

Original Research

View Article online



Received 17 April 2025

Revised 25 May 2025

Accepted 7 June 2025

Available Online 25 October 2025

Edited by Kannan RR Rengasamy

KEYWORDS:

Citrullus colocynthis (L.)
Histochemical Analysis
Pharmacognosy
Physicochemical
Purification
Siddha Medicine

Natr Resour Human Health 2025; 5 (4): 776–785

<https://doi.org/10.53365/nrfhh/206019>

eISSN: 2583-1194

Copyright © 2025 Visagaa Publishing House

Pharmacognostic Insights into *Citrullus colocynthis* (L.): Histochemical Analysis of Purification Process in Siddha Medicine

Karpagambal Ramamoorthy¹, Abarna Balasubramani¹,
Raghavi Marimuthu¹, Kalaarasi Angappan², Suresh Ramasamy³,
Manjari Venkatraman¹

¹Department of Nanju Maruthuvam (Siddha Toxicology), National Institute of Siddha (Affiliated with The Tamil Nadu Dr. M.G.R. Medical University), Ministry of Ayush, Chennai, Tamil Nadu, India

²Department of Siddhar Yoga Maruthuvam, Government Siddha Medical College (Affiliated with The Tamil Nadu Dr. M.G.R. Medical University), Ministry of Ayush, Chennai, Tamil Nadu, India

³Department of Gunapadam (Siddha Pharmacology), National Institute of Siddha (Affiliated with the Tamil Nadu Dr. M.G.R. Medical University), Ministry of Ayush, Chennai, Tamil Nadu, India

ABSTRACT: *Citrullus colocynthis* (L.) Schrad., commonly known as colocynth, is a member of the cucurbitaceae family predominantly found in arid regions. Its fruit is a key source of bioactive compounds used in the treatment of various ailments. In the Siddha system of medicine, purification is a crucial process employed to reduce toxicity and enhance therapeutic efficacy. This study evaluates the pharmacognostic differences in *C. colocynthis* (L.) fruit pulp before and after purification, with a focus on organoleptic characteristics, physicochemical parameters, and histochemical composition. Comparative analysis revealed that purification removed or degraded the potentially toxic constituents such as tannins, cutin, and resins, while also enriching beneficial bioactive compounds like phenolics. Key therapeutic constituents, including mucilage, alkaloids, oil globules, and lignin were retained, indicating selective histochemical modification. Physicochemical analysis further confirmed that the purification process improved the overall quality and stability of the sample, enhancing its medicinal potency. These findings provide valuable insights into the optimization of *C. colocynthis* (L.) for safe and effective medicinal use, while also laying the groundwork for future pharmacological research.

1. INTRODUCTION

Citrullus colocynthis (L.), commonly known as bitter apple or desert gourd, is a medicinal plant widely used in traditional medicine for its potent therapeutic properties. It is rich

in bioactive compounds such as cucurbitacins, flavonoids, and alkaloids contributing to its pharmacological activities, including anti-inflammatory, antimicrobial, and antidiabetic effects (Rahimi et al., 2012). The Drugs and Cosmetics Act classifies *C. colocynthis* (L.) as a Schedule E drug due to its

* Corresponding author.

E-mail address: karpagambsms97@gmail.com (Karpagambal Ramamoorthy)

This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

potent medicinal properties and potential toxicity (Juliet et al., 2021). In Siddha medicine, this plant, commonly known as *kalingam* or *varithumatti*, is traditionally used for treating *vata* disorders, menorrhagia, amenorrhea, and uterine disorders. *C. colocynthis* (L.), while beneficial in treating digestive disorders, joint pain, and skin diseases, must be used with caution to avoid adverse effects such as severe gastrointestinal irritation (Espinosa et al., 2022; Murugesamudhaliyaar, 2013). However, the raw form of *C. colocynthis* (L.) is known for its intense bitterness and potential toxicity, necessitating purification processes to enhance its safety and efficacy. The formulations containing *C. colocynthis* (L.) in Siddha medicine are recommended only after purification. The process of purification is traditionally employed to reduce toxicity and improve the bioavailability of its active constituents. The preliminary study of changes taking place during purification was assessed using physicochemical and pharmacognostic histochemical evaluation.

Pharmacognosy, the study of medicinal drugs derived from natural sources, plays a crucial role in analyzing the structural, physical, and chemical characterization, including isolation, identification, and quantification, of drug plant constituents (Alamgir, 2017). Histochemistry identifies and analyzes chemical distribution in biological cells using stains, indicators, and electron microscopy. It enables chemical characterization while preserving structural organization. In plant secretory structures, a detailed analysis is essential, as glands and cells can produce multiple metabolites simultaneously (Badria and Aboelmaaty, 2019).

Of interest here is to examine the pharmacognostical variations of *C. colocynthis* (L.) before and after purification, focusing on alterations in physicochemical and histochemical constituents. Understanding these transformations during the purification of *C. colocynthis* (L.) is essential for optimizing its therapeutic applications while ensuring its safety for medicinal use.

2. MATERIALS AND METHODOLOGY

2.1. Identification and authentication

The fruit of *C. colocynthis* (L.) was collected from Karunkulam, Tirunelveli district, during the spring season. The identification and authentication were done at the Department of Pharmacognosy, Siddha Central Research Institute, Arumbakkam, Chennai. The authentication code is C14072301C. The dried straw of *Paspalum scrobiculatum* L. (kodo millet) was authenticated at the Department of Medicinal Botany, National Institute of Siddha, Chennai. The authentication code is NISMB6472024.

2.2. Purification method

The purification process of *C. colocynthis* (L.) fruit was done after obtaining authentication. Then it was burned with the straw of kodo millet (*varagu vaikkol*) until soft, after which its outer layer of the peel and seeds were removed. After that, the pulp was collected and properly stored. This pulp was used for medicine preparation (Figure 1) (Juliet et al., 2021; Murugesamudhaliyaar, 2013). The two samples used for analysis were raw unpurified and purified *C. colocynthis* (L.) pulp.

2.3. Organoleptic characterization

The organoleptic characterization of *C. colocynthis* (L.) was carried out in line with the Pharmacopoeial Laboratory for Indian Medicine (PLIM) guidelines. The physical state of the samples was identified through direct visual inspection. For this, 5 g of each sample—unpurified and purified *C. colocynthis* (L.) pulp—were placed in clean glass containers and visually assessed for color. Each sample was then transferred to a separate beaker, where its odor was analyzed (Lohar, 2008; Ministry of Ayush, 2018).

2.4. Physicochemical analysis

The physicochemical parameters—including potential of hydrogen (pH), loss on drying (LOD) at 105°C, total ash content, water-soluble ash, and acid-insoluble ash as well as water and alcohol-soluble extractive values—were evaluated for both unpurified and purified pulp of *C. colocynthis* (L.) in accordance with PLIM guidelines (Lohar, 2008; Ministry of Ayush, 2018). These tests were analyzed in the Department of Chemistry at the Siddha Central Research Institute, Arumbakkam, Chennai.

2.5. Pharmacognostic study of histochemical analysis

Fruit sections were treated according to the standard procedures outlined in Table 1 (Fahn, 1980; Khandelwal, 2008; Wallis, 1965).

3. RESULTS AND DISCUSSION

The organoleptic characteristics of *C. colocynthis* before and after purification are presented in Table 2. The physicochemical parameters of unpurified and purified *C. colocynthis* (L.) pulp are provided in Table 3. The histochemical analysis

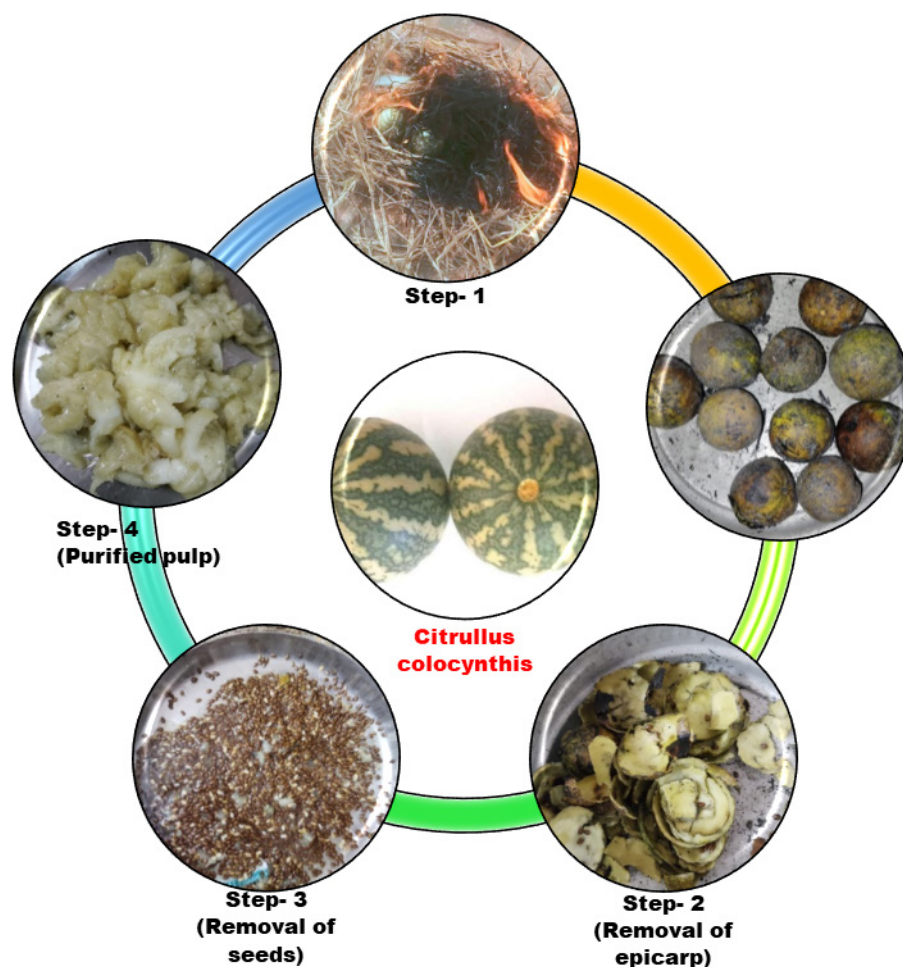


Figure 1. Purification process of *Citrullus colocynthis* (L.).

Table 1

Procedures for histochemical analysis.

Crystals	The section of both unpurified and purified pulp of <i>C. colocynthis</i> (L.) was mounted in water and one end of the cover slip was irrigated with acetic acid. While looking through the microscope, the water within the coverslip was replaced using a piece of filter paper at the opposite end of the coverslip. • Formation of air bubbles indicated calcium carbonate crystals. • If no air bubbles were formed, the experiment was repeated with conc. HCl, wherein dissolution of crystal and formation of needles of calcium sulphate indicated the presence of calcium oxalate crystals.
Fatty oils and resins	Approximately one to two drops of Sudan-IV were added to both pulp sections and left to stand for a few minutes. The presence of fatty oil substances was confirmed by the appearance of orange-red, pink, or red-colored globules, whereas red-colored irregular contents indicated the presence of resin.
Starch	A drop of 2% iodine water solution was added. Blue color indicated starch.
Phenolic compounds	A drop of alcoholic ferric chloride was added to bluish-black-colored contents indicated phenolic compounds like flavonoids/tannins and more.
Mucilage	A drop of ruthenium red was added—pink to red colored contents indicated mucilage.
Lignified cell walls	A drop of phloroglucinol was applied to the section and left to stand for approximately 2 minutes or until it nearly dried. Then, a drop of 50% HCl was added and the sample was observed under a cover glass. The cell walls stained pink to cherry red indicating the presence of lignin.
Cutin or suberin (cuticular cell walls or suberized cell walls)	A drop of Sudan red III was added and allowed to sit for a few minutes, with gentle warming if necessary. Cell walls stained orange-red or red indicated suberin or cutin deposition over cell wall.
Alkaloids	The addition of a drop of Wagners reagent resulted in the appearance of yellow to reddish-brown-colored contents, confirming the presence of alkaloids. This color change is indicative of the reaction between Wagners reagent and the alkaloids present in the sample, providing a clear visual indication of their presence.

Table 2

Organoleptic characteristics of *Citrullus colocynthis* before and after purification.

Characters	Before purification of <i>Citrullus colocynthis</i> (L.)	After purification of <i>Citrullus colocynthis</i> (L.) pulp
Color	Greenish with white	Dull white
Odor	Bitter odor	Bitter odor
Taste	Bitter	Bitter
Appearance	Hard and smooth surface	Softened and succulent nature

Table 3

Physicochemical parameters of unpurified and purified *Citrullus colocynthis* (L.) pulp.

S. No	Parameters	Unpurified sample Mean value	Purified sample Mean value
1	pH	5.23	4.82
2	Loss on drying	92.035%	92.935%
3	Total ash	4.515%	2.39%
4	Water-soluble ash	3.395%	1.85%
5	Acid-insoluble ash	0.701%	0.395%
6	Water-soluble extractive	4.37%	2.26%
7	Alcohol-soluble extractive	3.965%	2.415%

Table 4

Histochemical analysis of *Citrullus colocynthis* before purification.

Epicarp region	Cutin on epicarp walls and alkaloid present in epicarp cells.
Mesocarp region	Tannin deposition in mesocarp region, lignin in stone cells and xylem strands of mesocarp, and alkaloid detected in mesocarp cells.
Parenchyma cells of mesocarp	Mucilage present in the phloem cells and mucilage and also oil globules observed in parenchyma cells of mesocarp.

of *C. colocynthis* before purification is shown in Table 4 and Figure 2, while the histochemical analysis after purification is presented in Table 5 and Figure 3.

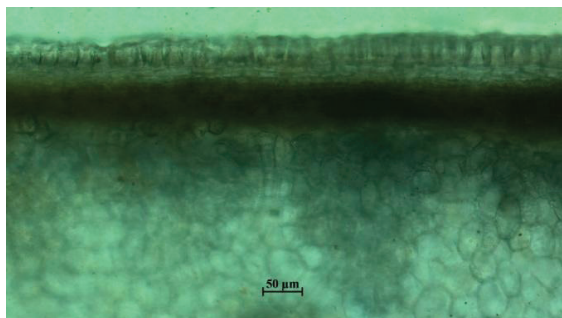
Observed through histochemical characterization, the purification process of *C. colocynthis* (L.) significantly alters its physical and chemical properties. The organoleptic evaluation revealed distinct changes in the pulps color, texture, and consistency. Before purification, the pulp exhibited a greenish-white hue, a hard texture, and a smooth surface. After purification, these attributes transformed, resulting in a dull white, softer, and more succulent pulp. Despite these physical changes, the characteristic bitter odor and taste remained

unaltered, indicating the persistence of certain phytochemical constituents even after purification. The increased succulence in the purified pulp is attributed to its substantial mass content, which constitutes nearly half of the total fruit mass (Rao and Poonia, 2023).

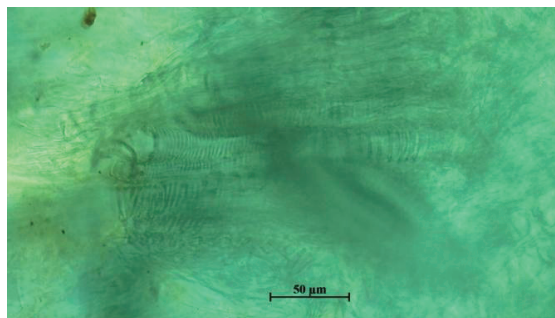
The pH level plays a vital role in assessing the quality of medicinal formulations, as it influences both chemical reactions and microbial activity. In this study, the pH of unpurified *C. colocynthis* pulp was 5.23, which slightly dropped to 4.82 after purification, indicating an increase in acidity. According to Allen (2003), herbal formulations with a low pH, due to the presence of acidic compounds, tend to have reduced bacterial counts. Conversely, higher or neutral pH values can support microbial growth. Optimal bacterial proliferation in medicinal products typically occurs within a pH range of 5.0–8.5. Therefore, the lower pH of the purified pulp may help inhibit microbial growth, making it more suitable for medicinal applications (Allen, 2003; Kumadiah et al., 2024). The moisture content, measured as LOD, was 92.035% in unpurified pulp and slightly increased to 92.935% in the purified sample. This rise may result from the removal of solids or breakdown of complex molecules during processing, allowing the pulp to retain more water. Given that *C. colocynthis* inherently contains more than 90% moisture, this increase is consistent with its natural composition; moisture also contributes to elevated absorption values (Hussain et al., 2014; Ibrahim et al., 2021). The total ash content, representing the inorganic matter in the sample, decreased significantly from 4.515% in the unpurified pulp to 2.39% after purification. This decline suggests that the purification process effectively removes a considerable portion of the mineral content. The low ash levels also indicate reduced contamination in the purified sample. Water-soluble ash, a fraction of the total ash that dissolves in water, was reduced from 3.395% to 1.85% post-purification. This reduction suggests that some water-soluble minerals or salts are eliminated during processing. Acid-insoluble ash, which indicates the presence of insoluble impurities such as silica, also decreased from 0.701% to 0.395%, confirming the removal of such contaminants through purification (Prakash et al., 2019).

Extractive values are used to determine and measure the chemical components that dissolve in certain solvents (Prakash et al., 2019). In the unpurified pulp, the water- and alcohol-soluble extractive values were 4.37% and 3.965%, respectively, which dropped to 2.26% and 2.415% after purification. This overall decline reflects a reduction in chemical constituents, likely due to the effects of heat processing. Prior studies have also reported that such processing leads to the breakdown of active components. The pulp primarily contains cucurbitane-type triterpene glycosides, along with triterpene saponins, phenolic acids, and flavonoids. The observed

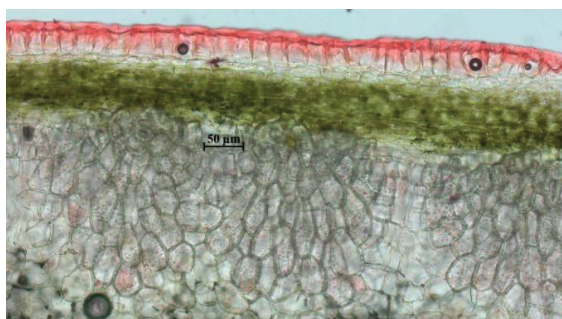
(a1) Tannin



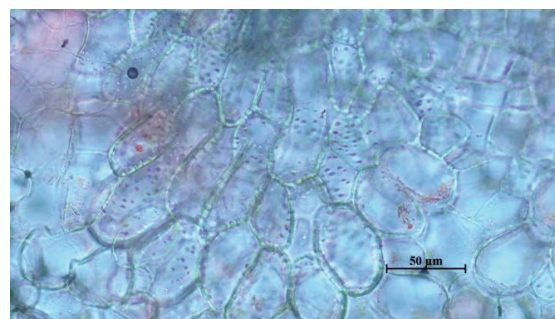
(a2) Tannin



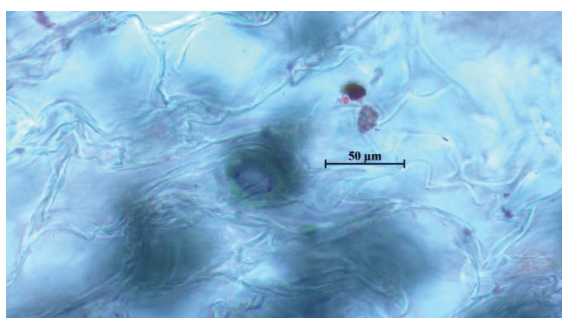
(b1) Cutin



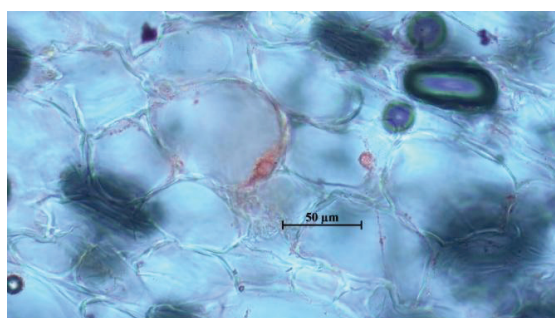
(c1) Oil globules



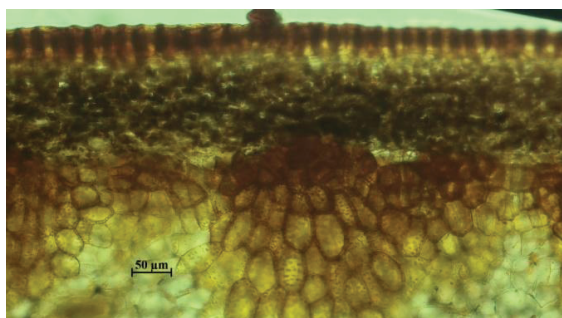
(c2) Oil globules



(c3) Oil globules



(d1) Alkaloid



(d2) Alkaloid

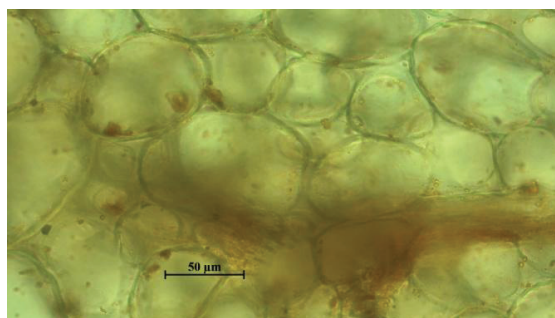
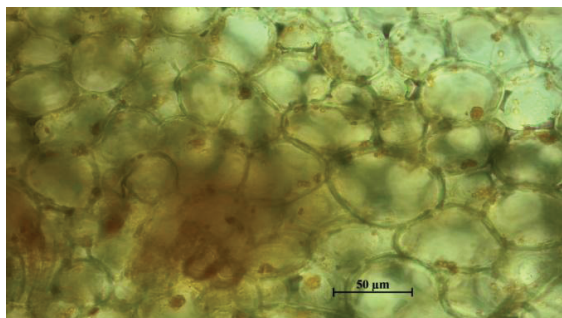


Figure 2. Histochemical analysis of *Citrullus colocynthis* (L.) pulp before purification (a to f).

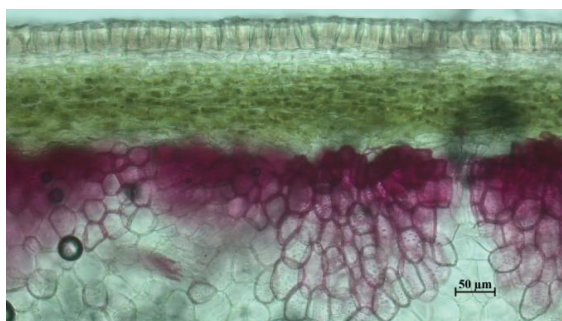
decrease in extractive values suggests that these glycosides may be converted into their corresponding genins during purification (Darwish et al., 2021). The histochemical analysis of *C. colocynthis* (L.) pulp before and after purification has revealed

significant alterations in its phytochemical composition of cells and tissues. One of the most notable findings was the presence of cutin on the epicarp walls in the unpurified pulp, which was absent in the purified pulp. Cutin, a protective

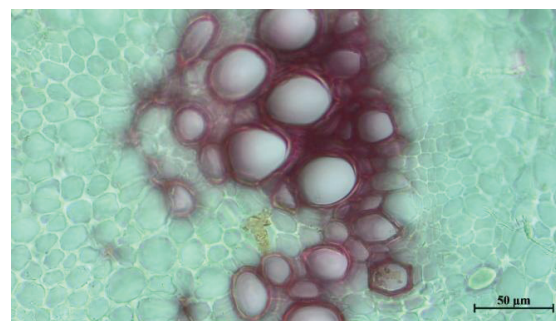
(d3) Alkaloid



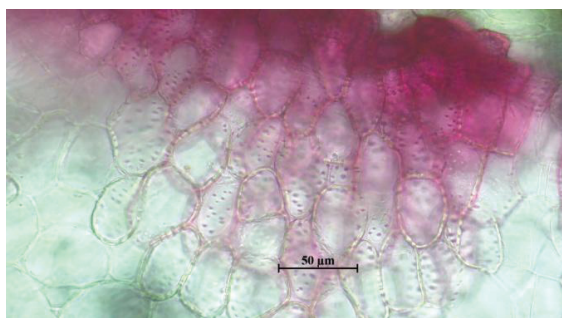
(d4) Alkaloid



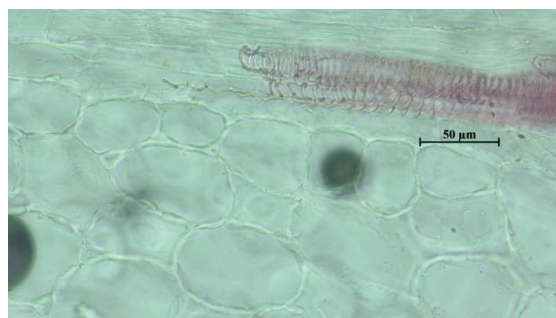
(e1) Lignin



(e2) Lignin



(f1) Mucilage



(f2) Mucilage

**Figure 2.** Continued.

Table 5

Histochemical analysis of *Citrullus colocynthis* (L.) after purification.

Epicarp region	-
Mesocarp region	1. Alkaloids, phenolic compounds, mucilage, and oil globules were detected in the mesocarp region of the fruit. 2. Lignin was present in the xylem of vascular strands and absent in stone cells. 3. Starch grains and resins were absent in the fruit.

waxy layer, serves as an effective barrier against water loss through transpiration, particularly in desert plants like *C. colocynthis* (Ladaniya, 2008). Therefore, this cutin deficiency after purification led to an increased proportion of hydrophilic constituents, contributing to a more succulent nature (Isacson et al., 2009).

Tannins, which were found in the mesocarp of the unpurified pulp but not in the purified pulp, are polyphenolic compounds known for their astringent properties. While tannins offer a range of beneficial effects, such as antioxidant and antimicrobial properties, they are also associated with potential antinutritional effects. Tannins can inhibit the bioavailability of essential nutrients, such as vitamins and minerals, and interfere with enzymatic activity. The absence of tannins in the purified pulp potentially improves the bioavailability of vitamins such as B12 and D (Sharma et al., 2019). Lignin, an essential component of the plants vascular system, was detected in the xylem strands and stone cells of the unpurified pulp. However, in the purified pulp, lignin was only found in the xylem of vascular strands, with no presence in the stone cells. Lignin is crucial for water conduction and structural support within the plant, and its selective removal from stone cells may imply changes in the mechanical properties and succulence of the pulp (Encyclopedia Britannica, 2025).

Both the unpurified and purified pulp contained mucilage, which was identified in the phloem cells and mesocarp parenchyma. Mucilage plays a vital role in water retention, seed dispersal, and potentially in the production of nanocarriers for pharmaceutical applications. Its persistence in both unpurified and purified pulp suggests that mucilage remains unaffected by the purification process, maintaining its functional properties in both forms of the pulp (Tosif et al., 2021). The presence of alkaloids in both the epicarp and mesocarp cells of the unpurified and purified pulp highlights the presence of bioactive compounds that may have pharmacological or toxic effects. Alkaloids, known for their diverse range of biological activities, could either contribute to the fruits medicinal properties or pose toxicity risks (Zheng et al., 2020).

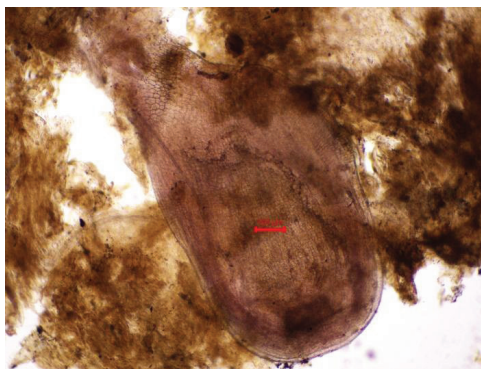
Oil globules, detected in the parenchyma cells of both unpurified and purified pulp, indicate the storage of lipids, which are important for energy provision and may also contribute to the fruits potential therapeutic properties. The preservation of oil globules in the purified pulp suggests that lipid content is not significantly altered during processing, which could be beneficial for maintaining the nutritional value of the fruit (Kalebar et al., 2019).

Phenolic compounds, which were detected only in the purified pulp, are known for their antioxidant properties. The increase in phenolic content following purification enhances the potential health benefits of *C. colocynthis* pulp, providing an additional mechanism for disease prevention and health promotion. The purification process enhances phenolic compounds, indicating antioxidant capacity of the pulp (Minatel et al., 2017). Starch grains, absent in the purified pulp, suggest that starch degradation occurred during the purification process, leading to a softening of the pulp. This change in texture could be due to the breakdown of starch into simpler sugars, making the fruit more easily digestible and possibly altering its glycemic index (Xiao et al., 2018). Resins, which are metabolic by-products of plant tissues, consist of a combination of volatile and nonvolatile terpenoid and/or phenolic secondary compounds, along with fatty substances that naturally exude from plants (McCreath and Clement, 2024). Cucurbitacins, a group of triterpenoids, are the main active compounds found in *C. colocynthis* (L.), known for their bitter taste and potential toxicity (Barton et al., 1999; Cheng et al., 2023). The absence of resins in the purified pulp suggests that these compounds, which may have influenced the potential medicinal properties, either eliminated or broken down during processing. The removal or degradation of potentially harmful compounds like tannins, cutin, and resins, along with the enhancement of bioactive components such as phenolics, could make the purified pulp a more beneficial and less toxic option for consumption. The preservation of beneficial compounds like mucilage, alkaloids, oil globules, and lignin in specific tissue types suggests that the purification process selectively alters the fruits chemical composition without completely removing its functional components. These findings illuminate the potential for using purification techniques to improve the therapeutic potential of *C. colocynthis* (L.) pulp.

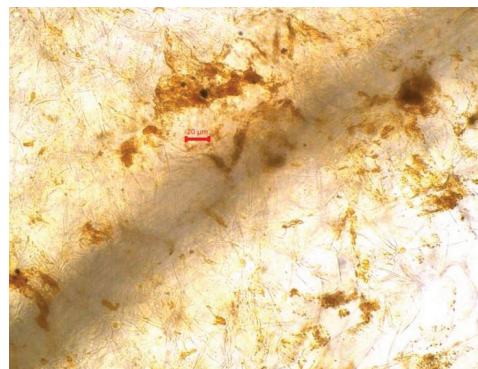
4. CONCLUSION

The histochemical localization of various phytochemicals in the pulp of *C. colocynthis* (L.) before and after purification reveals notable alterations in its physical and chemical characteristics, enhancing its usability and therapeutic value

(a) Mucilage



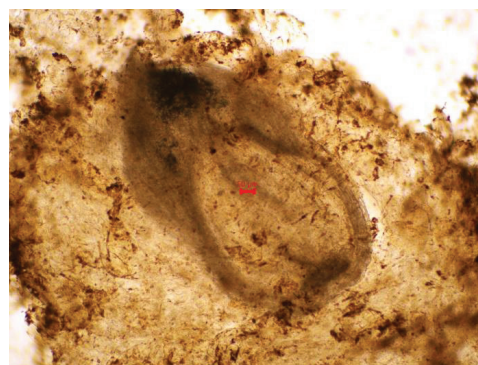
(b1) Phenolic compound



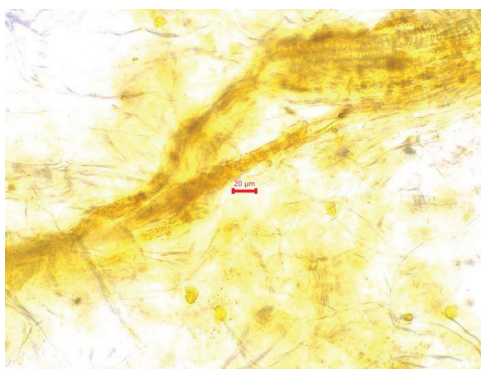
(b2) Phenolic compound



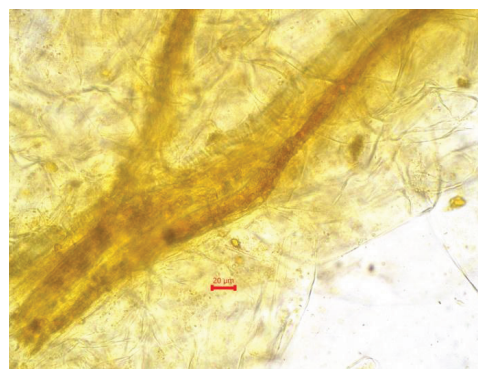
(b3) Phenolic compound



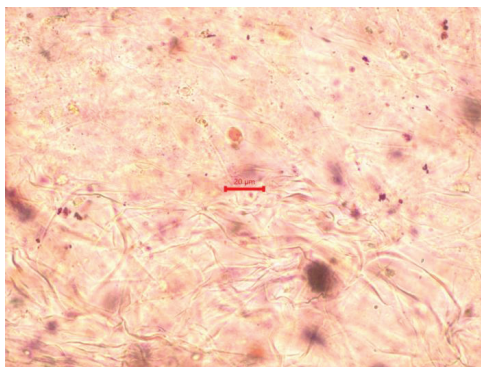
(c1) Alkaloids



(c2) Alkaloids



(d) Oil globules



(r) Lignin in vascular strands



Figure 3. Histochemical analysis of *Citrullus colocynthis* (L.) pulp after purification (a to e).

by minimizing toxic effects. The comprehensive organoleptic, physicochemical, and histochemical analyses aid in precise identification and standardization of the fruit, forming a basis for future pharmacological investigations. This preliminary assessment documents the transformations occurring through purification, offering valuable insights into its pharmacognostic features. The distribution of phytochemicals within the fruit tissues supports its traditional application in treating various conditions, underscoring the significance of *C. colocynthis* (L.) in both Siddha medicine and contemporary pharmacognosy.

ACKNOWLEDGMENTS

I sincerely acknowledge the Department of Pharmacognosy, Siddha Central Research Institute, Arumbakkam, and the faculty of the Department of Nanju Maruthuvam, National Institute of Siddha, for their invaluable technical support in conducting this research.

AUTHOR CONTRIBUTIONS

Conceptualization: KR; Medicine preparation: KR and AB; Data collection and compilation: KR, AB and RM; Manuscript writing: KR, AB, RM, RS, and MV; Proofreading and editing: KR, AB, RM, KA, RS, and MV.

CONFLICTS OF INTEREST

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

FUNDING

The author(s) received no financial support for the research, authorship, and/or publication of this article.

ORCID

Karpagambal Ramamoorthy	0009-0006-8481-1186
Abarna Balasubramani	0009-0005-1157-7205
Raghavi Marimuthu	0009-0006-5537-1310
Kalaiarasi Angappan	0009-0008-1925-6651
Suresh Ramasamy	0009-0004-9403-0629
Manjari Venkatraman	0000-0002-6946-5610

REFERENCES

- Alamgir, A.N.M., 2017. Introduction. In *Therapeutic use of medicinal plants and their extracts: Volume 1. Progress in Drug Research* (Vol. 73). Cham: Springer. https://doi.org/10.1007/978-3-319-63862-1_1
- Allen, L.V., 2003. pH and solubility, stability, and absorption. *International Journal of Pharmaceutical Compounding*.
- Badria, F., Aboelmaaty, W., 2019. Plant histochemistry: A versatile and indispensable tool in localization of gene expression, enzymes, cytokines, secondary metabolites and detection of plant infection and pollution. *Acta Scientifica Pharmaceutical Sciences*. 3(7), 88–100.
- Barton, D., Nakanishi, K., Meth-Cohn, O., 1999. *Comprehensive natural products chemistry* (pp. 263–375). Pergamon. ISBN 978-0-08-091283-7
- Cheng, X., Qin, M., Chen, R., Jia, Y., Zhu, Q., Chen, G., Wang, A., Ling, B., Rong, W., 2023. *Citrullus colocynthis* (L.) Schrad.: A promising pharmaceutical resource for multiple diseases. *Molecules*. 28(17), 6221.
- Darwish, R.S., Abdulmunem, O.A., Khairy, A., Ghareeb, D.A., Yassin, A.M., Abdulmalek, S.A., Shawky, E., 2021. Comparative metabolomics reveals the cytotoxic and anti-inflammatory discriminatory chemical markers of raw and roasted colocynth fruit (*Citrullus colocynthis* L.). *RSC Advances*. 11(59), 37049–37062. <https://doi.org/10.1039/d1ra07751a>
- Espinosa, J., Lucerna, A., Bassett, R., 2022. A bitter experience—A bitter apple (*Citrullus colocynthis* (L.)) ingestion in a child. *American Journal of Emergency Medicine*. 60, 225.e5–225.e6. <https://doi.org/10.1016/j.ajem.2022.07.011>
- Fahn, A., 1980. *Plant anatomy* (3rd ed.). Oxford: Pergamon Press.
- Hussain, A.I., Rathore, H.A., Sattar, M.Z.A., Chatha, S.A.S., Sarker, S.D., Gilani, A.H., 2014. *Citrullus colocynthis* (L.) Schrad (bitter apple fruit): A review of its phytochemistry, pharmacology, traditional uses, and nutritional potential. *Journal of Ethnopharmacology*. 155(1), 54–66. <https://doi.org/10.1016/j.jep.2014.06.011>
- Ibrahim, A., Alghannam, A., Eissa, A., Firtha, F., Kaszab, T., Kovacs, Z., Helyes, L., 2021. Preliminary study for inspecting moisture content, dry matter content, and firmness parameters of two date cultivars using an NIR hyperspectral imaging system. *Frontiers in Bioengineering and Biotechnology*. 9, 720630. <https://doi.org/10.3389/fbioe.2021.720630>
- Isaacson, T., Kosma, D.K., Matas, A.J., Buda, G.J., He, Y., Yu, B., Pravitasari, A., Batteas, J.D., Stark, R.E., Jenks, M.A., Rose, J.K.C., 2009. Cutin deficiency in the tomato fruit cuticle consistently affects resistance to microbial infection and biomechanical properties, but not transpirational water loss. *The Plant Journal*. 60(2), 363–377. <https://doi.org/10.1111/j.1365-313X.2009.03969.x>
- Juliet, L., Karthik, S., Reena, V.L., Aarthi, V., Kumar, K.N.S., Parameswaran, S., Kanakavalli, K., Hano, C., 2021. Traditional methods of purification (detoxification process) for Schedule E poisonous drugs. *Indian Journal of Traditional Knowledge*. 20(3), 740–748.
- Kalebar, V.U., Hoskeri, J.H., Hiremath, S.V., Hiremath, M.B., 2019. Pharmacognostical and phytochemical analysis of *Solanum macranthum* (Dunal) fruits. Volume 8. 284–290.
- Khandelwal, K.R., 2008. *Practical pharmacognosy* (19th ed.). Pune: Nirali Prakashan.

- Kumadoh, D., Amekyeh, H., Archer, M.-A., Kyene, M.O., Yeboah, G.N., Brew-Daniels, H., Adi-Dako, O., Osei-Asare, C., Adase, E., Appiah, A.A., 2024. Determination of consistency in pH of some commercial herbal formulations in Ghana. *Journal of Herbal Medicine*. 45, 100876. <https://doi.org/10.1016/j.hermed.2024.100876>
- Ladaniya, M.S., 2008. Fruit biochemistry. In *Citrus fruit* (pp. 125–190). San Diego: Academic Press. <https://doi.org/10.1016/B978-012374130-1.50008-5>
- Lohar, D., 2008. *Quality control manual for Ayurvedic, Siddha and Unani medicines*. New Delhi: Ministry of AYUSH, Government of India.
- McCreath, S.B., Clement, Y.N., 2024. *Pharmacognosy* (2nd ed., pp. v—xvi). Academic Press. ISBN 978-0-443-18657-8
- Minatel, I.O., Borges, C.V., Ferreira, M.I., Gomez, H.A.G., Chen, C-Y.O., Lima, G.P.P., 2017. Phenolic compounds: Functional properties, impact of processing and bioavailability. In *Phenolic Compounds — Biological Activity*. InTech.
- Murugesamudhaliyaar, K.S., 2013. *Materia Medica Gunapadam* (1st Part, 9th ed., pp. 61—62). Chennai: Indian Medicine and Homeopathy Department.
- Pharmacopeial Laboratory for Indian Medicine (PLIM). (n.d.). *Guideline for standardization and evaluation of Indian medicine which include drugs of Ayurveda, Unani and Siddha systems*. New Delhi: Department of Ayush, Ministry of Health & Family Welfare, Government of India.
- Prakash, A., Janmeda, P., Pathak, P., Bhatt, S., Sharma, V., 2019. Development and standardization of quality control parameters of different parts of *Trianthema portulacastrum* L. *SN Applied Sciences*. 1, 1108. <https://doi.org/10.1007/s42452-019-1074-3>
- Rahimi, R., Amin, G., Ardekani, M.R., 2012. A review on *Citrullus colocynthis* (L.) Schrad.: From traditional Iranian medicine to modern phytotherapy. *Journal of Alternative and Complementary Medicine*. 18(7), 551–554. <https://doi.org/10.1089/acm.2011.0297>
- Rao, V., Poonia, A., 2023. *Citrullus colocynthis* (L.) (bitter apple): Bioactive compounds, nutritional profile, nutraceutical properties and potential food applications: A review. *Food Production, Processing and Nutrition*. 5(4). <https://doi.org/10.1186/s43014-022-00118-9>
- Sharma, K., Kumar, V., Kaur, J., Tanwar, B., Goyal, A., Sharma, R., Gat, Y., Kumar, A., 2019. Health effects, sources, utilization and safety of tannins: A critical review. *Toxin Reviews*. 40, 1–13. <https://doi.org/10.1080/15569543.2019.1662813>
- The Editors of Encyclopaedia Britannica. (2025, February 21). Lignin. *Encyclopedia Britannica*. <https://www.britannica.com/science/lignin>
- Tosif, M.M., Najda, A., Bains, A., Kaushik, R., Dhull, S.B., Chawla, P., Walasek-Janusz, M., 2021. A comprehensive review on plant-derived mucilage: Characterization, functional properties, applications, and its utilization for nanocarrier fabrication. *Polymers*. 13(7), 1066. <https://doi.org/10.3390/polym13071066>
- Wallis, T.E., 1965. *Analytical microscopy: Its aims and methods in relation to foods, water, spices and drugs* (3rd ed.). Boston: Little, Brown and Company.
- Xiao, Y.Y., Kuang, J.F., Qi, X.N., Ye, Y-J., Wu, Z-X., Chen, J-Y., Lu, W-J., 2018. A comprehensive investigation of starch degradation process and identification of a transcriptional activator MabHLH6 during banana fruit ripening. *Plant Biotechnology Journal*. 16(1), 151–164. <https://doi.org/10.1111/pbi.12756>
- Zheng, M.S., Liu, Y.S., Yuan, T., Liu, L-Y., Li, Z-Y., Huang, X-L., 2020. Research progress on chemical constituents of *Citrullus colocynthis* (L.) and their pharmacological effects. *Zhongguo Zhong Yao Za Zhi*. 45(4), 816–824. <https://doi.org/10.19540/j.cnki.cjcmm.20191104.201>