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Genus *Ceiba* Mill. in Brazil: A comprehensive review on its ethnopharmacology, phytochemistry and bioactivities

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ABSTRACT: Introduction: Neotropical genus *Ceiba* Mill. is known for having tall trees, trunks with a robust part and fruits of economic interest. In Brazil, there are eleven native species, from those five are endemic. It is used in folk medicine to treat diabetes, inflammation, pain and diarrhea; however, most of the species have no scientific validation for such activities. This review aims to compile information about the genus *Ceiba* in Brazil, to update and allow an integrated understanding of its medicinal uses, chemical composition, and biological activities. Methodology: *Ceiba* species reported in Flora e Funga do Brasil were used for this literature review, using ScienceDirect, PubMed, and Google Scholar as articles databases. Results: In traditional use, the most cited species was *C. pentandra* (25 citations), the most used part was stem/bark (30 citations), preparation method was decoction (19 citations), and main administration route was oral ingestion (12 reports) for digestive system, skin and subcutaneous tissues diseases treatment. Genus chemical composition is wide, presenting metabolites such as proteins, sugars, fatty acids, tannins, flavonoids, and alkaloids. Bioactivities as antidiabetic, antioxidant, antitumor and antimicrobial were observed, especially for *C. pentandra*. Conclusion: Some species of *Ceiba* have extensive scientific literature, presenting several isolated compounds and bioactivities. On the other hand, some species, especially those that are endemics only in Brazil, do not have studies that evaluate their biological properties. Such knowledge is essential to confirm the medicinal potential cited in ethnobotanical reports, and the possibility of new compounds of biotechnological interest.

1. INTRODUCTION

Malvaceae Juss. family has a global distribution, especially in tropical and temperate regions, composed by almost 250 botanical genera (APG IV, 2017). Particularly, the neotropical genus *Ceiba* Mill. has 18 species that occur mainly in seasonally dry tropical forests (SDTFs) and tropical forests. The largest trees, about 50 m tall, are present in seasonally flooded várzea forests and the smallest, reaching 2 m, in rocky outcrops (Pezzini et al., 2021).

Ceiba are morphologically characterized by the presence of robust and aculeate trunks, leaves alternate compound-digitate, 5-7 leaflets with serrate margins, solitary flowers or in inflorescences, these being pentamerous and dichlamydeous, and fused staminal filaments that form a staminal tube (Figueiredo et al., 2020).

In Brazil, these species are distributed on all Brazilian geographic regions (North, South, Midwest, Northeast and Southeast) and in its six phytogeographic domains (Amazon, Caatinga, Cerrado, Atlantic Forest, Pampa and Wetlands).

There are 11 Brazilian native *Ceiba* species, six of which are also considered endemic, with highest prevalence in country's northeast region (REFLORA, 2022).

These genus species are used in landscaping and urbanism due to their large branches and brightly colored flowers, in addition to their economic and commercial applications, such as wood and vegetable oils and fibers production (as from *C. speciosa* and *C. pentandra* fruits) (Tripathi et al., 2019). Medicinal application of these species is also observed, in traditional medicine it is used for treating diabetes (Pereira Júnior et al., 2014), high blood pressure (Albuquerque et al., 2007; Almeida et al., 2005), inflammations (Sobrinho et al., 2021), chest and spine pains (Ribeiro et al., 2014; Silva et al., 2015), diarrhea (Ndenecho, 2009; Nwachukwu et al., 2008), and rheumatism (Agra et al., 2007, 2008; Sanz-Biset et al., 2009). Consequently, the number of scientific studies that seek to validate such knowledge has been increased in the last decade.

There are several literature reports about these species bioactive potential, examples are *C. pentandra*, as antidiabetic (Fofe

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et al., 2019), antimicrobial (Santos et al., 2021), antiviral (Dewi et al., 2019), and antitumor (Abouelela et al., 2018). Another that is frequently cited is *C. speciosa*, for its antidiabetic (Santos et al., 2020) and antioxidant properties (Braga et al., 2019).

Although there are a relevant number of publications about *Ceiba* spp., their restricted geographic distribution, reduce the scientific studies interest. Moreover, the absence of ethnopharmacological and/or ethnobotanical surveys restrains their information, especially for Brazilian endemic species. Thus, this review aims to compile the available information about *Ceiba* species to offer a scientific literature update and to help on understanding its ethnopharmacology, chemical composition, and biological activities in an integrated manner.

2. MATERIALS AND METHODS

For this integrative review design, Flora e Funga do Brasil database was used, due to its attribution on recording, grouping, and disseminating data regarding Brazil plant specimen's occurrence, and 11 *Ceiba* Mill. species were reported on this database in Brazil at the search period (<http://reflor.a.jbrj.gov.br>). Then, the review based on collection of evidences, especially for those with chemical, bioactivities, or ethnobotanical approaches.

Literature review was carried out between June 2020 and January 2023, through selecting articles in English, Portuguese, Spanish and French, indexed in databases ScienceDirect, PubMed, and Google Scholar, with the main keywords: *Ceiba crispiflora*, *Ceiba erianthos*, *Ceiba glaziovii*, *Ceiba jasminodora*, *Ceiba lupuna*, *Ceiba pentandra*, *Ceiba pubiflora*, *Ceiba rubriflora*, *Ceiba samauma*, *Ceiba speciosa* and *Ceiba ventricosa*, in addition to the synonyms *Chorisia crispiflora*, *Chorisia glaziovii*, *Chorisia pubiflora*, *Chorisia speciosa* and *Chorisia ventricosa*. In addition, the keywords ethnopharmacology, medicinal plants, biological activities, pharmacological potential were also used. The selection criteria were based on the presence of reports about botanical, phytochemical, or pharmacological activities of these 11 species. Studies regarding other *Ceiba* species, and non-pharmacological activities were removed.

3. MORPHOLOGY AND DISTRIBUTION IN BRAZIL

Ceiba Mill. taxonomic history presents divergences, because of species that currently integrate it have already composed other genera, such as *Chorisia* (APG IV, 2017). Morphological characteristics that differentiated the two genera are the lower staminal filaments arrangement in a corona-like structure, and the upper filaments fused to form the upper staminal tube. This last structure is absent in *Ceiba*, which has only the lower staminal tube, which divides into five free stamens at the top (Gibbs & Semir, 2003). However, after morphological analysis of pollen grains, a union between the two genera was proposed, and the classification of *Ceiba* is currently used for both genus (Gibbs et al., 1988).

Ceiba spp. trees most often have a height between 15 to 20 m, however *C. pentandra* can reach more than 50 m, being the tallest member of the group. Trunks are straight, robust, and

aculeate, and, in some species, such as *C. speciosa*, *C. glaziovii* and *C. pubiflora*, may have ventricose or “swollen” trunks, presenting a more robust area in stem middle portion. This particularity in its structure explains some common names given to these trees in South America, such as “barriguda” (in literal translation would be “paunchy”), in Brazil, and “palo borracho” (in literal translation would be “drunken tree”) in Peru, Bolivia and Argentina (Gómez-Maqueo & Gamboa-Debuen, 2022; Pezzini et al., 2021). Trunk also culminates in a dense canopy, with alternate compound-digitated leaves, long petioles, and leaflets (in number of 5 to 7) with serrated margins. Flowers can be solitary or organized in fasciculate inflorescences of few flowers, presenting deciduous bracts (Gibbs & Semir, 2003).

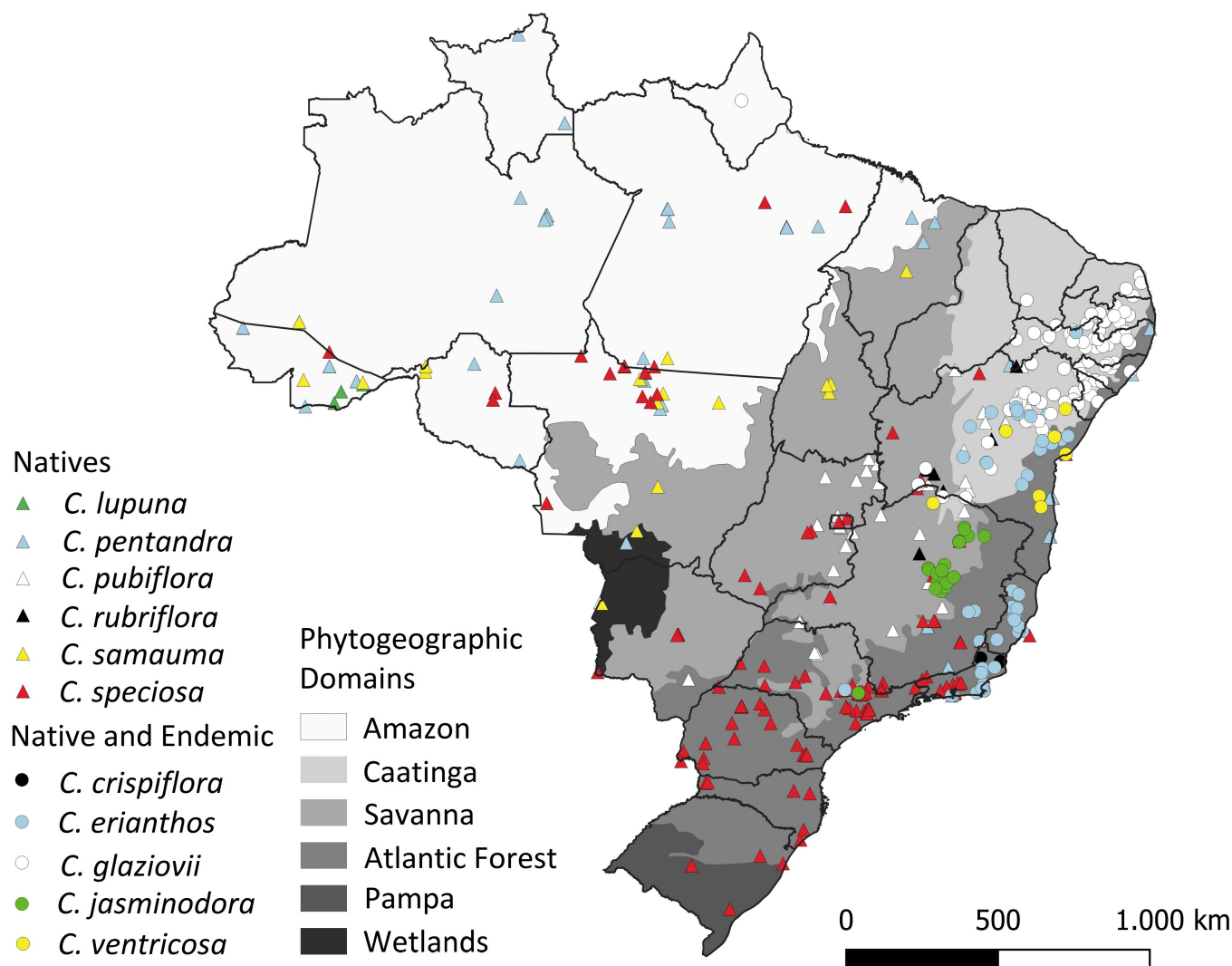
Fruits have an ellipsoidal capsule shape and endocarp develops into a fine white cotton fiber (named kapok), which is why they are known as “silk cotton tree” and “kapok tree”. This fiber is capable of protecting the seeds and assist in wind dispersion. This genus representatives show great morphological similarity but can be distinguished by differences in staminal filaments, in which these can be fused, forming a tube around the ovary (called staminal tube). Staminal filaments also can divide into five free filaments in some species, or remain fused just above the stem appendages, forming the upper staminal tube (Gibbs & Semir, 2003).

Among the cataloged species, eleven are found in Brazil and six of them are native. The ones found in Brazil are: *Ceiba lupuna* P.E.Gibbs & Semir, *Ceiba pentandra* (L.) Gaertn., *Ceiba pubiflora* (A.St.-Hil.) K.Schum., *Ceiba rubriflora* Carv.-Sobr. & L.P.Queiroz, *Ceiba samauma* (Mart.) K. Schum. and *Ceiba speciosa* (A.St.-Hil.) Ravenna, and five others species are native and endemic to the Brazilian territory: *Ceiba glaziovii* (Kuntze) K. Schum., *Ceiba crispiflora* (Kunth) Ravenna, *Ceiba erianthos* (Cav.) K.Schum., *Ceiba jasminodora* (A.St.-Hil.) K. Schum. and *Ceiba ventricosa* (Nees & Mart.) Ravenna (REFLORA, 2022).

These species are distributed on all Brazilian geographic regions (North, South, Midwest, Northeast, and Southeast) and in its six phytogeographic domains (Amazon, Caatinga, Cerrado, Atlantic Forest, Pampa, and Pantanal), as can be seen in Figure 1 (SiBB, 2022). They occur, mainly, in rocky outcrops, rocky fields, limestone fields and seasonally dry tropical forests (STDFs) (Figueiredo et al., 2020; REFLORA, 2022). Although most of the species are present in STDFs, *C. jasminodora* is found only in rocky outcrops, and other species are also present in humid habitats, such as *C. speciosa* and *C. samauma*, present in several South American countries. *C. pentandra* has a pantropical distribution and *C. lupuna* is restricted to rainforests (Gibbs & Semir, 2003; Pezzini et al., 2021).

4. MEDICINAL TRADICIONAL USE

Medicinal knowledge is part of the characteristic habits and customs of traditional communities and is used for the prevention and treatment of diseases. Traditional medicinal knowledge is transmitted orally or through written narratives,



Data for this map construction was obtained from Instituto Brasileiro de Geografia e Estatística (IBGE), Interface Integrada para Internet de Ferramentas de Geoprocessamento (i3GEO) created by Ministério do Meio Ambiente (MMA), and Sistema de Informação sobre a Biodiversidade Brasileira (SiBBR). Map was constructed using QGIS software.

Figure 1. Geographic distribution of *Ceiba* genus species in Brazil.

being fundamental in maintaining the physical, emotional, and spiritual integrity of the community (García-Flores et al., 2019).

This knowledge is extremely relevant, about 80 % of the world population uses herbal medicines and, in tropical countries, two thirds of the plant species used are obtained directly from nature (Albuquerque et al., 2011). In traditional rural communities, these practices are even more evident, as access to health services can be restricted (Saraiva et al., 2015).

From the traditional knowledge to prospecting new drugs, the first one cannot be totally discarded since it helps in the search for compounds with the most diverse bioactivities, which has been a successful tool for new medicinal alternatives

development over the last few decades (Ribeiro et al., 2017; Saraiva et al., 2015).

Several ethnopharmacological uses are reported for *Ceiba* species in Brazil and in other tropical countries of the world, as can be seen in Table 2. 42 reports were found, among them, *C. pentandra* was the most cited (25) followed by *C. glaziovii* (11), *C. speciosa* (3) and *C. pubiflora*, *C. samauma* and *C. ventricosa*, with only 1 report each. *C. crispiflora*, *C. erianthos*, *C. jasminodora*, *C. lupuna* and *C. rubriflora* were not mentioned in the consulted literature as used for traditional medicinal purposes.

Interestingly, *C. glaziovii* is found exclusively in Caatinga phytogeographic domain, in Brazil (Gibbs & Semir, 2003), and

although it has a restricted distribution, it presents a significant number of ethnobotanical studies, which places this species in prominence. Although there is considerable literature on its ethnopharmacological uses, there are no experimental studies that confirm the medicinal properties of this species (Saraiva et al., 2015).

4.1. Modes of preparation

The use of various plant elements was found in the consulted literature, totaling 57 reports contained in 42 studies. The bark/stem (bark, inner bark, stem and trunk) was the most cited element (30), followed by leaves (8), roots (7), flowers (6), fruits (3), xylem/heartwood (2) and resin (1). The higher recurrence of the popular use of elements related to the bark/stem can be observed in other ethnobotanical approaches. This may be related to the perenniality of these elements in the plant during seasonal changes, easy collection and storage, higher concentration of bioactive substances, and preservation of therapeutic properties for long periods of time (Pereira Júnior et al., 2014; Saraiva et al., 2015; Sobrinho et al., 2021).

Regarding the methods of medicinal plants preparations, 42 citations were found in the 31 articles consulted, distributed in five different ways: decoction, infusion, maceration, syrup, and percolation. Decoction and infusion have the highest number of reports, with 19 and 6 citations, respectively. They consist of the extraction of water-soluble constituents; in the decoction, the vegetable raw material is boiled in an open extractor to obtain thermostable compounds, and in the infusion, the vegetable material macerated in cold or boiling water is used to obtain the phytochemicals (Manousi et al., 2019).

The least mentioned methods of preparation were maceration, syrup, and percolation, with 3, 2 and 1 citations, respectively. In maceration, the material is ground and placed in a container, covered by the solvent, and kept for, at least, three days. Percolation is a more sophisticated technique that uses a dedicated device, in which the plant raw material is placed inside it as an immobile bed, and extraction occurs by passing the solvent in a continuous flow (Abubakar & Haque, 2020). Syrups are prepared with sugar or honey and are generally used to treat diseases in children, being popularly known as “lambedor” in Brazil (Agra et al., 2008).

4.2. Treated diseases and administrations forms

In the consulted literature, there are 17 reports of forms of medicinal administration of *Ceiba* species. The most cited is the oral intake (12 reports), with the use of teas, juices, and syrups, followed by topical use (3 reports), washing the affected areas (2), and mouthwash and contact with the cervix, with 1 citation each.

Several therapeutic indications are cited for the medicinal use of *Ceiba* species (64 occurrences were reported) in 42 articles, categorized according to the classification proposed by Albuquerque et al. (2007) with modifications. Use indications for treatment Diseases of the digestive system and Diseases of the skin and subcutaneous tissues are the most

cited categories (8 reports each), as can be seen in Table 1. Followed by afflictions and pains not defined (7), Infectious and parasitic diseases and Injury, poisoning and other infirmities with external causes (6 each), Diseases of the respiratory system and endocrine, nutritional and metabolic diseases (5 each), Diseases of the musculoskeletal system and connective tissues and Pregnancy, childbirth and the puerperium (4 each), Diseases of the genitourinary system, Diseases of the circulatory system, and Diseases of the blood and blood-forming organs (3 each), and Neoplasias (2).

Table 1

Number of medicinal indications for *Ceiba* species in traditional use, grouped into categories.

Categories of therapeutic indications	Number of occurrences
Diseases of the digestive system	8
Diseases of the skin and subcutaneous tissues	8
Afflictions and pains not defined	7
Infectious and parasitic diseases	6
Injury, poisoning, and other infirmities with external causes	6
Diseases of the respiratory system	5
Endocrine, nutritional, and metabolic diseases	5
Diseases of the musculoskeletal system and connective tissues	4
Pregnancy, childbirth, and the puerperium	4
Diseases of the genitourinary system	3
Diseases of the circulatory system	3
Diseases of the blood and blood-forming organs	3
Neoplasias	2

5. PHYTOCHEMICAL COMPOSITION

Plants produce several chemical constituents that play critical roles in these sessile and multicellular organisms, making them capable of inhabiting various environmental niches. These compounds can be from primary metabolism, such as amino acids, carbohydrates, nucleotides, and fatty acids, or from secondary metabolism (also called specialized), as phenols, terpenes, phenolic acids, and alkaloids (Maeda, 2019).

Secondary metabolites are essential for interaction with the environment, although their presence may diverge for each species. Biosynthesis of secondary metabolites varies according to the environmental conditions on the plant imposed, such as climatic aspects, seasonality, temperature, and humidity, generating specific responses on the phenology (Isah, 2019). *Ceiba* spp. native of Brazil have a great diversity of primary and secondary metabolites, which are obtained from different parts of these plants.

Table 2
Uses of *Ceiba* genus species in traditional medicine.

Scientific name	Vernacular name	Part used	Therapeutic indication	Mode of preparation	Form of use	Country of the study	Reference
<i>Ceiba</i> . sp.	Barriguda	Bark	Leprosy, cancer, and hemorrhoids	Maceration	NR	Brazil	Ribeiro et al. (2017)
	Barriguda de espinho	Inner bark	Hearth and high blood pressure	NR	NR	Brazil	Almeida et al. (2005)
	Barriguda	Leaves	Rheumatism and edemas	Decoction (handful/ liter) in water	Wash the affected area until the symptoms disappear	Brazil	Agra et al. (2007)
	Barriguda, Barriguda de espinho	Bast, flowers and leaves	Cardiac problems, hypertension, influenza, and bronchitis	NR	NR	Brazil	Albuquerque et al. (2007)
<i>Ceiba glaziovii</i> (Kuntze) K.Schum.	Barriguda-de-espinho	Leaves and stem bark	Rheumatism and edemas	Decoction (handful/ liter) in water	Wash the affected area until the symptoms disappear	Brazil	Agra et al. (2008)
	Barriguda	Stem bark	Anemia	Leave soaking	Oral intake	Brazil	Cartaxo et al. (2010)
	Barriguda	Stem bark	Diabetes	Decoction	NR	Brazil	Pereira Júnior et al. (2014)
	Barriguda	Stem bark	(Spine) pain	Sirup	Oral intake	Brazil	Ribeiro et al. (2014)
	Barriguda	Inner bark	Prostate inflammation	Immersion in water	Oral intake	Brazil	Saraiva et al. (2015)
	Barriguda	Bast	Spine pain	Tea	Oral intake	Brazil	Silva et al. (2015)
	Barriguda	Bark	Inflammation	Tea	Oral intake	Brazil	Sobrinho et al. (2021)
<i>Ceiba pentandra</i> (L.) Gaertn.	NR	Bark, branch xylem and root	Inflammation and analgesia	Decoction	NR	Taiwan	Kan (1975)
	Gotton Harrery	Branches	Emetic	Decoction	NR	Sudan	El-Kamali and El-Khalifa (1999)
	Douma, Dum, Bouma, Djam	Gum (Stem bark and leaves)	Abortion	Infusion	Putting the gum in contact with the cervix	Cameroon	Noumi and Tchakonang (2001)
	pixtyiñ, pix ti'ink	NR	Dermatological properties	NR	NR	Mexico	Leonti et al. (2003)

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Table 2 continued

Gon	Fruit	Diarrhoea, inflammation and malária	NR	NR	Vietnam	Nguyen et al. (2004)
Gounga	Bark	Mouth sores and gingivitis	Decoction	Use as a mouthwash until recovery	Burkina Faso	Tapsoba and Deschamps (2006)
Banan	Stem bark, root bark and leaves	Antitrypanosomal	Decoction	NR	Mali	Bizimana et al. (2006)
Mfuma	Leaves	Antidrepanocytary	Percolation	NR	Congo	Mpiana et al. (2007)
NR	Aerial parts	Chest pain	NR	NR	Nigeria	Kubmarawa et al. (2007)
Mbouma	Trunk bark	Stomachache	Decoction	Oral ingestion, 3x a day for 3 days.	Congo	Ibara et al. (2007)
Rimii	Stem bark	Fibrosis	NR	NR	Nigeria	Abubakar et al. (2007)
Rimi, apku, araba	Bark	Infections, bowel complains and diarrhoea	NR	NR	Nigeria	Nwachukwu et al. (2008)
Kaboka tree, akpu, ogwu, rimi, ogungun, araba	Stem bark	Threatened miscarriage treatment	Decoction	Oral ingestion, 3x a day	Nigeria	Akaneme (2008)
Kapas, randu	Leaves	Dermatitis	Decoction	Topical use	Indonesia	Roosita et al. (2008)
NR	Leaves, bark and roots	Chest pain, purgative, heart palpitations, diabetes, diarrhea, headaches, and rheumatism.	NR	NR	Cameroon	Ndenecho (2009)
cài hão	Roots	Fracture	Mashed material	Cataplasm	China	Zheng and Xing (2009)
Safed samal	Roots	Antigonorrhoeal	NR	NR	India	Shivhare et al. (2011)
Kumaka	Bark	Antileishmanial	Decoction	Juice and/or poultice	France	Odonne et al. (2011)
NR	Stem bark	Trating old wounds	Decoction	NR	Ivory Coast	Kone et al. (2012)
NR	NR	Antidrepanocytary	Extract (type not specified)	NR	India	Mohanty et al. (2012)

Continued on next page

Table 2 continued

	NR	Leaves	Ribs pains	Decoction prepared with feet of fowl	Oral ingestion, 3x a day	Ghana	Wodah and Asase (2012)
	NR	Trunk bark and branches	Antithrombotic / sickle cell disease	Decoction	NR	Congo	Nsimba et al. (2013)
	Araba, akpu-owu, rimi	Leaves	Antidiabetic	Decoction	Oral ingestion, 3x a day	Nigeria	Oyedemi et al. (2017)
	NR	NR	Dermatological properties and women's health issues	NR	NR	Mexico	García-Flores et al. (2019)
	Akpuogwu, cotton tree	Stem bark	Uterotonic	NR	NR	Nigeria	Ijioma et al. (2020)
<i>Ceiba pubiflora</i> (A.St.-Hil.) K.Schum.	Cuconeja	NR	Wounds	NR	NR	Paraguay	Schmeda-Hirschmann (1993)
<i>Ceiba samauma</i> (Mart.) K.Schum.	Wimba lupuna	Root and trunk bark	Rheumatism	Decocted or macerated in fresh water	NR	Peru	Sanz-Biset et al. (2009)
<i>Ceiba speciosa</i> (A.St.-Hil.) Ravenna	Algodão	NR	Medicinal and abortive	NR	NR	Brazil	Leitão et al. (2009)
	Paineira	Fruits, leaves, flowers, and bark	Diseases of the cardiovascular and respiratory system	Infusion, decoction and syrup	Oral intake	Brazil	Bolson et al. (2015)
	Silk tree, kapok, barriguda	Trunk bark	Reduction of serum cholesterol, triglycerides and glucose levels	Decoction	Oral intake	Brazil	Malheiros et al. (2017)
<i>Ceiba ventricosa</i> (Nees & Mart.) Ravenna	Barriguda, paineira	Resin, bark and wood	inguinal hernia	NR	NR	Brazil	Moraes (1881)

NR: Not reported.

5.1. *Ceiba pentandra*

C. pentandra has a vast scientific literature about its chemical composition around the world. Stem bark has phenols (Nwachukwu et al., 2008), tannins, catechins, mucilage (Kone et al., 2012), C-glycosides, reducing sugars, triterpenes (Bairwa et al., 2010), tannins, flavonoids, alkaloids, terpenoids, saponins (Syihabudin et al., 2018), resins, proteins and steroids (Akaneme, 2008). Aerial parts have saponins, tannins, alkaloids, essential oils (Kubmarawa et al., 2007), hydrocarbons and fatty acids (Abouelela et al., 2018), and the leaves of *C. pentandra* have carotenes (Smith et al., 1996), vitamins, proteins (Dickson et al., 2012), fatty acids, mono and polysaccharides, fiber (Herzog et al., 1993), alkaloids, phenols (Bhavani et al., 2016), saponins, flavonoids, terpenoids and steroids (Bhuvanawari et al., 2014).

Seeds were also investigated for their composition, having phenolic compounds, flavonoids, alkaloids, tannins (Kiran et al., 2015), fatty acids, proteins, fibers and phytosterols (Anwar et al., 2014). Essential oil obtained from stem bark present as major constituents β -caryophyllene (28.7%), β -elemene (18.5%), α -muurolene (7.8%), caryophyllene oxide (4.8%) and α -humulene (4.2%), and heartwood essential oil have α -eudesmol (21.1%), 2-ethoxyacetate (11.3%) and nonanal (7.3%) as main compounds (Alade et al., 2021). In addition to these compounds mentioned above, literature also report a great diversity of isolated phytochemicals from this species, as can be seen in Table 3. Despite the high number of studies that use *C. pentandra* as an object of investigation, no research produced in Brazil was found, even though this species has a wide distribution in the Brazilian territory (Figure 1).

5.2. *Ceiba speciosa*

Phytochemical analyzes of *C. speciosa* are reported around the world, using several parts of the plant, as bark, seeds, leaves, and fruits. Branches aqueous extract has a wide variety of phenols and flavonoids, such as quercetin, rutin, and gallic, caffeic and chlorogenic acids, and a total dosage of phenolic compounds of 117.4 ± 6.2 mg GAE/g (milligrams equivalent of gallic acid per gram of extract) (Dörr et al., 2019). In dosages to determine the quantity of flavonoids and polyphenolic compounds from trunk aqueous extract, values of 240 μ g of quercetin/g and 425 μ g of gallic acid/g were found, respectively (Malheiros et al., 2017). Also, in this study, the phytochemical profile of *C. speciosa* bark ethanolic extract was evaluated, with total flavonoids dosage (429 μ g of quercetin/g), total polyphenols (470 μ g of gallic acid/g), and identification by high performance liquid chromatography (HPLC) of various phenolic compounds, such as kaempferol, quercetin, rutin, caffeic, chlorogenic, gallic, and ellagic acids.

Leaves aqueous extract also has a low content of phenols (3.3 mg gallic acid/g extract) and flavonoids (7.7 mg quercetin/g extract) (Krishnaveni et al., 2013), and leaves ethanolic extract has triterpenes (β -amyryn), sterols (stigmasterol), organic acids (p-hydroxybenzoic and succinic acids) and glycosides (ver-

bascoside, β -sitosterol-3-O- β -d-glucopyranoside, astragalín, cinnaroside, tytyroside and rhiofolin) (Nasr et al., 2018).

Fruits, more specifically mesocarp and the exudate gum, have several carbohydrates, such as rhamnose, arabinose, xylose, mannose, glucose, and galactose (Beleski-Carneiro et al., 1999). Seed oil, submitted to gas chromatography followed by mass spectroscopy (GC-MS), presented a rich lipid profile, being found as major constituents: linoleic (28.22 %), palmitic (19.56 %), malvalic (16.15 %), sterculic (11.11 %), and dihydrosterculic acids (2.74 %) (Rosselli et al., 2020).

C. speciosa is also capable of producing essential oils, which, after being obtained from the leaves, were identified through GC-MS as major components: caryophyllene (32.26 %), bicyclogermacrene (9.88 %), humulene (9.26%), α -selinene (9.18 %), and β -elemene (8.06 %) (Kausar et al., 2020).

5.3. Other species

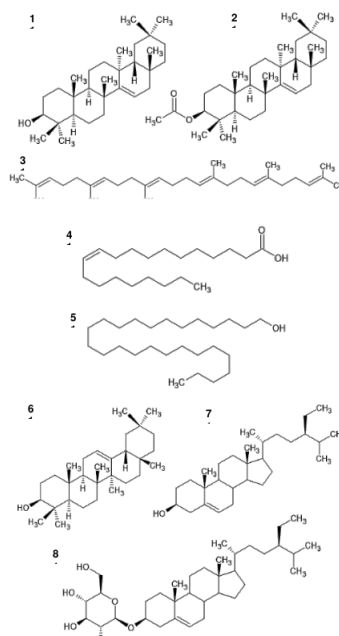
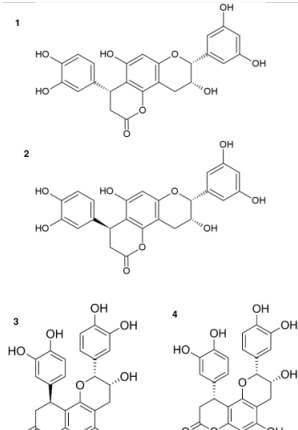
Other species of *Ceiba* genus that are native to Brazil had qualitative and quantitative analysis of its phytochemicals. In studies with the ethanolic extract of *C. glaziovii* bark, the presence of phenols, tannins, triterpenes and quinones was identified (Almeida et al., 2005). In bark and leaf hydroalcoholic extracts, catechin tannins, flavonols, phenols, flavones, xanthenes, alkaloids, anthocyanins, anthocyanidins, saponins and proteins were found (Leal et al., 2011). In studies with *Ceiba pubiflora* flowers hydroethanolic extract, a quantitative analysis of total phenols was performed, obtaining 7.26 ± 0.16 mg GAE/100g. In qualitative analysis, presence of chemical groups such a cyanogenic glycoside, reducing and non-reducing sugars, tannins, flavonoids, alkaloids, coumarins, saponins, organic acids and steroids were identified (Menezes-Filho et al., 2022).

6. BIOLOGICAL ACTIVITIES

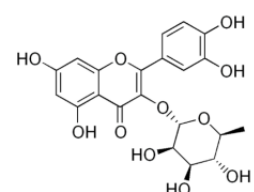
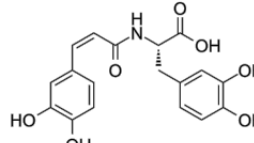
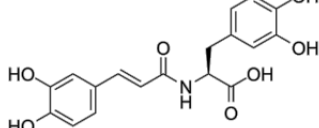
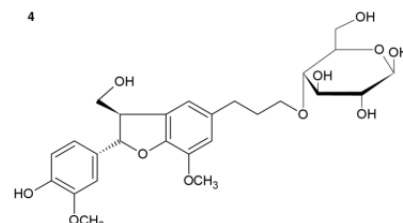
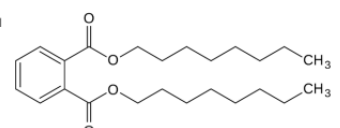
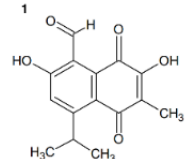
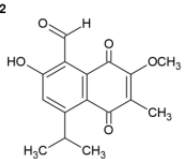
6.1. *Ceiba pentandra*

The main biological activity cited for *C. pentandra* is antidiabetic. Studies report its ability to inhibit carbohydrate digestive enzymes, a property that can help control blood glucose of patients, especially in reducing postprandial hyperglycemia. Stem bark methanolic and aqueous extracts showed inhibition of α -amylase with IC₅₀ values of 6.15 and 54.52 μ g/ml, and α -glucosidase of 76.61 and 86.49 μ g/ml, respectively (Nguelefack et al., 2020). Leaves methanolic extract showed weak inhibition of α -amylase (5.86 %), and high total content of phenolic compounds and flavonoids, in addition to potent free radical scavenging, and efficient reducing power (RP) as compared with other plant in this study, showing a low correlation between antioxidant activity and enzymatic inhibition (Oyedemi et al., 2017). Bark hydroalcoholic extract fractions show promising α -glucosidase inhibition activity, in which IC₅₀ values of 4.60, 8.55 and 5.61 μ g/mL for n-hexane, ethyl acetate and aqueous fractions, respectively, having approximate values to quercetin (6.04 μ g/mL), a flavonoid used as a standard (Syihabudin et al., 2018).

Table 3
Isolated compounds from *C. pentandra*.

Part of Plant	Plant Material	Compounds	Chemical Structure	Autor
Aerial parts (leaves, petioles, and young twigs)	Methanol/chloroform, aqueous, ethyl acetate and n-butanol partitions (Methanolic extract 80%)	3 β -Taraxerol(1), 3 β -Taraxerol acetate(2), All-trans-squalene(3), Oleic acid(4), 1-Hexacosanol(5), β -Amyrin(6), β -Sitosterol(7), β -Sitosterol-3-O- β -D-glucopyranoside(8)		Abouelela et al. (2018)
Aerial parts	Ethyl Acetate Extract	ceibapentains A (1) and B (2), cinchonains Ia (3) and Ib (4)		Abouelela, Orabi, Abdelhamid, Abdelkader, Madkor, et al. (2020)

Continued on next page

<i>Table 3 continued</i>				
Aerial parts	Ethyl Acetate fraction (Methanolic extract)	Quercitrin (1), cis-clovamide (2), trans-clovamide (3), and glochidioboside (4)	<p>1 </p> <p>2 </p> <p>3 </p> <p>4 </p>	Abouelela, Orabi, Abdelhamid, Abdelkader, Darwish, et al. (2020)
Leaves	Methanolic extract	Diocetyl phthalate (1)	<p>1 </p>	Ibrahim et al. (2012)
Heartwood	N-hexane, acetone and methanol extract	2,7-dihydroxy-8-formyl-5-isopropyl-3-methyl-1,4-naphthoquinone (1), 8-formyl-7-hydroxy-5-isopropyl-2-methoxy-3-methyl-1,4-naphthoquinone (2)	<p>1 </p> <p>2 </p>	Kishore et al. (2003)

Continued on next page

Table 3 continued				
Stem bark	Fractions of n-hexane, dichloromethane and ethyl acetate (Methanolic extract 80%)	5,5'-dihydroxy-7,3',4'-trimethoxyisoflavone (1) and 5,5'-dihydroxy-7,3',4'-trimethoxyisoflavone 5'-O- β -D-glucoside (2)		Ngounou et al. (2000)
Bark	Hydroalcoholic extract (70%)	5-Hydroxy-7,4',5'-trimethoxyisoflavone 3'-O- β -D-glucoside (1), 5,3'-Dihydroxy-7,4',5'-trimethoxyisoflavone (2)		Noreen et al. (1998)
Root bark	Hexane, chloroform, and methanol extract	8-formyl-7-hydroxy-5-isopropyl-2-methoxy-3-methyl-1,4-naphthoquinone (1), 2,7-dimethoxy-5-isopropyl-3-methyl-8,1-naphthalene carbolactone (2), 7-hydroxycadalene (3) and 2-hydroxy-5-isopropyl-7-methoxy-3-methyl-8,1-naphthalene carbolactone (4)		Rao et al. (1993)
Bark	Hydroalcoholic extract (80%)	5-hydroxy-7,4',5'-trimethoxyisoflavone 3'-O- α -L-arabinofuranosyl(1 \rightarrow 6)- β -D-glucopyranoside (1)		Ueda et al. (2002)

In vitro antihyperglycemic activity of bark decoction was also observed, in hyperglycemic milieu, there was a significant increase in glucose uptake by the liver (56.57 %) and skeletal muscle (94.19 %), and in hypoglycemic milieu, there was a reduction in glucose release by the liver (33.94 %) (Fofie et al., 2014). In *in vivo* evaluations, trunk bark aqueous and methanolic extracts of *C. pentandra* can improve peripheral glucose uptake, insulin resistance, lipid metabolism, and oxidative state. The methanolic extract (200 mg/Kg) caused a glycemic reduction of 62.4%, when administered to diabetic rats, in addition to improving oral glucose tolerance, and physiological changes caused by diabetes, such as cholesterol and triglycerides, body and pancreas weight, and antioxidant enzymes concentration such as glutathione and malondialdehyde (Fofie et al., 2019).

Bark decoction (150 mg/Kg) can reduce blood glucose by 33 % in oral treatment, and correct impaired glycaemia in rats with insulin resistance induced by dexamethasone, improving dyslipidemia and increasing levels of nitric oxide and antioxidant enzymes such as catalase and glutathione (Fofie et al., 2018). Ethyl acetate fraction of leaves ethanolic extract was able to reduce glycemia in rats with alloxan-induced diabetes, with high reduction observed to the treatment with 200 mg/Kg. In addition to reduction of serum levels of kidney (urea and creatinine) and liver (ALP, ALT and AST) injury markers, compared with untreated group (Lami et al., 2015).

Antifungal and antiviral potential of *C. pentandra* derivatives were also investigated, as well as the antimicrobial activity. Leaves ethanolic extract acts on virulence factors of *Pseudomonas aeruginosa*, showing significant inhibition of biofilm formation (1.00 mg/mL) compared to control norfloxacin (2.33 mg/mL) (Santos et al., 2021). Bark hexane and dichloromethane extracts and their fractions show inhibitory activity on the quorum sensing system (a communication system between bacteria) of *P. aeruginosa* and *Chromobacterium violaceum*. They also can act on virulence factors such as pyocyanin, alkaline protease and violacein production, being associated with the large quantity of steroids, terpenes, and flavonoids present (Muñoz-Cázares et al., 2018).

Ethanol and aqueous extracts from different parts of *C. pentandra* show weak activity against fungi that cause dermatitis (*Epidermophyton floccosum*, *Microsporium canis*, *Trichopyton rubrum* and *Candida albicans*), with values of minimum inhibitory concentration (MIC) higher than 50 mg/mL for all microorganisms (Nwachukwu et al., 2008).

Afzal et al. (2022) investigated the antibacterial and antibiofilm activity of silver nanoparticles (AgNPs) synthesized with methanolic extracts from the bark (AgNPb) and leaf (AgNPl) of *C. pentandra*. Using the plate diffusion methodology, significant antimicrobial activity of AgNPl was observed, with zones of inhibition of 8.6 ± 1.2 mm, 15.0 ± 1.0 mm, and 9.3 ± 2.3 mm against the microorganisms *P. aeruginosa*, *E. coli* and *S. aureus*, respectively, and for AgNPb, with 14.7 ± 1.2 mm, 15.3 ± 0.6 mm, and 12.7 ± 2.3 mm for the same microorganisms. The minimum inhibitory

concentration (MIC) for AgNPb was 63 $\mu\text{g/mL}$ against *S. aureus*, and 125 $\mu\text{g/mL}$ against *P. aeruginosa* and *E. coli*, and for AgNPl, it was 125 $\mu\text{g/mL}$ against *E. coli* and *S. aureus* and 250 $\mu\text{g/mL}$ against *S. aureus* and *P. aeruginosa* for AgNPb. Regarding biofilm formation, AgNPb showed greater inhibition against *E. coli*, and AgNPl showed moderate activity against the tested microorganisms. The presented antimicrobial potential is attributed to the presence of active biomolecules, such as alkaloids, flavonoids, tannins, and glycosides.

Antiviral action can be observed within its leaves ethanolic extract, which shows expressive inhibition of human liver cells infectivity to dengue virus (DENV), with an IC_{50} of 15.49 $\mu\text{g/mL}$, and low half-cytotoxic concentration (CC_{50}) of 81.1 $\mu\text{g/mL}$ at the lowest concentrations tested, resulting in an infectivity index of 5.23 (Dewi et al., 2019). This antimicrobial and antiviral activity is attributed to the presence of a great diversity of secondary metabolites compounds, especially belonging to flavonoids and tannins classes.

Stem bark ethanolic extract showed strong antitrypanosomal activity against *Trypanosoma brucei brucei*, with an IC_{50} of 11.70 $\mu\text{g/mL}$. Anthelmintic activity was also observed, killing *Phetima posthuma* after the experimental exposure time at 2.5, 5 and 10 mg/mL. These activities may be due to a high concentration and number of tannins and presence of n-hexadecanoic acid, a compound known for its antitrypanosomal potential (Wahab Obeng et al., 2022).

Anticancer potential was also observed as in dichloromethane fraction of the aerial parts methanolic extract, which showed promising cytotoxicity against cancer cells, presenting IC_{50} values of 14,89 and 18,86 $\mu\text{g/mL}$, against cell lines of hepatocellular carcinoma (HepG2) and breast cancer (MCF-7), respectively. Molecules isolated from this same fraction also had anticancer activity through molecular docking analysis. The compounds 3 β -taraxerol, 3 β -taraxerol acetate, all-trans-squalene and β -amyrin, have predicted activity for the treatment of diseases proliferative, apoptosis agonists and antineoplastic agents (Abouelela et al., 2018).

Silver nanoparticles synthesized with bark ethanolic extract produce a significant cytotoxic effect against human colon cancer cells (HCT-116) with a IC_{50} of 60 $\mu\text{g/mL}$ in MTT assay, inducing increased production of reactive oxygen species, which leads to cell membrane damage and subsequently apoptosis (Brian & Selvi, 2019).

Petroleum ether (PE) and acetone (AC) extracts from *C. pentandra* bark showed robust short-term cytotoxic effects against Ehrlich ascites carcinoma (EAC) cells and long-term cytotoxic effects on human breast cancer cell lines (MCF-7) and melanoma cell lines (B16F10). Mice inoculated by EAC-induced liquid tumor and treated with the extracts had increased survival, and in Dalton's lymphoma ascites (DLA)-induced solid tumor model, there was a significant reduction in tumor weight and volume (about 50%) compared to negative control. Hematological parameters (WBC, RBC and hemoglobin content) and endogenous antioxidant levels in liver (catalase, SOD, GSH, GST and MDA) were improved, and

these may play an important role in antitumor activity of these extracts (Kumar et al., 2016).

Seeds fixed oil has in vivo anti-inflammatory potential confirmed, through oral administration in mice, reducing the serum concentration of the inflammatory marker C-reactive protein. It was observed values of 1.08 ± 0.02 and 1.08 ± 0.03 mg/L to 50 and 100 mg/Kg, respectively; similar results to the control sodium diclofenac (1.04 ± 0.04 mg/L). The fixed oil shown blood cells membrane-stabilization, with maximum inhibition 28.8 % at 50 mg/mL. Membrane-stabilizing compounds can act in the initial phase of inflammation, preventing the release of phospholipases that trigger the formation of cytokines. The inhibition of production and function of these inflammatory mediators are fundamental in controlling the inflammatory process (Ravi Kiran & Raghava Rao, 2014).

Antipyretic activity was observed in leaves ethanolic extract, using murine models, with higher effect at 189 mg/Kg. There was a temperature variation of -0.85 ± 0.58 °C after 4h of fever induction, compared to the pre-treatment temperature. One of the possible mechanisms for the antipyretic action may be associated with inhibition of prostaglandin E2 (PGE2), responsible for adjusting body thermoregulation in cases of fever (Saptarini & Deswati, 2015). Parhan (2021) evaluated antipyretic potential of leaves ethanolic extract in rats with high body temperature induced by DPT-HB vaccine, finding moderate potential at a concentration of 300 mg/kg of body weight.

6.2. *Ceiba speciosa*

Recent studies show that *C. speciosa* has antidiabetic properties, which confirms its ethnopharmacological use, used to reduce cholesterol, triglycerides, and blood glucose levels (Malheiros et al., 2017). Bark aqueous extract can increase glucose utilization and regulate insulin levels. These results were observed in protocols with the nematode *Caenorhabditis elegans*, with reduced body glucose levels and increased longevity of the animals, protecting them against glucotoxicity (Santos et al., 2020). Seed fixed oil has anti-obesity potential, by slightly inhibiting α -amylase, α -glucosidase, and pancreatic lipase enzymes activity, with IC_{50} values of 135.69, 158.22 and 127.57 μ g/mL, respectively (Rosselli et al., 2020).

Antioxidant capacity of *C. speciosa* stem bark aqueous extract was evaluated observing high values by the 2,2-Diphenyl-1-picrylhydrazyl (DPPH) radical scavenging methodology (IC_{50} 42.87 μ g/mL), moderate effect on ferric reducing antioxidant power assay (FRAP) (236 ± 18 μ M $FeSO_4/g$) and slight activity by the oxygen radical absorbent capacity assay (ORAC) (2351 ± 136 μ mol Trolox Equivalent/g). These results are attributed to kaempferol presence, a flavonoid known for its antioxidant activity, in addition to other phenolic compounds present in this extract (Dörr et al., 2019).

In another study, the antioxidant activity of bark trunk extracts against the DPPH radical was evaluated, obtaining percentages of 85.13 % and 88.95 % (50 μ g/mL) of

inhibition, to aqueous and ethanolic extracts, respectively. Other concentrations were also tested (10, 5 and 2 μ g/mL) for both extracts and decreasing of activity in dose-dependence was observed (Malheiros et al., 2017). These results differ significantly from those presented by Nasr et al. (2018), that, evaluating *C. speciosa* bark ethanolic extract, found a lower inhibition percentage, even though concentration tested was higher (1 mg/mL), obtaining 81.2 % of radical scavenging. The specimens collected in the studies come from different geographic locations and climatic conditions. It is expected that there will be variations in the synthesis of secondary metabolites, and therefore in the intensity of the biological activity evaluated.

The antioxidant potential of *C. speciosa* leaves ethanolic was 78.7 % for DPPH radical scavenging (at 1 mg/mL), with this activity attributed to the presence of phenolic compounds, especially flavonoids (Nasr et al., 2018). Methanolic extract of leaves and its dichloromethane fraction presented antioxidant potential similar to the standard, with IC_{50} of 15.48 and 12.37 μ g/mL, compared to ascorbic acid (7.60 μ g/mL) (Abdel-Aziz et al., 2021). Seed fixed oil showed strong IC_{50} values, with 10.21 μ g/mL and 77.44 μ g/mL for ABTS and DPPH radical scavenging activity assays, respectively (Rosselli et al., 2020).

Flower pigment of *C. speciosa* (FPCS) presents strong antioxidant effect, with scavenging the DPPH and ABTS free radical with an IC_{50} of 49.78 and 58.14 μ g/mL, respectively. Pretreatment with FPCS prevents lipopolysaccharide-induced hippocampal oxidative stress in a model using Kunming mice. There was a reduction in the expression of the oxidative stress biomarkers nitric oxide synthase (iNOS) and heat shock protein 60 (HSP60) at concentrations of 30 and 60 mg/kg. There was no acute oral toxicity at the concentrations tested (1 - 10 g/Kg) (Chen et al., 2022).

The antipyretic activity was evaluated in hydroalcoholic extracts of stem and leaves of mice previously inoculated with yeast. A relevant decrease in body temperature was observed in mice previously inoculated with yeast, with maximum effect (400 mg/Kg) reached about the third hour of the experiment, for both extracts. In the evaluation of anti-inflammatory activity by the paw edema methodology, there was a moderate reduction in edema. The most expressive inhibition values were observed after 1 h of test (3-5 h), both for the ethanolic extracts of leaves and stem, and for the isolated compound rhoifolin. This indicates that the extract acts in the second phase of the inflammatory process, and the presence of flavonoids, sterols and triterpenes in these extracts may be associated with inhibition of the release of lysosomal enzymes and prostaglandins (Nasr et al., 2018).

Antiulcerogenic and anti-inflammatory activity of stem bark ethanolic extract of *Ceiba speciosa* was evaluated, observing the viability of cells of gastric origin (MN01, ACP02 and ACP03), and murine macrophage (RAW264.7) in which ethanolic extract (100 μ g/mL) showed a proliferative effect. It is also observed the inhibition of the activity of p38 α (1.66 μ g/mL), JAK3 (5.25 μ g/mL), and JNK3 (8.34 μ g/mL), and reduction of the release of TNF- α and IL-6 by murine macrophage. In

an in vivo evaluation, it reduced the recruitment of leukocytes, significantly reverting the production of NO and reducing the formation of edema in a dose-dependent manner at the concentrations tested (10, 50 and 100mg/Kg), reversing the effects caused by carrageenan. The extract showed significant prevention of ulcer formation at the tested concentrations (20-400mg/Kg) with a greater reduction than the reference drug, Omeprazole (40mg/Kg). This potential is attributed to the phenolic compounds present in the extract, which have antioxidant activity, inhibiting oxidative stress, reducing NO production, and preventing the activation of p38, which, in turn, inhibits TNF- α and IL-6, exhibiting activity anti-inflammatory and anti-ulcerogenic properties (Dörr et al., 2022).

The anticancer activity was evaluated with fractions of the methanolic extract of the leaves of *C. speciosa*, being observed moderate cytotoxicity against the cell line HepG2. The petroleum ether, ethyl acetate and dichloromethane fractions presented IC₅₀ of 74.35, 79.73 and 57.30 μ g/mL, respectively. Dichloromethane fraction still present strong antimicrobial activity, with clear zone of 23, 22, 22 and 21 (ϕ mm) on *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Candida albicans* and *Aspergillus niger*, respectively (Abdel-Aziz et al., 2021).

6.3. Other species

Hydroalcoholic extract of *Ceiba glaziovii*, in an antimicrobial activity evaluation against *Staphylococcus aureus*, showed a not very expressive result in inhibiting bacterial growth, for both bark and leaf extracts (Leal et al., 2011). *C. pubiflora* leaves hydroethanolic extract has antioxidant potential evaluate by DPPH radical scavenging methodology, with an IC₅₀ of 217.4 \pm 0.19 mg/L (Menezes-Filho et al., 2022).

Purified compounds from *Ceiba* species are also investigated, such as rhoifolin, isolated from *C. crispiflora* leaves. This compound has neuroprotective properties evaluated in models with zebra fish, being able to improve symptoms of anxiety, memory deficit and brain oxidative stress, inhibiting acetylcholinesterase activity and reestablishing cholinergic activity, thus being a promising compound for amnesia and anxiety treatment (Brinza et al., 2020).

7. CONCLUSION

The number of studies with species of the genus *Ceiba* has been increased in the last decade, and this is due to the necessity of investigation on their use in folk medicine. Nonetheless, species endemic to Brazil are still poor in scientific evidence. This might be a result of their distribution, semi-arid region, where the flora is poorly studied, and the competition with other species that have economic value, often preferred targets for scientific studies.

Because of the high medicinal and pharmacological potential reported for this genus, it is necessary that new studies are carried out, in order to investigate the bioactive properties of *Ceiba* species not yet studied. This knowledge is fundamental for the process of preservation, validation of traditional uses,

and local economy development through the bioeconomy. In addition, new studies on these plants can help in the prospection of compounds of biotechnological interest that can be applied in the most diverse industry areas, such as food, cosmetics and pharmacological.

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REFERENCES

- Abdel-Aziz, A.W.A., Elwan, N.M., Abdallah, M.A., Shaaban, R., Mohamed, N.S., Mohamed, M.A., 2021. High-performance liquid chromatography-fingerprint analyses, in vitro cytotoxicity, antimicrobial and antioxidant activities of the extracts of *Ceiba speciosa* growing in Egypt. *Egyptian Journal of Chemistry*. 64(4), 1831–1843. <https://doi.org/10.21608/EJCHEM.2021.58716.3267>
- Abouelela, M.E., Orabi, M.A.A., Abdelhamid, R.A., Abdelkader, M.S., Madkor, H.R., Darwish, F.M.M., Hatano, T., Elsadek, B.E.M., 2020. Ethyl acetate extract of *Ceiba pentandra* (L.) Gaertn. reduces methotrexate-induced renal damage in rats via antioxidant, anti-inflammatory, and antiapoptotic actions. *Journal of Traditional and Complementary Medicine*. 10(5), 478–486. <https://doi.org/10.1016/j.jtcm.2019.08.006>
- Abouelela, M.E., Orabi, M.A.A., Abdelhamid, R.A., Abdelkader, M.S.A., Darwish, F.M.M., 2018. Chemical and cytotoxic investigation of non-polar extract from *Ceiba pentandra* (L.) Gaertn.: A study supported by computer based screening. *Journal of Applied Pharmaceutical Science*. 8(7), 57–64. <https://doi.org/10.7324/JAPS.2018.8710>
- Abouelela, M.E., Orabi, M.A.A., Abdelhamid, R.A., Abdelkader, M.S.A., Darwish, F.M.M., Hotsumi, M., Konno, H., 2020. Anti-Alzheimer's flavanolignans from *Ceiba pentandra* aerial parts. *Fitoterapia*. 143. <https://doi.org/10.1016/j.fitote.2020.104541>
- Abubakar, A.R., Haque, M., 2020. Preparation of medicinal plants: Basic extraction and fractionation procedures for experimental purposes. *Journal of Pharmacy and Bioallied Sciences*. 12, 1–10. https://doi.org/10.4103/jpbs.JPBS_175_19
- Abubakar, M.S., Musa, A.M., Ahmed, A., Hussaini, I.M., 2007. The perception and practice of traditional medicine in the treatment of cancers and inflammations by the Hausa and Fulani tribes of Northern Nigeria. *Journal of Ethnopharmacology*. 111(3), 625–629. <https://doi.org/10.1016/j.jep.2007.01.011>
- Afzal, S., Qurashi, A.W., Sarfraz, B., Liaqat, I., Sadiqa, A., Muhtaq, M., Andleeb, S., Ahsan, F., 2022. A Comparative Analysis of Antimicrobial, Antibiofilm and Antioxidant Activity of Silver Nanoparticles Synthesized from *Erythrina Suberosa* Roxb. and *Ceiba Pentandra*. *Journal of Oleo Science*. 71(4), 523–533. <https://doi.org/10.5650/jos.ess21347>
- Agra, M.F., Baracho, G.S., Nurit, K., Basílio, I.J.L.D., Coelho, V.P.M., 2007. Medicinal and poisonous diversity of the flora of “Cariri Paraibano”, Brazil. *Journal of Ethnopharmacology*. 111(2), 383–395. <https://doi.org/10.1016/j.jep.2006.12.007>
- Agra, M.F., Silva, K.N., Basílio, I.J.L.D., Freitas, P.F., Barbosa-Filho, J.M.,

2008. Survey of medicinal plants used in the region Northeast of Brazil. *Revista Brasileira de Farmacognosia*. 18(3), 472–508. <https://doi.org/10.1590/S0102-695X2008000300023>
- Akaneme, F.I., 2008. Identification and preliminary phytochemical analysis of herbs that can arrest threatened miscarriage in Orba and Nsukka towns of Enugu State. *African Journal of Biotechnology*. 7(1), 6–11. <https://doi.org/10.5897/AJB07.682>
- Alade, A., Satyal, P., Setzer, W.N., Aboaba, S.A., Alade, A.T., Aboaba, S.O., 2021. Chemical profiles and brine shrimp toxicity of volatile oils hydrodistilled from stem bark and heartwood of *Ceiba pentandra* Linn. *American Journal of Essential Oils and Natural Products*. 9(3), 22–26.
- Albuquerque, U.P., Medeiros, P.M., Almeida, A.L.S., Monteiro, J.M., Neto, L., Melo, E.M.F., Santos, J.G., P, J., 2007. Medicinal plants of the caatinga (semi-arid) vegetation of NE Brazil: A quantitative approach. *Journal of Ethnopharmacology*. 114(3), 325–354. <https://doi.org/10.1016/j.jep.2007.08.017>
- Albuquerque, U.P., Soldati, G.T., Sieber, S.S., Medeiros, P.M., Sá, J.C., Souza, L.C., 2011. Rapid ethnobotanical diagnosis of the Fulni-ô Indigenous lands (NE Brazil): Floristic survey and local conservation priorities for medicinal plants. *Environment, Development and Sustainability*. 13(2), 277–292. <https://doi.org/10.1007/s10668-010-9261-9>
- Almeida, C.F.C.B.R., Silva, T.C.L.E., Amorim, E.L.C., Maia, M.B.S., Albuquerque, U.P., 2005. Life strategy and chemical composition as predictors of the selection of medicinal plants from the caatinga (Northeast Brazil). *Journal of Arid Environments*. 62(1), 127–142. <https://doi.org/10.1016/j.jaridenv.2004.09.020>
- Anwar, F., Rashid, U., Shahid, S.A., Nadeem, M., 2014. Physicochemical and antioxidant characteristics of kapok (*Ceiba pentandra* Gaertn.) seed oil. *Journal of the American Oil Chemists' Society*. 91(6), 1047–1054. <https://doi.org/10.1007/s11746-014-2445-y>
- APG IV., 2017. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants. <http://www.mobot.org/MOBOT/research/APweb/> Date accessed: 22nd august 2022.
- Bairwa, N.K., Sethiya, N.K., Mishra, S.H., 2010. Protective effect of stem bark of *Ceiba pentandra* Linn. against paracetamol-induced hepatotoxicity in rats. *Pharmacognosy Research*. 2(1), 26–30. <https://doi.org/10.4103/0974-8490.60584>
- Beleski-Carneiro, E.B., Ganter, J.L.M.S., Reicher, F., 1999. Structural aspects of the exudate from the fruit of *Chorisia speciosa* St. Hil. *International Journal of Biological Macromolecules*. 26(4), 219–224. [https://doi.org/10.1016/S0141-8130\(99\)00086-0](https://doi.org/10.1016/S0141-8130(99)00086-0)
- Bhavani, R., Bhuvanewari, E., Rajeshkumar, S., 2016. Antibacterial and antioxidant activity of ethanolic extract of *Ceiba pentandra* leaves and its phytochemicals analysis using GC-MS. *Research Journal of Pharmacy and Technology*. 9(11), 1922–1926. <https://doi.org/10.5958/0974-360X.2016.00393.0>
- Bhuvanewari, S., Balaji, R., Prakash, U., Kannaian, N., 2014. Studies on the Phytochemistry and Bioactivity of Leaves of Trees in Chennai-I. *International Journal of ChemTech Research*. 6(9), 4078–4083.
- Bizimana, N., Tietjen, U., Zessin, K.H., Diallo, D., Djibril, C., Melzig, M.F., Clausen, P.H., 2006. Evaluation of medicinal plants from Mali for their in vitro and in vivo trypanocidal activity. *Journal of Ethnopharmacology*. 103(3), 350–356. <https://doi.org/10.1016/j.jep.2005.08.023>
- Bolson, M., Hefler, S.R., Chaves, E.I., Junior, A.G., Junior, E.L.C., 2015. Ethno-medicinal study of plants used for treatment of human ailments, with residents of the surrounding region of forest fragments of Paraná, Brazil. *Journal of Ethnopharmacology*. 161, 1–10. <https://doi.org/10.1016/j.jep.2014.11.045>
- Braga, P.M., Do, S., Barcelos, I.B., Calazans, R.D.S.P., Bulian, A.L.S., Gabler, J.C.R., Sobral, F.D.O.S., Salvi, J.D.O., 2019. Análise fitoquímica, toxicidade, potencial antioxidante e atividade antibacteriana da *Ceiba speciosa* (A.St.-Hil.) Ravenna. *Revista Fitos*. 13(1), 9–21. <https://doi.org/10.17648/2446-4775.2019.641>
- Brian, M.O., Selvi, S., 2019. Cytotoxic effects of *Ceiba pentandra* L. Mediated silver nanoparticles on HCT-116 colon cancer cell lines through ROS generation and cell membrane damage. *International Journal of Research in Pharmaceutical Sciences*. 10(4), 3236–3243. <https://doi.org/10.26452/ijrps.v10i4.1627>
- Brinza, I., Abd-Alkhalik, A.M., El-Raey, M.A., Boiangiu, R.S., Eldahshan, O.A., Hritcu, L., 2020. Ameliorative effects of rhoifolin in scopolamine-induced amnesic zebrafish (*Danio rerio*) model. *Antioxidants*. 9(7), 1–14. <https://doi.org/10.3390/antiox9070580>
- Cartaxo, S.L., Souza, M.M.A., Albuquerque, U.P., 2010. Medicinal plants with bioprospecting potential used in semi-arid northeastern Brazil. *Journal of Ethnopharmacology*. 131(2), 326–342. <https://doi.org/10.1016/j.jep.2010.07.003>
- Chen, B., Misrani, A., Long, C., He, Z., Chen, K., Yang, L., 2022. Pigment of *Ceiba speciosa* (A. St.-Hil.) Flowers: Separation, Extraction, Purification and Antioxidant Activity. *Molecules*. 27(11), 3555–3555. <https://doi.org/10.3390/molecules27113555>
- Dewi, B.E., Angelina, M., Ardiantara, S., Prakoso, A.R., Desti, H., Sudiro, T.M., 2019. Antiviral activity of *Ceiba pentandra* and *Eugenia uniflora* leaf extracts to dengue virus serotype-2 in Huh 7it-1 cell line. *AIP Conference Proceedings*. 2193, 030003. <https://doi.org/10.1063/1.5139340>
- Dickson, R.A., Annan, K., Fleischer, T.C., Amponsah, I.K., Nsiah, K., Oteng, J.A., 2012. Phytochemical investigations and nutritive potential of eight selected plants from Ghana. *Journal of Pharmacy and Nutrition Sciences*. 2(2), 172–177. <https://doi.org/10.6000/1927-5951.2012.02.02.9>
- Dörr, J.A., Bitencourt, S., Bortoluzzi, L., Alves, C., Silva, J., Stoll, S., Pinteus, S., Boligon, A.A., Santos, R.C.V., Laufer, S., Pedrosa, R., Goettert, M.I., 2019. In vitro activities of *Ceiba speciosa* (A.St.-Hil) Ravenna aqueous stem bark extract. *Natural Product Research*. 33(23), 3441–3444. <https://doi.org/10.1080/14786419.2018.1478823>
- Dörr, J.A., Majolo, F., Bortoluzzi, L., de Vargas, E.Z., Silva, J., Pasini, M., Stoll, S.N., Rosa, R.L.D., Figueira, M.M., Fronza, M., Beys-Da-Silva, W.O., Martins, A., Gaspar, H., Pedrosa, R.P., Laufer, S., Goettert, M.I., 2022. Antiulcerogenic Potential of the Ethanolic Extract of *Ceiba speciosa* (A. St.-Hil.) Ravenna Evaluated by In Vitro and In Vivo Studies. *International Journal of Molecular Sciences*. 22(22), 15634–15634. <https://doi.org/10.3390/ijms232415634>
- El-Kamali, H.H., El-Khalifa, K.F., 1999. Folk medicinal plants of riverside forests of the Southern Blue Nile district. *Fitoterapia*. 70(5), 73–81. [https://doi.org/10.1016/S0367-326X\(99\)00073-8](https://doi.org/10.1016/S0367-326X(99)00073-8)
- Figueiredo, S.S., Monteiro, F.K., Da, S., de Melo, J.I.M., 2020. Flora of Paraíba, Brazil: Bombacoideae burnett (Malvaceae). *Biota Neotropica*. 20(2), e20190837. <https://doi.org/10.1590/1676-0611-BN-2019-0837>
- Fofie, C.K., Katekhaye, S., Borse, S., Sharma, V., Nivsarkar, M., Nguetefack-Mbuyo, E.P., Kamanyi, A., Singh, V., Nguetefack, T.B., 2019. Antidiabetic properties of aqueous and methanol extracts from the trunk bark of *Ceiba pentandra* in type 2 diabetic rat. *Journal of Cellular Biochemistry*. 120(7), 11573–11581. <https://doi.org/10.1002/jcb.28437>
- Fofie, C.K., Nguetefack-Mbuyo, E.P., Tsbang, N., Kamanyi, A., Nguetefack, T.B., 2018. Hypoglycemic properties of the aqueous extract from the stem bark of *Ceiba pentandra* in dexamethasone-induced insulin resistant rats. *Evidence-based Complementary and Alternative*

- Medicine, 4234981. <https://doi.org/10.1155/2018/4234981>
- Fofie, C.K., Wansi, S.L., Nguielefack-Mbuyo, E.P., Atsamo, A.D., Watcho, P., Kamanyi, A., Nole, T., Nguielefack, T.B., 2014. In vitro anti-hyperglycemic and antioxidant properties of extracts from the stem bark of *Ceiba pentandra*. *Journal of Complementary and Integrative Medicine*. 11(3), 185–193. <https://doi.org/10.1515/jcim-2014-0031>
- García-Flores, J., González-Espinosa, M., Lindig-Cisneros, R., Casas, A., 2019. Traditional medicinal knowledge of tropical trees and its value for restoration of tropical forests. *Botanical Sciences*. 97(3), 336–354. <https://doi.org/10.17129/botsci.2122>
- Gibbs, P., Semir, J., 2003. A Taxonomic Revision of the Genus *Ceiba* Mill. (Bombacaceae). *Anales del Jardín Botánico de Madrid*. 60(2), 259–300. <https://doi.org/10.3989/ajbm.2002.v60.i2.92>
- Gibbs, P.E., Semir, J., Cruz, N.D., 1988. A proposal to unite the genera *Chorisia* Kunth and *Ceiba* Miller (Bombacaceae). *Notes Of The Royal Botanic Garden Edinburgh*. 45, 125–136.
- Gómez-Maqueo, X., Gamboa-Debuen, A., 2022. The Biology of the Genus *Ceiba*, a Potential Source for Sustainable Production of Natural Fiber. *Plants*. 11(4), 521. <https://doi.org/10.3390/plants11040521>
- Herzog, F., Farah, Z., Amadò, R., 1993. Nutritive Value of Four Wild Leafy Vegetables in Côte d'Ivoire. *International Journal for Vitamin and Nutrition Research*. 63(3), 234–238.
- Ibara, J.R., Elion-Itou, R.D.G., Etou-Ossebi, A., Ouamba, J.M., Diatéwa, M., Abena, A.A., 2007. Enquête ethnobotanique à propos de plantes médicinales Congolaises présumées anti-ulcéreuses. *Phytothérapie*. 5, 118–120. <https://doi.org/10.1007/s10298-007-0236-4>
- Ibrahim, S., Nok, J.A., Abubakar, M.S., Sarkiyayi, S., 2012. Efficacy of Di-n-octyl Phthalate Anti Venom Isolated from *Ceiba pentandra* Leaves Extract in Neutralization of *Echis ocellatus* Venom. *Research Journal of Applied Sciences, Engineering and Technology*. 4(15), 2382–2387.
- Ijioma, S.N., Osim, E.E., Nwankwo, A.A., Kanu, K.C., Oriek, D., 2020. Southeast Nigerian polyherbal (AJUMBISE): A potential uterotonic and tocolytic agents. *Scientific African*. 8, e00393. <https://doi.org/10.1016/j.sciaf.2020.e00393>
- Isah, T., 2019. Stress and defense responses in plant secondary metabolites production. *Biological Research*. 52, 39–39. <https://doi.org/10.1186/s40659-019-0246-3>
- Kan, W.S., 1975. *Pharmaceutical Botany*. National Research Institute of Chinese Medicine. Taiwan, CN. Taiwan, CN.
- Kausar, F., Intisar, A., Din, M.I., Aamir, A., Hussain, T., Aziz, P., Mutahir, Z., Fareed, S., Samreen, B., Sadaqat, K., 2020. Volatile composition and antibacterial activity of leaves of *Chorisia speciosa*. *Journal of the Mexican Chemical Society*. 64(4), 339–348. <https://doi.org/10.29356/jmcs.v64i4.1440>
- Kiran, C.R., Rao, D.B., Sirisha, N., Rao, T.R., 2015. Assessment of phytochemicals and antioxidant activities of raw and germinating *Ceiba pentandra* (kapok) seeds. *Journal of Biomedical Research*. 29(5), 414–419. <https://doi.org/10.7555/JBR.27.20120145>
- Kishore, P.H., Reddy, M.V.B., Gunasekar, D., Caux, C., Bodo, B., 2003. A new naphthoquinone from *Ceiba pentandra*. *Journal of Asian Natural Products Research*. 5(3), 227–230. <https://doi.org/10.1080/1028602031000105812>
- Kone, W.M., Azokou, A., Bakayoko, A., Bi, F.H.T., 2012. Preliminary Study on Polysaccharides and Certain Secondary Metabolites of Medicinal Plants used in Cote D'Ivoire for Wound Healing. *Research Journal of Medicinal Plant*. 6(3), 214–224. <https://doi.org/10.3923/rjmp.2012.214.224>
- Krishnaveni, M., Amsavalli, L., Chandrasekar, R., Madhaiyan, P., Durairaj, S., 2013. Antioxidant activity of Plants at Govt. College of Engineering Campus, Salem, Tamil nadu, India. *International Journal of Pharmaceutical Sciences Review and Research*. 21(1), 160–163.
- Kubmarawa, D., Ajoku, G.A., Enwerem, N.M., Okorie, D.A., 2007. Preliminary phytochemical and antimicrobial screening of 50 medicinal plants from Nigeria. *African Journal of Biotechnology*. 6(14), 1690–1696.
- Kumar, R., Kumar, N., Ramalingayya, G.V., Setty, M.M., Pai, K.S.R., 2016. Evaluation of *Ceiba pentandra* (L.) Gaertner bark extracts for in vitro cytotoxicity on cancer cells and in vivo antitumor activity in solid and liquid tumor models. *Cytotechnology*. 68(5), 1909–1923. <https://doi.org/10.1007/s10616-016-0002-2>
- Lami, M.H., Yusuf, K.A., Ndaman, S.A., Bola, B.M., Damilola, B.O., Siddique, A.A., 2015. Ameliorative properties of ethyl acetate fraction of *Ceiba pentandra* on serum glucose, hematological and biochemical parameters of diabetic rats. *Asian Pacific Journal of Tropical Disease*. 5(9), 737–742. [https://doi.org/10.1016/S2222-1808\(15\)60923-9](https://doi.org/10.1016/S2222-1808(15)60923-9)
- Leal, A.J.B., Dantas, I.C., Chaves, T.P., Felismino, D.C., Vieira, K.V.M., 2011. Estudo Fitoquímico Antimicrobiano de *Ceiba glaziovii* Kuntze K. Schum. *Revista de Biologia e Farmácia*. 5(1), 73–77.
- Leitão, F., Fonseca-Kruel, V.S., Silva, I.M., Reinert, F., 2009. Urban ethnobotany in Petrópolis and Nova Friburgo. *Revista Brasileira de Farmacognosia*. 19(1B), 333–342. <https://doi.org/10.1590/S0102-695X2009000200026>
- Leonti, M., Sticher, O., Heinrich, M., 2003. Antiquity of medicinal plant usage in two Macro-Mayan ethnic groups (México). *Journal of Ethnopharmacology*. 88(2-3), 119–124. [https://doi.org/10.1016/S0378-8741\(03\)00188-0](https://doi.org/10.1016/S0378-8741(03)00188-0)
- Maeda, H.A., 2019. Harnessing evolutionary diversification of primary metabolism for plant synthetic biology. *Journal of Biological Chemistry*. 294(45), 16549–16566. <https://doi.org/10.1074/jbc.REV119.006132>
- Malheiros, C.K.C., Silva, J.S.B., Hofmann, T.C., Messina, T.M., Manfredini, V., Piccoli, J.C.E., Faoro, D., Oliveira, L.F.S., Machado, M.M., Farias, F.M., 2017. Preliminary in vitro assessment of the potential toxicity and antioxidant activity of *Ceiba speciosa* (A. St.-Hill) ravenna (paineira). *Brazilian Journal of Pharmaceutical Sciences*. 53(2), e16098. <https://doi.org/10.1590/s2175-97902017000216098>
- Manousi, N., Sarakatsianos, I., Samanidou, V., 2019. Extraction Techniques of Phenolic Compounds and Other Bioactive Compounds From Medicinal and Aromatic Plants. In: *Engineering Tools in the Beverage Industry*. Elsevier, 283–314. <https://doi.org/10.1016/b978-0-12-815258-4.00010-x>
- Menezes-Filho, A.C.P., Santos, M.C., Castro, C.F.S., Oliveira-Filho, J.G., Christofoli, M., 2022. Phytochemical prospection and biological activities of the floral extract *Ceiba pubiflora* St. -hill. (Malvaceae). *Revista Cubana de Química*. 34(2), 333–348.
- Mohanty, S., Sharma, M., Dubey, K., Alexander, A., Khan, J., Khan, S., Tripathi, D., 2012. Significance of Herbal Plants in the Treatment of Sickle Cell Anaemia. *Research Journal of Pharmacy and Technology*. 5(3), 297–301.
- Moraes, A.J., 1881. *Phytografia ou Botânica Brasileira Aplicada à Medicina, às Artes e à Indústria*. Garnier, Rio de Janeiro, BR, 74.
- Mpiana, P.T., Tshibangu, D.S.T., Shetonde, O.M., Ngbolua, K.N., 2007. In vitro antitrepanocytary activity (anti-sickle cell anemia) of some congolese plants. *Phytomedicine*. 14(2-3), 192–195. <https://doi.org/10.1016/j.phymed.2006.05.008>
- Muñoz-Cázarez, N., Aguilar-Rodríguez, S., García-Contreras, R., Soto-Hernández, M., Martínez-Vázquez, M., Palma-Tenango, M., Prado-Galbarro, F.J., Castillo-Juárez, I., 2018. Phytochemical screening and antivirulence properties of *Ceiba pentandra* and *Ceiba aesculifolia* (Malvaceae) bark extracts and fractions. *Botanical Sciences*. 96(3),

- 415–425. <https://doi.org/10.17129/botsci.1902>
- Nasr, E.M., Assaf, M.H., Darwish, F.M., Ramadan, M.A., 2018. Phytochemical and biological study of *Chorisia speciosa* A. St. Hil. Cultivated in Egypt. *Journal of Pharmacognosy and Phytochemistry*. 7(1), 649–656.
- Ndenecho, E.N., 2009. Herbalism and resources for the development of ethnopharmacology in Mount Cameroon region. *African Journal of Pharmacy and Pharmacology*. 3(3), 78–86.
- Ngounou, F.N., Meli, A.L., Lontsi, D., Sondengam, B.L., Rahman, A.U., Choudhary, M.I., Malik, S., Akhtar, F., 2000. New isoflavones from *Ceiba pentandra*. *Phytochemistry*. 54(1), 107–110. [https://doi.org/10.1016/S0031-9422\(00\)00035-2](https://doi.org/10.1016/S0031-9422(00)00035-2)
- Nguelefack, T.B., Fofie, C.K., Nguelefack-Mbuyo, E.P., Wuyt, A.K., 2020. Multimodal α -Glucosidase and α -Amylase Inhibition and Antioxidant Effect of the Aqueous and Methanol Extracts from the Trunk Bark of *Ceiba pentandra*. *BioMed Research International*. 3063674. <https://doi.org/10.1155/2020/3063674>
- Nguyen, M.T.T., Awale, S., Tezuka, Y., Tran, Q., Le, Watanabe, H., Kadota, S., 2004. Xanthine Oxidase Inhibitory Activity of Vietnamese Medicinal Plants. *Biological and Pharmaceutical Bulletin*. 27(9), 1414–1421. <https://doi.org/10.1248/bpb.27.1414>
- Noreen, Y., El-Seedi, H., Perera, P., Bohlin, L., 1998. Two New Isoflavones from *Ceiba pentandra* and Their Effect on Cyclooxygenase-Catalyzed Prostaglandin Biosynthesis. *Journal of Natural Products*. 61(1), 8–12. <https://doi.org/10.1021/np970198+>
- Noumi, E., Tchakonang, N.Y.C., 2001. Plants used as abortifacients in the Sangmelima region of Southern Cameroon. *Journal of Ethnopharmacology*. 76(3), 252–257. [https://doi.org/10.1016/S0378-8741\(01\)00252-5](https://doi.org/10.1016/S0378-8741(01)00252-5)
- Nsimba, M.M., Yamamoto, C., Lami, J.N., Hayakawa, Y., Kaji, T., 2013. Effect of a Congolese herbal medicine used in sickle cell anemia on the expression of plasminogen activators in human coronary aortic endothelial cells culture. *Journal of Ethnopharmacology*. 146(2), 594–599. <https://doi.org/10.1016/j.jep.2013.01.031>
- Nwachukwu, I.N., Allison, L.N., Chinakwe, E.C., Nwadiaro, P., 2008. Studies on the effects of *Cymbopogon citratus*, *Ceiba pentandra* and *Loranthus bengwelensis* extracts on species of dermatophytes. *The Journal of American Science*. 4(4), 58–67. <https://doi.org/10.7537/marsjas040408.10>
- Odonne, G., Berger, F., Stien, D., Grenand, P., Bourdy, G., 2011. Treatment of leishmaniasis in the Oyapock basin (French Guiana): A K.A.P. survey and analysis of the evolution of phytotherapy knowledge amongst Wayápi Indians. *Journal of Ethnopharmacology*. 137(3), 1228–1239. <https://doi.org/10.1016/j.jep.2011.07.044>
- Oyedemi, S.O., Oyedemi, B.O., Ijeh, I.I., Ohanyerem, P.E., Cooposamy, R.M., Aiyegoro, O.A., 2017. Alpha-Amylase Inhibition and Antioxidative Capacity of Some Antidiabetic Plants Used by the Traditional Healers in Southeastern Nigeria. *Scientific World Journal*, 3592491. <https://doi.org/10.1155/2017/3592491>
- Parhan, N.N., 2021. Antipyretic Effect of Ethanol Extract of *Randu* (*Ceiba pentandra* (L.) Gaertn.) Leaves on Male White Rat (*Rattus norvegicus*). *Jurnal Dunia Farmasi*. 5(2), 82–88. <https://doi.org/10.33085/jdf.v5i>
- Pereira Júnior, L.R., Andrade, A.P., Araújo, K.D., Barbosa, A.S., Barbosa, F.M., 2014. Espécies da caatinga como alternativa para o desenvolvimento de novos fitofármacos. *Floresta e Ambiente*. 21(4), 509–520. <https://doi.org/10.1590/2179-8087.024212>
- Pezzini, F.F., Dexter, K.G., Carvalho-Sobrinho, J.G., Kidner, C.A., Nicholls, J.A., Queiroz, L.P., Pennington, R.T., 2021. Phylogeny and biogeography of *Ceiba* Mill. (Malvaceae, Bombacoideae). *Frontiers of Biogeography*. 13(2), e49226. <https://doi.org/10.21425/f5fbg49226>
- Rao, K.V., Sreeramulu, K., Gunasekar, D., 1993. Two New Sesquiterpene Lactones From *Ceiba pentandra*. *Journal of Natural Products*. 56(12), 2041–2045. <https://doi.org/10.1021/np50102a003>
- Ravi Kiran, C., Raghava Rao, T., 2014. Lipid Profiling by GC-MS and Anti-inflammatory Activities of *Ceiba pentandra* Seed Oil. *Journal of Biologically Active Products from Nature*. 4(1), 62–70. <https://doi.org/10.1080/22311866.2014.890064>
- REFLORA., 2022. Flora e Funga do Brasil. <http://reflora.jbrj.gov.br> Date accessed: 19th July 2022.
- Ribeiro, D.A., Oliveira, L.G.S., Macêdo, D.G., Menezes, I.R.A., Costa, J.G.M., Silva, M.A.P., Lacerda, S.R., Souza, M.M.A., 2014. Promising medicinal plants for bioprospection in a Cerrado area of Chapada do Araripe, Northeastern Brazil. *Journal of Ethnopharmacology*. 155(3), 1522–1533. <https://doi.org/10.1016/j.jep.2014.07.042>
- Ribeiro, R.V., Bieski, I.G.C., Balogun, S.O., Martins, D.T.O., 2017. Ethnobotanical study of medicinal plants used by Ribeirinhos in the North Araguaia microregion. *Journal of Ethnopharmacology*. 205(9), 69–102. <https://doi.org/10.1016/j.jep.2017.04.023>
- Roosita, K., Kusharto, C.M., Sekiyama, M., Fachrurrozi, Y., Ohtsuka, R., 2008. Medicinal plants used by the villagers of a Sundanese community in West Java, Indonesia. *Journal of Ethnopharmacology*. 115(1), 72–81. <https://doi.org/10.1016/j.jep.2007.09.010>
- Rosselli, S., Tundis, R., Bruno, M., Leporini, M., Falco, T., Candela, R.G., Badalamenti, N., Loizzo, M.R., 2020. *Ceiba speciosa* (A. St.-Hil.) seeds oil: Fatty acids profiling by GC-MS and NMR and bioactivity. *Molecules*. 25(5), 1037–1037. <https://doi.org/10.3390/molecules25051037>
- Santos, F.B., Quines, C.B., Pilissão, L.E., Dal Forno, A.H.C., Rodrigues, C.F., Denardin, C.C., Farias, F.M., Avila, D.S., 2020. Aqueous Bark Extract of *Ceiba speciosa* (A. St.-Hill) *Ravenna* Protects against Glucose Toxicity in *Caenorhabditis elegans*. *Oxidative Medicine and Cellular Longevity*. <https://doi.org/10.1155/2020/1321354>
- Santos, R.I., Jacinto, W.R., Cruz, K.G.J., 2021. Philippine Ethnobotanicals Downregulate *lasR* Expression Linked to Quorum Sensing-Mediated Biofilm Formation In *Pseudomonas aeruginosa*. *Biotechnology and Food Sciences*. 10(4), 592–597. <https://doi.org/10.15414/jmbfs.2021.10.4.592-597>
- Sanz-Biset, J., de-la Cruz, J.C., Epiquién-Rivera, M.A., Cañigual, S., 2009. A first survey on the medicinal plants of the Chazuta valley (Peruvian Amazon). *Journal of Ethnopharmacology*. 122(2), 333–362. <https://doi.org/10.1016/j.jep.2008.12.009>
- Saptarini, N.M., Deswati, D.A., 2015. The antipyretic activity of leaves extract of *Ceiba pentandra* better than *Gossypium arboreum*. *Journal of Applied Pharmaceutical Science*. 5(7), 118–121. <https://doi.org/10.7324/JAPS.2015.50718>
- Saraiva, M.E., Ulisses, A.V.R.A., Ribeiro, D.A., Oliveira, L.G.S., Macêdo, D.G., Sousa, F.F.S., Menezes, I.R.A., Sampaio, E.V.S.B., Souza, M.M.A., 2015. Plant species as a therapeutic resource in areas of the savanna in the state of Pernambuco, Northeast Brazil. *Journal of Ethnopharmacology*. 171(2), 141–153. <https://doi.org/10.1016/j.jep.2015.05.034>
- Schmeda-Hirschmann, G., 1993. Magic and medicinal plants of the Ayoreos of the Chaco Boreal (Paraguay). *Journal of Ethnopharmacology*. 39(2), 90025–90025. [https://doi.org/10.1016/0378-8741\(93\)90025-Z](https://doi.org/10.1016/0378-8741(93)90025-Z)
- Shivhare, Y., Singh, P., Singh, S., Bharti, P.K., Tiwari, R., 2011. Ethnomedicinal Plants for the Prevention and Treatment of Gonorrhoea. *Research Journal Pharmacy and Technology*. 4(2), 182–183.
- SiBBr., 2022. Sistema de Informação sobre a Biodiversidade Brasileira. Global Diversity Information facility. <https://sibbr.gov.br>. Date

accessed: 14th october 2022.

- Silva, C.G., Marinho, M.G.V., Lucena, M.F.A., Costa, J.G.M., 2015. Levantamento etnobotânico de plantas medicinais em área de Caatinga na comunidade do Sítio Nazaré, município de Milagres. *Revista Brasileira de Plantas Medicinais*. 17(1), 133–142. https://doi.org/10.1590/1983-084X/12_055
- Smith, G.C., Dueker, S.R., Clifford, A.J., Grivetti, L.E., 1996. Carotenoid values of selected plant foods common to Southern Burkina Faso. West Africa. *Ecology of Food and Nutrition*. 35(1), 43–58. <https://doi.org/10.1080/03670244.1996.9991474>
- Sobrinho, A.C.N., Nunes, J.A., Souza, R.U., Lucena, L.S., Silva, F.B.G., Figueiredo, D., 2021. Estudo etnobotânico de plantas medicinais comercializadas no mercado público de Iguatu-Ceará. *Brasil. Research, Society and Development*. 10(6), e14310615478. <https://doi.org/10.33448/rsd-v10i6.15478>
- Syihabudin, V., Sari, B.L., Utami, N.F., Apriliani, N.A., 2018. Inhibitory activity of α -glucosidase of bark of *Ceiba pentandra* Linn. *Indonesian Journal of Pharmacy*. 29(4), 206–213. <https://doi.org/10.14499/indonesianjpharm29iss4pp206>
- Tapsoba, H., Deschamps, J.P., 2006. Use of medicinal plants for the treatment of oral diseases in Burkina Faso. *Journal of Ethnopharmacology*. 104(1-2), 68–78. <https://doi.org/10.1016/j.jep.2005.08.047>
- Tripathi, S., Farooqui, A., Singh, V.K., Singh, S., Kumar Roy, R., 2019. Morphometrical analysis of *Ceiba* Mill. (Bombacoideae, Malvaceae) pollen: a sacred plant of the Mayan (Mesoamerican) civilisation. *Palynology*. 43(4), 551–573. <https://doi.org/10.1080/01916122.2018.1467350>
- Ueda, H., Kaneda, N., Kawanishi, K., Alves, S.M., Moriyasu, M., 2002. A New Isoflavone Glycoside from *Ceiba pentandra* (L.) GAERTNER. *Chemical and Pharmaceutical Bulletin*. 50(3), 403–404. <https://doi.org/10.1248/cpb.50.403>
- Wahab Obeng, A., Boakye, Y.D., Agana, T.A., Boamah, V.E., Oppong-Kyekyeku, J., Brobbey, A.A., Adu, F., Gbedema, S.Y., Agyare, C., 2022. Antitrypanosomal and Anthelmintic Properties of Ethanol Extracts of *Carica papaya* Linn. and *Ceiba pentandra* (L) Gaertn. *Journal of Chemistry*, 5251930. <https://doi.org/10.1155/2022/5251930>
- Wodah, D., Asase, A., 2012. Ethnopharmacological use of plants by Sisala traditional healers in northwest Ghana. *Pharmaceutical Biology*. 50(7), 807–815. <https://doi.org/10.3109/13880209.2011.633920>
- Zheng, X., Xing, F., 2009. Ethnobotanical study on medicinal plants around Mt.Yinggeling, Hainan Island. China. *Journal of Ethnopharmacology*. 124(2), 197–210. <https://doi.org/10.1016/j.jep.2009.04.042>