus microcarpa*) Peel Essential Oil: Physicochemical, Chemical, and Biological Properties

Nguyen Ngoc Thuan, Le Pham Tan Quoc*, Lam Bach Bao Phuong, Pham Thi Quyen

Institute of Biotechnology and Food Technology, Industrial University of Ho Chi Minh City, Viet Nam

ABSTRACT: Calamondin (*Citrus microcarpa*) peel essential oil (CmEO) contains bioactive compounds with potential uses in food preservation, cosmetics, and aromatherapy. This study investigates its chemical composition, antioxidant properties, and antibacterial activity to evaluate its potential as a natural preservative. CmEO was obtained through steam distillation, and its chemical composition was characterized by gas chromatography–mass spectrometry (GC-MS). Its antioxidant potential was evaluated using both DPPH and ABTS radical scavenging assays, while antibacterial effects were tested against *Staphylococcus aureus*, *Escherichia coli*, *Salmonella enteritidis*, and *Bacillus cereus* through the disk diffusion method. GC-MS analysis identified D-Limonene as the predominant compound, accounting for 95.56% of the total composition. The essential oil (EO) exhibited strong antioxidant activity, demonstrating remarkable antioxidant potential, achieving IC_{50} values of 9.46 mg/mL in the DPPH assay and 13.95 mg/mL in the ABTS assay. Antibacterial evaluation demonstrated notable antimicrobial efficacy, particularly against *Staphylococcus aureus*, evidenced by a distinct zone of inhibition measuring 15.73 mm. The findings suggest that CmEO possesses promising antioxidant and antibacterial properties, supporting its potential application as a natural preservative in food and other industries. The findings of this study demonstrate CmEO's promising use as a natural replacement for synthetic preservatives, supporting the advancement of safer and more sustainable preservation strategies.

GRAPHICAL ABSTRACT



* Corresponding author.

E-mail address: lephamtanquoc@iuh.edu.vn (Le Pham Tan Quoc)

This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. INTRODUCTION

Calamondin, or *Citrus microcarpa*, is a small citrus fruit widely cultivated in Southeast Asia, particularly in Vietnam. This fruit is highly prized for its uniquely tart and aromatic flavor, which makes it a popular culinary ingredient, medicinal component, and suitable for decorative applications (Alinejhad et al., 2016). It is a hybrid between *Citrus reticulata* and *Fortunella japonica*, characterized by its small round shape, thin peel, and high juice content (Lim, 2012).

The peel of calamondin is an important source of essential oil (CmEO), containing bioactive compounds with various biological effects. Studies have shown that CmEO mainly consists of monoterpenes such as D-Limonene, γ -Terpinene, and β -Pinene. Furthermore, the presence of oxygenated compounds, including linalool and citronellal, was noted (Chen et al., 2024). These components contribute to the citrus aroma of the essential oil (EO) and its functional properties, including antioxidant, antibacterial, and anti-inflammatory activities (Husni et al., 2020). It has been proven to be effective against common foodborne pathogenic bacteria, specifically *Escherichia coli* and *Staphylococcus aureus*, positioning it as a potential natural preservative (Rubiatul et al., 2015).

Beyond culinary applications, CmEO is gaining recognition in the cosmetic and aromatherapy industries because of its refreshing scent and skin-enhancing properties. Research indicates that it can inhibit melanin production, suggesting potential applications in skin-whitening and anti-aging products (Arifin et al., 2024). In addition, its calming and stress-relieving effects have been studied in aromatherapy treatments (Husnayanti et al., 2022).

In spite of its numerous benefits, calamondin peel is often considered a waste in Vietnam. The region has a long tradition of cultivating calamondin, yet the utilization of its peel remains largely untapped. Currently, most calamondin peels are discarded as byproducts after juice processing, leading to resource wastage. Therefore, the study about CmEO contributes to creating added value and jobs for local people. At the same time, this study also provides additional necessary data to supplement important information on this oil from different sources.

2. MATERIALS AND METHODS

2.1. Plant extraction

Calamondin was grown extensively in Vietnam's Dong Nai. Following harvest, calamondin fruit peels are used to extract EOs, and the fruit's juice is used to make food and

beverages. The extraction efficiency of the steam distillation process, which is run at 100°C for around 3 hours, is 1.2% (v/w). To maintain the EO's quality and inherent fragrant qualities, it is kept at room temperature in dark glass bottles.

2.2. Bacterial strains

The study employed four bacterial strains from the Institute of Biotechnology and Food Technology, Industrial University of Ho Chi Minh City. The strains consisted of two Gram-positive (*Staphylococcus aureus* - ATCC 33591 and *Bacillus cereus* - ATCC 11778) and two Gram-negative bacteria (*Escherichia coli* - ATCC 25922 and *Salmonella enteritidis* - ATCC 13076).

2.3. Experimental reagents

For this study, the following chemicals were employed: 2,2'-azinobis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS, $\geq 98\%$ purity, Sigma, USA), 2,2-diphenyl-1-picrylhydrazyl (DPPH, $\geq 97\%$ purity, Sigma, USA), and dimethyl sulfoxide (DMSO, $\geq 99.5\%$ purity, China). Analytical-grade reagents were employed alongside culture media for antibacterial tests, including nutrient agar (NA) and Mueller–Hinton agar (MHA) (HiMedia, India).

2.4. Characterization of the physical properties of CmEO

Physical properties of CmEO, such as relative density (RD), absolute density (AD), freezing point (FP), acid value (AV), ester value (EV), and saponification value (SV), were measured according to ISO standards: ISO 279 (1998), ISO 1041 (1973), ISO 1242 (2023), and ISO 7660 (1983).

2.5. Fragrance retention (FR)

The concentration and longevity of CmEO were used to evaluate its FR, following the procedure of Bandyopadhyay and Das (2017) with minor modifications. CmEO solutions were prepared at 20, 40, 60, 80, and 100% (v/v) by dilution in 96% ethanol. A few drops of each solution were applied to scent test paper, allowing the mixture to spread evenly after a brief waiting period. The time taken for the fragrance to completely dissipate under normal conditions was recorded to assess its retention.

2.6. Gas chromatography-mass spectrometry (GC-MS) analysis

A 1 μ L aliquot of the CmEO was injected into an Agilent 7890A GC system coupled with a 5977E MSD detector for chemical composition analysis. The analysis was performed using GC-MS (Agilent Technologies, USA). The compounds were separated on a Carbowax 20MTM capillary column (30 m \times 0.25 mm i.d., 0.25 μ m film thickness). The flow of the helium carrier gas was set to 10 mL/minute with a split ratio of 10:1. The injector temperature was 250°C. The oven program started with a 2-minute hold at 50°C, followed by an increase to 250°C at a rate of 10°C/minute (held for 5 minutes), and a final ramp to 280°C for 3 minutes. The resulting mass spectra were recorded using electron ionization (EI) at 70 eV.

2.7. Antioxidant capacity by DPPH assay

To evaluate the antioxidant capacity (AC) of CmEO, the free radical scavenging activity (RSA) was measured with a modified DPPH method, based on the procedure described by Quoc (2020). CmEO solutions at different concentrations were prepared in 96% ethanol. Then, 0.3 mL of the CmEO solution was combined with 2.7 mL of a 0.1 mM DPPH solution and incubated in the dark at room temperature for 30 minutes. The subsequent decrease in DPPH absorbance was measured at 517 nm using a spectrophotometer, with vitamin C serving as the reference standard. The inhibition percentage was calculated as a function of CmEO concentration, and the IC₅₀ value (concentration required to inhibit 50% of DPPH radicals) was determined. AC was subsequently calculated using the following equation:

$$\%DPPH_{RSA} = \frac{A_0 - A_{sample}}{A_0} \times 100$$

where A₀ represents the absorbance of the DPPH solution, and A_{sample} denotes the absorbance of the CmEO solution in the presence of the DPPH solution.

2.8. Antioxidant capacity by ABTS assay

The ABTS⁺ was determined by a modified version of the method described by Tsvetkova et al. (2023). A solution of ABTS⁺ (7 mM) and potassium persulfate (2.45 mM) was mixed in a 1:1 (v/v) ratio and incubated for 16 hours in the dark at room temperature to produce ABTS⁺ radicals. This stock solution was then diluted with distilled water to an absorbance of 0.70 \pm 0.02 at 734 nm. Finally, 0.1 mL of CmEO at various concentrations was combined with 3 mL of the ABTS⁺ solution, and ethanol was added to bring the total

volume to 5 mL. After incubation for 6 minutes in the dark, absorbance was measured at 734 nm. Inhibition percentage and IC₅₀ values were calculated, and AC was expressed using the corresponding formula:

$$\%ABTS_{RSA} = \frac{A_0 - A_{sample}}{A_0} \times 100$$

where A₀ is the absorbance of the ABTS⁺ solution without the sample, and A_{sample} is the absorbance of the CmEO solution in the presence of the ABTS⁺ solution

2.9. Evaluation of antibacterial properties

The antibacterial activity (AA) was assessed using the paper disk diffusion method, adapted from the procedure described by Al-Daihan and Bhat (2012) with some modifications. For the antibacterial assay, 100 μ L of a bacterial suspension at a density of approximately 1.5 \times 10⁸ CFU/mL (0.5 McFarland standard) was uniformly spread onto the MHA plates. Five μ L of the EO was loaded onto 6 mm sterile paper disks. The study used gentamicin (10 μ g/disc) as a positive control and dimethyl sulfoxide (DMSO) (5%, v/v) as a negative control. The plates were incubated at 37°C for 24 hours. Antibacterial efficacy was determined by measuring the diameter of the inhibition zones surrounding the paper disks.

2.10. Data analysis

Data analysis was performed using ANOVA, with post-hoc mean comparisons conducted using Statistics 20 software. A significance level of $p \leq 0.05$ was used, and the least significant difference (LSD) test was applied to assess differences. All results are reported as mean \pm standard deviation (mean \pm SD).

3. RESULTS AND DISCUSSION

3.1. Determination of physicochemical properties of CmEO

CmEO exhibits notable physical properties, with a characteristic pale yellow color, a density of 0.86 g/mL, and a FP below -18°C (Table 1). With a pH of 4.12, this CmEO is mildly acidic, making it suitable for applications in cosmetics and aromatherapy (Mehlich et al., 2021). However, for safe direct application on the skin, dilution is required. Research by Kamal et al. (2011) has also reported similar properties in other citrus EOs, such as the density of *Citrus reticulata*

Table 1.

Physicochemical properties of *Citrus microcarpa* peel essential oil.

No.	Physicochemical properties	Value
1	pH	4.12 ± 0.26
2	Freezing point (FP, °C)	< -18 °C
2	Relative density (RD)	0.8626 ± 0.0001
3	Absolute density (AD, g/mL)	0.8600 ± 0.0011
4	Acid value (AV, mg KOH/g EO)	0.8436 ± 0.0256
5	Saponification value (SV, mg KOH/g EO)	2.4664 ± 0.2303
6	Ester value (EV, mg KOH/g EO)	1.6228 ± 0.2057
7	Fragrance retention (FR, h):	
	20% EO	1.48 ± 0.38
	40% EO	2.39 ± 0.18
	60% EO	3.72 ± 0.27
	80% EO	4.98 ± 0.32
	100% EO	5.83 ± 0.29

(0.834 ± 0.02 g/cm³), *Citrus paradisi* (0.833 ± 0.02 g/cm³), and *Citrus sinensis* (0.833 ± 0.02 g/cm³) in Pakistan.

The AV of 0.8436 mg KOH/g and the SV of 2.4664 mg KOH/g of CmEO are relatively low, suggesting a high degree of purity with only minimal amounts of free fatty acids and complex esters (Belarbi et al., 2000). A low AV helps maintain a fresh fragrance but also means that the oil tends to evaporate quickly and has a short-lasting scent (Ba et al., 2013).

The longevity of CmEO's fragrance depends on its concentration. When used at 100% purity, it retains its scent for approximately 5.83 h. However, as it is diluted, the fragrance duration decreases: 80% (4.98 h), 60% (3.72 h), 40% (2.39 h), and 20% (1.48 h). Overall, even in its pure form, CmEO falls into the category of short-lasting EOs, with a scent duration ranging from 1.5 to 6 hours.

Because of its high volatility, CmEO is commonly used in fresh-scented products such as light perfumes, diffusers, and natural cosmetics. To enhance its longevity, it can be blended with carrier oils like jojoba or coconut oil or combined with long-lasting EOs such as sandalwood or patchouli to extend its diffusion time and improve scent retention (Chandran et al., 2017).

3.2. Chemical composition of CmEO

GC-MS analysis showed that D-Limonene is the major constituent of CmEO, representing 95.56% of its total volatile compounds (Table 2). This compound overwhelmingly dominates over other identified constituents such as β-Myrcene (2.61%), α-Pinene (1.50%), and Sabinene (0.33%). The high abundance of D-Limonene is characteristic of citrus EOs and is primarily responsible for the distinctive citrus aroma commonly associated with Citrus species. In addition to its

Table 2.

Chemical composition of *Citrus microcarpa* peel essential oil.

No.	Compounds	RT. (min)	Content (%)
1	α-Pinene	3.27	1.50
2	β-Myrcene	4.24	2.61
3	D-Limonene	4.59	95.56
4	Sabinene	4.69	0.33
	Monoterpene hydrocarbons		100.00

sensory contribution, D-Limonene exhibits a wide spectrum of biological and industrially valuable properties, with notable antioxidant, anti-inflammatory, antibacterial, and solvent properties, which makes it especially attractive for incorporation into cosmetic formulations, perfumery, and eco-friendly cleaning products (Han et al., 2019).

These findings align with previous reports, further supporting the chemical stability and reproducibility of CmEO compositions across different geographical regions. For example, Dao et al. (2022) investigated CmEO from the Ben Tre province in Vietnam and reported an even higher D-Limonene content of 96.93%, with other minor constituents such as β-Myrcene (1.43%) and α-Pinene (0.56%) present in comparable amounts. Similarly, Goncalves et al. (2018) analyzed CmEO samples from China and found that D-Limonene constituted 92.70% of the oil. The convergence of these findings supports the hypothesis that CmEO exhibits a relatively conserved chemotype, likely driven by the species' genetic makeup and the biosynthetic dominance of monoterpene hydrocarbons, particularly D-Limonene.

In conclusion, the consistent dominance of D-Limonene in CmEO, verified by multiple independent studies, underscores its robustness and versatility as a natural ingredient. The presence of supportive bioactive compounds further enhances its broad-spectrum applicability. Overall, these results emphasize CmEO as a promising candidate for the formulation of safe, multifunctional, and eco-friendly products, with potential applications spanning food preservation, cosmetics, health care, and household use.

3.3. Determination of AC of CmEO

The AC of CmEO was assessed by its free RSA, which was measured using both the DPPH and ABTS assays with IC₅₀ values compared to vitamin C. In the DPPH assay, vitamin C exhibited strong AC with an IC₅₀ of 4.83 µg/mL, significantly lower than CmEO (IC₅₀ = 9.46 mg/mL) (Table 3). This indicates that while CmEO can scavenge free radicals, its AC is significantly weaker than that of vitamin C. The difference may be attributed to the nature of the compounds

Table 3.Antioxidant capacity of *Citrus microcarpa* peel essential oil.

Test sample	IC ₅₀ -DPPH	IC ₅₀ -ABTS
Vitamin C (µg/mL)	4.83 ^a ± 0.28	4.98 ^a ± 0.11
CmEO (mg/mL)	9.46 ^b ± 0.14	13.95 ^b ± 0.43

Note: Different small letters in the same column indicate significant differences ($p \leq 0.05$) between samples.

in CmEO, which primarily consist of monoterpenes and sesquiterpenes—compounds with antioxidant properties but weaker than polyphenols or vitamin C (Ciesla et al., 2016).

Similarly, in the ABTS assay, vitamin C demonstrated superior AC ($IC_{50} = 4.98 \mu\text{g/mL}$), whereas CmEO had a significantly higher IC_{50} value (13.95 mg/mL). Compared to the DPPH method, the AC of CmEO was lower in the ABTS⁺ assay. This variation could be due to the different reaction mechanisms of the two assays: DPPH primarily reacts with hydrogen-donating compounds, whereas ABTS⁺ interacts with both hydrophilic and lipophilic antioxidants (Christodouleas et al., 2015).

Overall, although CmEO possesses antioxidant activity, it is considerably weaker than vitamin C. However, its moderate antioxidant potential makes it a viable candidate for applications in natural food preservation or cosmetics. To enhance its efficacy, CmEO could be combined with stronger antioxidants or optimized extraction methods to increase the bioactive compound content.

3.4. Determination of AA of CmEO

The antibacterial testing results for *E. coli*, *S. enteritidis*, *S. aureus*, and *B. cereus* revealed significant differences in the inhibitory effects of gentamicin and CmEO. For *S. aureus*, CmEO exhibited a relatively strong inhibition, with an inhibition zone diameter of 15.73 mm, which is close to that of gentamicin (16.57 mm) (Table 4). On the other hand, for *B. cereus*, both gentamicin and CmEO showed relatively weak inhibitory effects, with the diameters of the inhibition zones measuring 14.78 mm and 13.26 mm, respectively, indicating that neither was highly effective against this strain.

Meanwhile, *E. coli* and *S. enteritidis* were more strongly inhibited by gentamicin, with inhibition zone diameters of 20.40 and 20.45 mm, respectively, compared to 10.97 and 7.87 mm for CmEO. This demonstrates that gentamicin is a more effective antibacterial agent against these bacterial strains. However, the results also suggest that CmEO could be a viable alternative to gentamicin for treating *S. aureus* infections. A study by Iseppi et al. (2021) highlighted that combining EOs with antibiotics offers a potential therapeutic approach for combating antibiotic-resistant bacteria including

Table 4.Antibacterial activity of *Citrus microcarpa* peel essential oil.

Test strains	Diameter of the inhibitory zones of gentamicin (mm)	Diameter of the inhibitory zones of CmEO (mm)
<i>Escherichia coli</i>	20.40 ^{Cb} ± 0.47	10.97 ^{Ba} ± 0.70
<i>Salmonella enteritidis</i>	20.45 ^{Cb} ± 0.73	7.87 ^{Aa} ± 0.50
<i>Staphylococcus aureus</i>	16.57 ^{Bb} ± 0.38	15.73 ^{Da} ± 0.18
<i>Bacillus cereus</i>	14.78 ^{Ab} ± 0.44	13.26 ^{Ca} ± 0.26

Note: Within a row (a–b) or a column (A–D), different letters denote significant differences ($p \leq 0.05$) between samples or microorganisms, respectively.

those capable of surviving in biofilms. This approach could significantly reduce the required antibiotic dosage, thereby minimizing side effects and the risk of resistance.

The antibacterial mechanism of CmEO is likely associated with its major component, D-Limonene, which disrupts bacterial cell membranes by altering lipid membrane permeability and causing intracellular component leakage, ultimately inhibiting bacterial growth (Han et al., 2021). Although CmEO exhibits AA, it does not match the overall effectiveness of gentamicin. However, it may serve as a complementary treatment option, particularly for *S. aureus*, which has shown sensitivity to the EO.

4. CONCLUSION

This research offers important information on the physicochemical characteristics, AC, and antibacterial effects of CmEO. Its high D-Limonene content (95.56%) suggests promising applications in natural food preservation and cosmetic formulations. CmEO exhibited moderate antioxidant activity, with IC_{50} -DPPH of 9.46 mg/mL and IC_{50} -ABTS of 13.95 mg/mL. In addition, it demonstrated antibacterial efficacy, which resulted in inhibition zones with diameters between 7.87 and 15.73 mm. These findings indicate that CmEO is a viable bioactive compound with potential commercial applications.

ACKNOWLEDGEMENT

The authors extend their appreciation to all colleagues who supported this work.

AUTHOR CONTRIBUTIONS

Research concept and design were done by N.N.T. and L.P.T.Q.; collection and assembly of data were looked into by

N.N.T. and L.P.T.Q.; data analysis and interpretation were the responsibility of P.T.Q. and L.B.B.P.; writing the article was done by L.B.B.P. and L.P.T.Q.; critical revision of the article was accomplished by P.T.Q. and N.N.T.; final approval of the article was the responsibility of N.N.T. and L.P.T.Q. All authors have approved the manuscript for publication.

CONFLICTS OF INTEREST

The authors declare that there is no conflict of interest.

DATA AVAILABILITY

All data generated or analyzed during this study are included in this published article.

FUNDING

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

ORCID

Nguyen Ngoc Thuan	0000-0003-2797-3139
Le Pham Tan Quoc	0000-0002-2309-5423
Lam Bach Bao Phuong	0009-0004-4152-609X
Pham Thi Quyen	0000-0003-3695-3703

REFERENCES

Al-Daihan, S., & Bhat, R.S. 2012. Antibacterial activities of extracts of leaf, fruit, seed and bark of *Phoenix dactylifera*. African Journal of Biotechnology 11, 10021–10025. <https://doi.org/10.5897/AJB11.4309>

Alinejihad, D., Asayesh, M.A., & Asayesh, M. 2016. Determination of the anti-inflammatory property of tannins from the rind of calamansi (*Citrus microcarpa*, Rutaceae). Journal of International Oral Health 8, 546–553.

Arifin, A., Nada, S.Y., Mawarni, R.S., & Adi, A.V. 2024. Formulasi dan uji aktivitas antioksidan sediaan sheet mask dari sari buah lemon cui (*Citrus microcarpha* B.) sebagai anti-aging. Sasambo Journal of Pharmacy 5, 26–31. <https://doi.org/10.29303/sjp.v5i1.322>

Ba, H.V., Amna, T., & Hwang, I. 2013. Significant influence of particular unsaturated fatty acids and pH on the volatile compounds in meat-like model systems. Meat Science 94, 480–488. <https://doi.org/10.1016/j.meatsci.2013.04.029>

Bandyopadhyay, S., & Das, D. 2017. Retention and sustained release of fragrance by cyclodextrin functionalized cotton fabric modified using maleic anhydride. Flavour and Fragrance Journal 32, 207–211. <https://doi.org/10.1002/ffj.3377>

Belarbi, E.H., Molina, E., & Chisti, Y. 2000. A process for high yield and scaleable recovery of high purity eicosapentaenoic acid esters from microalgae and fish oil. Enzyme and Microbial Technology 26, 516–529. [https://doi.org/10.1016/S0141-0229\(99\)00191-X](https://doi.org/10.1016/S0141-0229(99)00191-X)

Chandran, J., Nayana, N., Roshini, N., & Nisha, P. 2017. Oxidative stability, thermal stability and acceptability of coconut oil flavored with essential oils from black pepper and ginger. Journal of Food Science and Technology 54, 144–152. <https://doi.org/10.1007/s13197-016-2446-y>

Chen, Y., Liang, Z., Chen, S., Yan, F., He, J., Zhou, Y., & Gao, T. 2024. Unveiling the floral scent dynamics of calamondin (*Citrus × microcarpa*) across developmental stages. Separations 11, 359. <https://doi.org/10.3390/separations11120359>

Christodouleas, D.C., Fotakis, C., Nikokavoura, A., Papadopoulos, K., & Calokerinos, A.C. 2015. Modified DPPH and ABTS assays to assess the antioxidant profile of untreated oils. Food Analytical Methods 8, 1294–1302. <https://doi.org/10.1007/s12161-014-0005-6>

Ciesla, L.M., Wojtunik-Kulesza, K.A., Oniszczuk, A., & Waksmundzka-Hajnos, M. 2016. Antioxidant synergism and antagonism between selected monoterpenes using the 2,2-diphenyl-1-picrylhydrazyl method. Flavour and Fragrance Journal 31, 412–419. <https://doi.org/10.1002/ffj.3330>

Dao, P.T., Nguyen, M.V., Tran, Q.N., Le, D.T., Tran, T.Y.N., & Lam, T.V. 2022. Experimental and kinetic modeling studies on extraction of essential oil from Vietnamese calamondin (*Citrus microcarpa*) by hydro-distillation process. Iranian Journal of Chemistry & Chemical Engineering 4, 3824–3834.

Goncalves, D., Koshima, C.C., Batista, F.R.M., & da Costa Rodrigues C.E. 2018. Performance of continuous countercurrent extractor on the fractionation of *Citrus bergamia* essential oil using ethanol/water mixtures as solvents. Chemical Engineering Research and Design 137, 566–576. <https://doi.org/10.1016/j.cherd.2018.08.008>

Han, Y., Sun, Z., & Chen, W. 2019. Antimicrobial susceptibility and antibacterial mechanism of limonene against *Listeria monocytogenes*. Molecules 25, 33. <https://doi.org/10.3390/molecules25010033>

Han, Y., Chen, W., & Sun, Z. 2021. Antimicrobial activity and mechanism of limonene against *Staphylococcus aureus*. Journal of Food Safety 41, 12918. <https://doi.org/10.1111/jfs.12918>

Husnayanti, A., Sinulingga, S.R., & Sudirman, M. 2022. Aromatherapy candle formulation from *Citrus microcarpa* peel. Pharmacy Reports 2, 49. <https://doi.org/10.51511/pr.49>

Husni, E., Ismed, F., & Afriyandi, D. 2020. Standardization study of simplicia and extract of calamondin (*Citrus microcarpa* Bunge) peel, quantification of hesperidin and antibacterial assay. Pharmacognosy Journal 12, 777–783. <https://doi.org/10.5530/pj.2020.12.111>

Iseppi, R., Mariani, M., Condò, C., Sabia, C., & Messi, P. 2021. Essential oils: a natural weapon against antibiotic-resistant bacteria responsible for nosocomial infections. Antibiotics 10, 417. <https://doi.org/10.3390/antibiotics10040417>

ISO 1041, 1973. Essential oils—Determination of freezing point. International Organization for Standardization, Geneva, Switzerland.

ISO 279, 1998. Essential oils—Determination of relative density at 20°C. International Organization for Standardization, Geneva, Switzerland.

ISO 1242, 2023. Essential oils—Determination of acid value by two titration methods, manual and automatic. International Organization for Standardization, Geneva, Switzerland.

ISO 7660, 1983. Essential oils—Determination of ester value of oils containing difficult-to-saponify esters. International Organization for Standardization, Geneva, Switzerland.

Kamal, G.M., Anwar, F., Hussain, A.I., Sarri, N., & Ashraf, M.Y. 2011. Yield and chemical composition of Citrus essential oils as affected by drying pretreatment of peels. International Food Research Journal 18, 1275–1282.

Lim, T.K. 2012. *Citrus x microcarpa. Edible medicinal and non-medicinal plants*. Springer: Dordrecht, Netherlands. 865–870.

Mehlich, A., Doberenz, C., & Janssens-Böcker, C. 2021. Acidification of the skin and maintenance of the physiological skin pH value by buffered skin care products formulated around pH 4. Journal of Cosmetics, Dermatological Sciences and Applications 11, 44. <https://doi.org/10.4236/jcdsa.2021.111005>

Quoc, L.P.T. 2020. Physicochemical properties and antibacterial activity of essential oil of *Ageratum conyzoides* L. leaves. Agriculturae Conspectus Scientificus 85, 139–144.

Rubiatul, A.S., Helya, I.N., Zarina, Z., Dachyar, A., & NurulAin, H.A. 2015. Antibacterial properties of limau kasturi (*C. microcarpa*) peels extract. Advances in Environmental Biology 9, 5–10.

Tsvetkova, D., Kostadinova, I., Landzhov, B., Vezenkov, L., Marinov, L., & Ivanova, I. 2023. Application of ABTS method for assessment of radical-binding effect of creatine monohydrate. Journal of Advanced Pharmacy Education and Research 13, 92–98. <https://doi.org/10.51847/rxgOGbIUkJ>