# Natural Resources for Human Health



#### Review

#### View Article Online

() Check for updates

Received 23 September 2022 Revised 03 November 2022 Accepted 03 November 2022 Available online 10 November 2022

Edited by Rohit Sharma

#### **KEYWORDS:**

Grape polyphenols hepatoprotection hepatotoxicity hepatotoxic agents

Natr Resour Human Health 2023; 3 (2): 196-213 https://doi.org/10.53365/nrfhh/156261 eISSN: 2583-1194 Copyright © 2023 Visagaa Publishing House

# A systematic review on hepatoprotective potential of grape and polyphenolic compounds: molecular mechanism and future prospective

Shabina Khan<sup>1</sup>, Mohammad Ibrahim<sup>2,\*</sup>

<sup>1</sup>Department of Pharmacy, Anjuman-I-Islam Kalshekhar Technical Campus, Mumbai - 410206, Maharashtra, India

<sup>2</sup>Pharmacology, Research and Development, Abhinav Healthcare Products Private Limited, Mumbai - 400053, Maharashtra, India

ABSTRACT: The hepatoprotective potential of grapes and polyphenols has been scientifically well demonstrated in several experimental studies. However, there is no systematic review performed on their hepatoprotective activity. Based on scientific evidence, the goal of this systematic review was to describe and make inferences on the therapeutic advantages of grape and polyphenols against various chemical hepatotoxicities. Grape and polyphenols from preclinical and clinical studies were included for the online search utilising electronic databases such as PubMed, Web of Science, Scopus, Springer Nature, and Google Scholar. The searched words and terms were: "Hepatotoxicity" OR "Hepatoprotective" OR "Liver injury" AND "Hepatotoxic drug" AND "Grape" OR "Grapefruit" OR "Grape seed" OR "Grape peels" OR "Bioactive compounds" OR "Polyphenols" AND "Preclinical studies" OR "Clinical studies". A total of sixty-five studies were included based on the rigorous inclusion and exclusion criteria. The systematic review revealed that grape and polyphenolic compounds could significantly alleviate hepatic markers (ALP, ALT and AST), antioxidant status (CAT, GPx, GSH and SOD), hepatic anti-inflammatory status (TNF- $\alpha$ , IL-1, IL-6, COX-2, and NF- $\kappa$ B pathways) and apoptosis markers [caspase-3 (CASP) and caspase-9] in chemical induced-hepatotoxicity. Furthermore, improvements in the structural, functional, and physiological characteristics of the liver were observed after treatment with grape and polyphenols, reflecting modulation in the cellular and molecular alterations. Conclusively, this study supports the therapeutic potential of grape and polyphenols for the management of hepatotoxicity in various conditions based on available scientific evidence.

#### 1. INTRODUCTION

The liver is the largest and most important vital organ of the human body. It plays a crucial role in the management of many physiological processes associated with several essential processes like metabolism, secretion, and storage. It has a propensity to detoxify endogenous (waste metabolites) and exogenous (toxic chemicals) compounds of the organisms (Mani et al., 2022). In addition, metabolic processes of development, nutrient provision, energy production, reproduction, bile secretion, vitamin storage, and the metabolism of fats and carbohydrates are also carried out by the liver. Because of all, hepatic disorders are still one of the biggest health challenges and these are serious public issues around the globe (Hajdarevic et al., 2020).

The liver is negatively affected by biological factors, autoimmune, toxic chemicals [carbon tetrachloride (CCl<sub>4</sub>), aluminium chloride (AlCl<sub>3</sub>)], drugs [paracetamol (PCM), thioacetamide, dimethylnitrosamine (DMN), diclofenac, isoniazid, valproic acid, d-galactosamine/lipopolysaccharide (GalN/LPS)], infections, disturbances in the blood supply and unquestionably, excessive consumption of alcohol (Singh et al., 2016). Excessive exposure to toxins and dosage of pharmaceuticals results in altered homeostatic status that leads to reactive oxygen species (ROS) generation. Altered ROS generation increases lipid peroxidation, oxidative stress and inflammations leading to organ malfunction that resulting hepatic steatosis, hepatitis, cirrhosis, haemochromatosis, cholestatic and hepatocellular carcinoma (Singh et al., 2016).

Drug-induced liver injury (DILI) and herb-induced liver injury (HILI) are well-known concerns that can symptomatically reflect both acute and chronic liver disorders. The clinical manifestations of DILI are similar to HILI. It occurs due to exposure to chemical drugs either produced naturally or synthesized in the laboratory. Annually, out of 10.000-100.000 people about 10-15 people suffered from hepatotoxicity after



\* Corresponding author.

*E-mail address:* ibrahimhayat91@gmail.com (Mohammad Ibrahim)

This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

taking prescripted or OTC medicines (Raschi & De Ponti, 2015).

Despite significant advances in modern medicine, there are no entirely effective drugs that improve liver function, offer complete protection for the organ, or facilitate the regeneration of hepatic cells. Although the majority of synthetic drugs for liver diseases are effective pro-oxidant scavengers, their prolonged use could have substantial adverse effects. (Farzaei et al., 2018). Taking this into consideration, researchers are inclined to employ plant-based drugs because they will have greater efficacy, be more cost-effective, and have little to no toxicity. Therefore, there has long-established empirical evidence for using herbal treatments to alleviate hepatic problems; grapefruit and its polyphenols are one of them (Ibrahim et al., 2022). Cited literature reported that grape (fruit/seed/peels) contains a huge number of polyphenols that are capable of significantly modulating the levels of alanine phosphatase and transaminase (ALP and ALT), aspartate transaminase (AST), thiobarbituric acid-reactive substances (TBARS), malondialdehyde (MDA), lipid peroxides (LPO), protein carbonyl content, total bilirubin (TLB), lactate dehydrogenase (LDH), and enzymes increasing the activity of catalase (CAT), glutathione peroxidase (GPx) glutathione (GSH), superoxide dismutase (SOD), alcohol dehydrogenase (ADH), and aldehyde dehydrogenase (ALDH), as well as the levels of vitamins C and E in the hepatocytes (Madrigal-Santillán et al., 2014; Wiegand & Berg, 2013).

In the current scenario, numerous studies have been reported on the hepatoprotective potential of grapes and polyphenols. However, we have presented a comprehensive systematic analysis of grapefruit/seed/peel and polyphenolic compounds based on rigorous inclusion and exclusion criteria for their hepatoprotective potential.

#### 2. METHODS

#### 2.1. Search Strategy

According to PRISMA guidelines, a systematic review was done to find the information available for grapes and polyphenols for hepatoprotective activity. The following keywords were used: "Hepatotoxicity" OR "Hepatoprotective" OR "Liver injury" AND "Hepatotoxic drug" AND "Grape" OR "Grapefruit" OR "Grape seed" OR "Grape peels" OR "Bioactive compounds" OR "Polyphenols" AND "Preclinical studies" OR "Clinical studies".

#### 2.2. Study Selection

PubMed, Google Scholar, Springer, Scopus, Taylor & Francis, and Science Direct were searched for studies published online without language and time restrictions. We have limited our search to focus on the hepatoprotective potential of grapefruit/seeds/peels and bioactive compounds especially polyphenols, whereby only online research papers having complete studies are considered. Based on rigorous inclusion and exclusion criteria total of 65 studies were included in the current study and Mendeley was used as a database manager

# (Figure 1).

### 3. RESULTS

Figure 1 summarizes the research selection procedure as per PRISMA principles. Using the online database search, a total of 184 records were retrieved. After 79 duplicate publications were excluded, 105 papers were included, and further 27 papers were rejected based on the full text, title and/or abstract. Moreover, 13 papers were removed for not meeting the inclusion criteria of the eligible studies (n = 78) read. Finally, 65 papers were included after the selection procedure was complete.

# 3.1. Rationale for the use of grape and polyphenols for the management of hepatotoxicity

For thousands of years, plants have been used as sources of medicine, grape is one of them. Grapefruit/seeds/peels contain a variety of polyphenols that could serve as the foundation for the development of future medicine. These may be used for hepatoprotection either alone or in combination with synthetic medicine, making them an attractive target for the future (Nada et al., 2015; Saha et al., 2019). By putting forth this effort, existing liver disease medicines may be given a wider range of potential applications and it makes sense that liver disease treatments will be cured using grapes in their entirety or their bioactive components. Because of their natural origin, cultural acceptability, minimal or no side effects, affordability, and accessibility, plant-derived medicines have been utilized for the management of various diseases for a long time, with reportedly inconclusive findings for improving outcomes in developed and developing countries. Many medicines derived from plant origin to manage various diseases and disorders including atropine, aspirin, artemisinin, resveratrol, ephedrine, digoxin, reserpine, quinine, cannabidiol, vinblastine, vincristine, dronabinol, and many more (Amedei & Niccolai, 2014). Hence, to replace currently prescribed drugs with dubious efficacy and safety, we need to look for novel, inexpensive, and effective plant-based therapeutics for the treatment of liver illnesses.

#### 3.2. Mechanisms of conventional drug-induced hepatotoxicity

The liver lesion is indicated by hepatocytes damage, which manifests by elevation in ALT, AST and cholestatic lesion when ALP increases twice than actual. Hepatocytes, cholangiocytes, ductal cells, Kupffer cells and endothelial cells are all involved in the processes by which drugs cause hepatotoxicity; they also directly affect cellular organelles such as the endoplasmic reticulum, mitochondria, cytoskeleton, microtubules, or nucleus (Guicciardi et al., 2013; Morales et al., 2016; Xu et al., 2020). Moreover, the development of drug metabolites in the liver through biotransformation may result in hepatic injury due to toxic chemicals or free radicles compounds and may trigger several chemical reactions (Xu et al., 2020). These pathways may cause apoptosis, necrosis, or both. The primary culprits of liver damage include the following mechanism mentioned below and examples of some conventional drug-induced hepatotoxicity



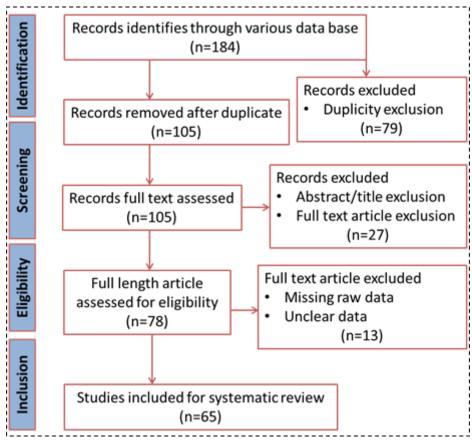


Figure 1. Schematic representation of the systematic review.

#### (Table 1).

Mitochondrial dysfunction: It may occur due to the disruption of  $\beta$ -oxidation of lipids and oxidative energy production within the hepatic cells. In addition, steatosis may exist as a result of aberrant mitochondrial activity and fat accumulation, which can also result in hepatic apoptosis, ATP depletion and consequent necrosis, and mitochondrial membrane permeabilization or rupture.

Immune response: The creation of new antigens is believed to be responsible for the immune response, resulting cause idiosyncratic hepatotoxicity. Additionally, it may be characterized by inflammatory cells like neutrophils and lymphocytes.

Oxidative stress (OS): The liver is the most frequent target organ for drug toxicity. It has been suggested that the production of free radical species, specifically ROS and RNS, is an early sign of drug toxicity and a sign of its propensity to be harmful to the liver. Additionally, OS is generated when ATP is depleted and the concentration of calcium inside hepatic cells increases, which may result in liver necrosis.

Lipid peroxidation: Free radicals and fatty acids in the membranes interact to cause lipid peroxidation, which might result in the formation of electrophilic metabolites that can damage cellular DNA.

#### 3.3. Clinical manifestations

In both acute and chronic use, several medications cause liver damage. DILI can relate to either asymptomatic liver injury or liver failure which is clinically severe. However, the most reliable sign of a drug's propensity for severe hepatotoxicity is an increase in blood TBL and aminotransferase levels. The liver enzymes that are most frequently upregulated by the use of modern medicines without any clinical signs are AST followed by ALT and ALP (Figure 2) (Bessone et al., 2021). The identification of a higher rate of these elevations in drugusing participants compared to a placebo group is a sensitive signal of the potential to generate severe DILI. According to the published reports, a higher rate of the more pronounced peak in ALT elevations (10x, 15x upper limit of normal-ULN), with cases of rises to >1,000 U/L raises more worries and is considered a definite indicator of hepatotoxicity (Devarbhavi, 2012).

Another potentially helpful indicator of liver function that might point to DILI is an elevated plasma prothrombin (PT) time or its international normalised ratio as a result of decreased hepatic synthesis of vitamin K-dependent clotting components. However, the occurrence of clinically severe liver injury or impaired liver function (with prolonged PT or hypoalbuminemia) that leads to deficient protein synthesis is rare (Figure 2). Jaundice, gastrointestinal discomfort, itching,



# Table 1

Some conventional drugs induce hepatotoxicity

Drug	Possible Mechanism of hepatotoxicity	References
Acetaminophen	Through the CYP450 system, it is converted to hazardous intermediates. High acetaminophen doses lower GSH levels or stop the pathway; this intermediate accumulates and binds to intracellular macromolecules that can cause cell damage, typically by caspase-independent apoptosis started by PARP-1 activation.	Yoon et al. (2016)
Aspirin	Direct intrinsic hepatotoxic. Inhibit mitochondrial function, elevates serum ALT levels and mild bilirubin	Laster and Satoskar (2014)
Dabigatran	Hepatic injury due to marked elevation in enzymes in the liver	Rochwerg et al. (2012)
Glipizide	Liver damage may be exacerbated by hypersensitivity. Hepatocellular, cholestatic, and mixed injuries have all been linked to sulfonylurea-induced liver damage.	Kamal and Bhabhra (2019)
Interferon beta (ß-1a and ß-1b)	Chronic hepatitis is suggested by persistent ALT increases.	Liao et al. (2013)
Isoniazid, rifampicin, pyrazinamide	Lipid peroxidation (necrosis). Formation of hepatotoxic metabolites	Yew and Leung (2006)
Ketoconazole	Potentially inhibited human CYP 3A4, which can alter the serum levels of many drugs that are metabolized via the P450 system and ultimately increase liver toxicity.	Yan et al. (2013)
Lovastatin	Lovastatin is largely metabolized by CYP 3A4 and metabolites are excreted in bile. The mild ALT elevations are likely due to a toxic metabolite. The onset of clinical injury (usually cholestatic, but can be hepatocellular) can vary from weeks to years.	Perdices et al. (2014)
Methimazole	The liver injury occurs due to an increased level of serum ALT	Gomez-Peralta et al. (2018)
Methotrexate	Direct toxicity. Lipid peroxidation (necrosis). Dysfunction of mitochondrial and increase in liver enzymes	Conway and Carey (2017)
Methylphenidate	It is extensively metabolized in the liver and has many drug-drug interactions as well as increases serum aminotransferase level	Alam and Ikram (2018)
Olanzapine	Olanzapine has extensive hepatic metabolism, partially by CYP450 enzymes and some cases of clinically apparent hepatotoxicity may be due to the production of a toxic metabolite and increased level of serum ALT	Dönmez et al. (2017)
Orlistat	Hypersensitivity is likely to be the cause of liver injury	Douglas et al. (2013)
Phenytoin	DILI is caused by phenytoin due to a hypersensitivity reaction, mostly in typical cases of immunoallergic hepatotoxicity. It is metabolized by the CYP450 system to arene oxide, which may result in toxic or immunogenic metabolite formation.	Ganesan et al. (2015)
Rifampin (Rifampicin)	The cause of injury is likely to be due to toxic metabolic products that directly induce an immunologic reaction.	Fountain et al. (2009)
Tacrolimus	Liver injury related to serum enzyme elevation.	Ko et al. (2015)
Tamoxifen	Disrupt $\beta$ -oxidation of lipids. Steatohepatitis, necrosis, and inflammatory infiltrate due to higher levels of ALT	Gao et al. (2016)
Valproic acid	It lowers tissue carnitine levels, which can lead to inhibition of beta-oxidation and loss of mitochondrial function, hyperammonemia and microvesicular steatosis.	Gayam et al. (2018)
Vancomycin	Direct toxicity or immune adverse reactions	Brunetti et al. (2020)

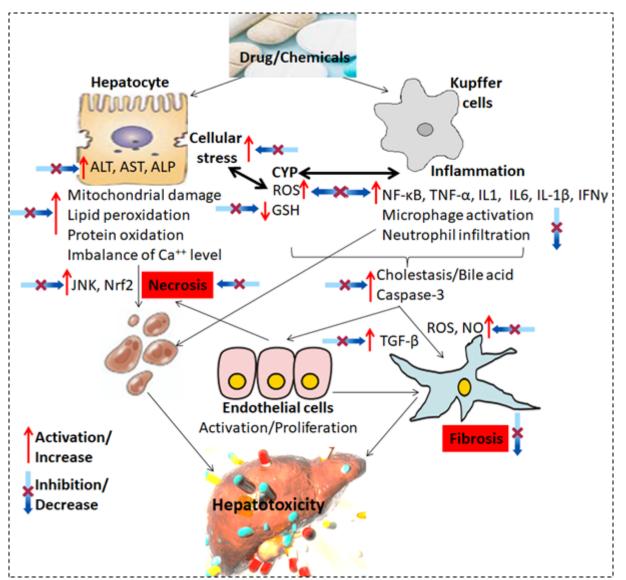
and other symptoms are among them (Fernandez et al., 2021). In addition to medications, DILI also covers injuries caused by dietary supplements, herbal remedies, and medicinal plants.

#### 3.4. Therapeutic potential of grape and polyphenols

Diets are essential for improving health care and preventing disease and disorders because they contain a huge number of bioactive molecules. Grapes are well known for their useful remedial functions as they are rich in polyphenols and have beneficial health effects. Some important polyphenols found in grapes like quercetin, gallic acid, catechin, syringic acid, naringenin and many more play a key role to protect liver disorder, oxidative stress, inflammation, brain disorders, diabesity, cardiovascular disease and disorders etc., (S. Li et al., 2018; Nada et al., 2015; Saha et al., 2019).

The hepatoprotective mechanism of grapes and polyphenolic compounds is assumed to be an elevation in antioxidant enzyme activities, down-regulation of CYP2E1, iNOS, CASP-3, and NF-kB expression, up-regulation of trace element concentration levels, and activation of SIRT1 gene expression, which results in an improvement of the antioxidant and anti-inflammatory status in the hepatic tissues (Ismail et al., 2016). A previously published report suggested that ethanolic extract of grape impressively inhibited the level of serum ALP and TLB, whereas resveratrol significantly reduced oxidative stress, hepatic inflammation (IL-1, IL-6, NF-





**Figure 2.** Diagrammatic presentation of hepatotoxicity in response to conventional drugs and protective effect of grape and polyphenols. Kupffer and endothelial cells are activated by harmfulchemicals and chemotactic molecules secreted by injured hepatocytes. These occurrences could lead to hepatocyte stress, which is exacerbated by the presence of ROS and NO species produced by active endothelial cells. Hepatocyte death and Kupffer cell fibrosis enhancement are the end results. Nf-kB: IL-1: Interleukin-1; IL-6: Interleukin-6; I-1 $\beta$ : Interleukin 1beta; IFN $\gamma$ :Interferon-gamma; JNK: nuclear factor erythroid 2–related factor 2: Nrf2; NO:nitrogen oxides; TGF- $\beta$ : Transforming growth factor beta.

 $k\beta$ , TNF- $\alpha$ , and IKK), and apoptosis markers CASP-3, CASP-9, and Bax] in streptozotocin-induced diabetic rats (Giribabu et al., 2018). Moreover, grape polyphenols reduce severe inflammation either by regulating inflammatory pathways or by reducing ROS levels. Grape flavonoid derivatives may a better remedial choice than synthetic ones because they are natural constituents that can choose numerous paths to reduce chronic inflammation (Haghighatdoost et al., 2020). According to another study, grape seed extract controls LDH leakage from liver cells while maintaining antioxidant enzyme levels. The enrichment of bioactive substances such as polyphenols, vitamin E, and ascorbic acid may be responsible for these actions (Dogan & Celik, 2012). Although several polyphenols are cited in the literature, few of them are described. Caffeic acid exhibited potential anti-inflammatory and antioxidant qualities and is successful in the treatment of severe liver damage. By engaging with the Nrf2 binding site, inhibiting it from binding to the liver cancer factor kelch-like ECH-associated protein-1 (Keap1), and increasing the expression of key antioxidative signals like HO-1, it can alter Keap1's expression (Yang et al., 2017). Quercetin scavenges ROS, and inhibits xanthine oxidase as well as lipid peroxidation, in addition to scavenging and stabilizing iron. It also reduces the level of ALT and AST which will help to minimise liver damage (Barros et al., 2017).



# Table 2

Grape and polyphenolic compounds used for the management of hepatotoxicity against hepatotoxic drugs and chemicals

Constituents	Dose	Mechanism	Inducing agents	References
Grapefruit/seed/peel	27ml/kg/b.w./rat	Attenuates the level of ALT and AST and protects against liver damage.	AlCl <sub>3</sub>	Al-Amoudi (2017)
	200mg/kg/b.w./rat	Ameliorates the activity of MDA, GSH, and SOD activity in liver cells and protects against cell injury	LPS/ d-GalN	Gad et al. (2013)
	200-600 mg/kg/b.w./rat	Reduced the activities of ALP, ALT and AST	Paracetamol	Udom et al. (2018)
	Conventional/organic juice	Modulates CAT, SOD and GPx activities. Also, inhibit hepatocellular degeneration and steatosis	High fat diet	Buchner et al. (2014)
	100 mg/kg/b.w./mice	Diminished elevated levels of ALT, AST, and MDA while enhancing GSH, SOD and CAT levels. Protect infiltration, irregular nuclei and altered mitochondria in live tissue	Ehrlich solid tumor	M. Li et al. (2015)
	100-200 mg/kg/b.w./rat	Significantly reduced MDA, NO contents in liver homogenates and AST, ALT, ALP, and TLB levels in serum. While significantly increased serum GSH content in liver homogenate. Attenuates disruption of the hepatic architecture and collagen fibres deposition in the portal tract	Thioacetamide	Nada et al. (2015)
	100 mg/kg/b.w./rat	Significantly decrease MDA level and efficiently increases GSH level and catalase CAT, SOD activity. Attenuated liver injury to normal	Diethylnitrosamine	Bilgic et al. (2018)
	150 mg/kg/b.w./rat	Decreased AST, ALT and LDH serum enzyme levels and MDA contents in liver tissues	Ethanol	Dogan and Celik (2012)



		<i>Table 2 continued</i>		
Gallic acid	50 mg/kg/b.w./rat	It increases antioxidant enzyme (SPD, CAT) activities, decreased plasma hepatic enzyme (ALT, ALP, AST) and pro-inflammatory markers (IL-1 $\beta$ , IL-6, COX-2, TNF- $\alpha$ )	CCl <sub>4</sub>	Ojeaburu and Oriakhi (2021)
	100 mg/kg/b.w./mice	Modulates AST, ALT, ALP, lipid peroxidation and TNF- $\alpha$ levels in serum and antioxidant status (OSD, CAT and GSH)	РСМ	Rasool et al. (2010)
	20 mg/kg/b.w./rat	Diminished hepatic damage via upregulated hepatic mRNA expression of GLUT-4, Wnt1 and $\beta$ -catenin with inhibitory effects on the elevated expression of ERK1/2/NF- $\kappa$ B	STZ	Bashar et al. (2021)
	100 mg/kg/b.w./rat	Attenuating the inflammatory mediators COX-2 through the NF- $\kappa$ B inhibition pathway increases SOD and ATPases (Ca <sup>2+/</sup> Mg <sup>2+)</sup> expression	N'- Nitrosodiethylamine	Latief et al. (2016)
	10-20 mg/kg/b.w./rat	Decreased level of TBARS and MDA in liver homogenates. Improve antioxidant status and liver injury.	NaF	Nabavi et al. (2013)
Catechin	50-100 mg/kg/b.w./rat	Prevented mitochondrial membrane damage, up-regulation of p53, Bax and down-regulation of Bcl-2 mRNA levels in the liver. Protect hepatic cell damage.	d-GalN	Raj et al. (2010)
	100-400 mg/kg/b.w./mice	Increased SOD, CAT, GX, and GSH levels and markedly decreased the MDA level. Protect liver cell damage	CCl <sub>4</sub>	Liu et al. (2015)
Epicatechin	50-100 mg/kg/b.w./mice	Decrease the level of ALT, AST, LPO, IL1 $\beta$ , TNF- $\alpha$ , and NO while increasing CAT, SOD and GSH levels. Reduced histological damage.	MTX	Azadnasab et al. (2021)



		Table 2 continued		
	20 mg/kg/b.w./rat	Declined MDA level whereas GSH, CAT, GST, GPx, and CYP450 levels were upregulated. Regenerated damage of hepatic tissue.	CCl <sub>4</sub>	Alkinani et al. (2021)
Quercetin	50-100 mg/kg/b.w./rat	Reduction of ALT, AST and amelioration of liver damages.	РСМ	Alkinani et al. (2021)
	10 mg/kg/b.w./rat	Protect the liver against DNA damage in mild liver injury	ACR	Ansar et al. (2016)
	70 mg/kg/b.w./rat	Enhance the level of SOD, and GPx while decreasing AST, ALT, LPO, direct bilirubin, and TLB level and also protect tissue damage	ACR	Abo-Salem et al. (2011)
	20 mg/kg/b.w./rat	Decreased level of MDA and increased GSH and CAT activities. Protect liver injury	CCl <sub>4</sub>	Huang et al. (2018)
	50 mg/kg/b.w./rat	Increases level of SOD, CAT, GPx, and GSH while decreasing MDA level.	РСМ	Faras and Elsawaf (2017)
	50mg/kg/b.w./mice	Attenuated the NF- $\kappa$ B, TNF- $\alpha$ , COX-2, TGF- $\beta$ 1 and expression of iNOS	$\mathrm{CCl}_4$	Domitrović et al. (2012)
Kaempfrol	20mg/kg/b.w./mice	Decreased AST, ALT, LPO, TG, and MDA levels in serum and increases the level of SOD, and GSH. Reduced the expression level and activity of hepatic CYP2E1	Alcohol	Wang et al. (2015)
	50mg/kg/b.w./mice	Reduced ALT, AST activities, and increased GSH, SOD, GPx, CAT and ameliorates liver injury	CCl <sub>4</sub>	Zang et al. (2017)
	250 mg/kg/b.w./rat	Increased GSH, Bcl-2 levels while decreasing the level of MDA, ROS, TNF- $\alpha$ , Bax. It also reduced the expression of CYP2E1, and the activity and protein levels of SIRT1.	АРАР	Binmowyna and Alfaris (2021)



		<i>Table 2 continued</i>		
Rutin	20 mg/kg/b.w./rat	Reduced ALT, MDA and total bilirubin while increasing GSH, SOD, GPx and CAT levels. Attenuates liver cell damage through uniform mitochondrial distribution, regeneration of rough ER, and vacuolations in the cytoplasm	АРАР	Reddy et al. (2017)
	40 mg/kg/b.w./rat	Decreased level of MDA, ALP, AST and ALT while increasing SOD and CAT. Attenuates infiltration of hepatic tissue	Isoniazid	Abdel-Ghaffar et al. (2017)
	10-250 mg/kg/b.w./rat	Attenuated the NF- $\kappa$ B, TNF- $\alpha$ , COX-2, TGF- $\beta$ 1 and expression of iNOS	$\mathrm{CCl}_4$	Domitrović et al. (2012)
	70 mg/kg/b.w./rat	Downregulates expression levels of IL-6, Bcl-XL, MEK5, FADD, EGF, STAT3 and JAK while upregulates the Bcl2 expression	CCl <sub>4</sub>	Hafez et al. (2015)
	50 mg/kg/b.w./rat	Decreased level of AST, ALT, LDH, ALP and CASP-3 while increasing GPx and GSH	5-FU	Gelen et al. (2017)
	40 mg/kg/b.w./rat	Decreased ALT activity, TLB, TNF- $\alpha$ ), LPO, NF $\kappa$ B and Increased hepatic GSH level.	d-GalN	Mahmoud et al. (2013)
Chlorogenic acid	60 mg/kg/b.w./rat	Inhibited the activities of mitochondrial NADH dehydrogenase and ATP synthase and the contents of ATP, and AMP in the liver and protect the liver	LPS	Z. Chen et al. (2019)
	50mg/kg/b.w./mice	Decreased the elevated plasma levels of ALT and AST and also suppressed hepatic mRNA expression of toll-like receptor 4 (TLR4), TNF- $\alpha$ and NF-k $\beta$ p65 subunit	LPS	Xu et al. (2010)



		Table 2 continued		
	60-200 mg/kg/b.w./mice	Decrease the serum levels of AST, ALT, TC, TG, and total albumin and elevated the serum total albumin and the activities of GSH, CAT, SOD, GSH-Rd, and GSH-Px increased. Decreased the formation of lesions in liver	CCl <sub>4</sub>	Hsu et al. (2022)
		Protect liver injury markers (AST, ALT and ALP), hepatic inflammatory cytokines (IL-1, IL-6, TNF- $\alpha$ and TNF- $\beta$ ) and dyslipidemia (TC, TG and LDL-C)	High l-carnitine	X. Zhang et al. (2021
	100 mg/kg/b.w./mice	Significantly reverse in LPO, enzymatic leakage, Cyt P450 inactivation and produced enhancement of cellular antioxidant defence		Kapil et al. (1995)
Resveratrol	200 mg/kg/b.w./mice	Increased SOD protein and mRNA expression levels in hepatic cells	Ethanol	W.M. Chen et al. (2016)
	20 mg/kg/b.w./rat	Ameliorates the level of AST and ALT, MDA while increasing the activities of SOD, CAT, GPx and protects against serious congestion and haemorrhage related to the hepatic tissues injury	Cadmium	Al-Baqami and Hamza (2021)
Syringic acid	10mg/kg/b.w./mice	Attenuates the level of AST, and ALT and protects liver fibrogenesis	$\mathrm{CCl}_4$	Itoh et al. (2010)
	50-100 mg/kg/b.w./rat	Effectively reduced AST, ALT, ALP, LDH, MDA, and ROS while increasing SOD and GSH levels. Attenuated inflammatory injury by suppressing TNF- $\alpha$ , IL-1 $\beta$ and NF- $\kappa$ B and also down-regulates CASP-3 expression in the liver	Thioacetamide	Okkay et al. (2022)
Caftric acid	40 mg/kg/b.w./rat	Decreased the level of AST, ALT, ALP, and MDA while increasing SOD, Cat and GPx	Methamphetamine	Koriem and Soliman (2014)



#### Khan and Ibrahim

		<i>Table 2 continued</i>		
Caffeic acid	5 mg/kg/b.w./rat	Increases endogenous level of GSH, GPx, SOD, and CAT	Mercuric chloride	Manju and Jagadeesa (2019)
	3-12 mg/kg/b.w./rat	Markedly decreased MDA levels while increased GSH, CAT and SOD activities in liver	CCl <sub>4</sub>	M. Li et al. (2015)
Ellagic acid	50-100 mg/kg/b.w./mice	Attenuates centrizonal necrosis, fatty changes and liver cytochrome P450 enzymes. Also improve the level of LPO, GSH and SOD	PCM	Girish et al. (2009)
	40 mg/kg/b.w./rat	Enhance the level of SOD and CAT in liver tissue. Ameliorates degree of liver fibrosis	Diclofenac	Gupta et al. (2021)
	60 mg/kg/b.w./rat	A marked decrease in the level of AST, ALT, ALP, MDA, NO and expression of TNF- $\alpha$ and NF-kB. In addition, it significantly increased protect liver damage	Valproic acid	Abdelkader et al. (2020)
	50-100 mg/kg/b.w./mice	Alleviated hepatic antioxidant activities (GSH, CAT, MDA and SOD), pro-inflammatory cytokines levels (IL-6, IL-1 $\beta$ , and TNF- $\alpha$ ), genes expressions (Tlr4, Myd88, Cd14, Cox2, Nos2, and Nf $\kappa$ b1). Also, protect the histological architecture of the liver injury	Alcohol	Zhao et al. (2021)
	100 mg/kg/b.w./rat	Attenuates liver function (AST, ALT and ALP), antioxidant enzymes (GSH, CAT and SOD and fibrosis-related genes (MMP9 and MMP2)	thioacetamide	Afifi et al. (2018)
Naringenin	500 mg/kg/b.w./mice	Attenuates hepatocytes architecture by maintaining extended centrilobular necrosis, steatosis and fibrosis.	CCl <sub>4</sub>	Hermenean et al. (2014)
	20-80 mg/kg/b.w./rat	Increase the antioxidant status (SOD, CAT, GPx) while decreasing ALT, AST and LOP levels.	Nickel	Pari and Amudha (2011)



	Table 2 continued		
30-120 mg/kg/b.w./mice	Markedly decreased cytochrome P4502E1 expression, suppressed oxidative stress, inflammation, apoptosis, ALT, AST, and MAPK levels while increasing SOD, CAT and GSH level	CCl <sub>4</sub>	Dong et al. (2015)
100 mg/kgb.w./mice	Mitigated inflammation by suppressing NF-κB and NLRP3 pathways, CASP-3 and p53. Alleviated oxidative stress by inducing Nrf2 and increasing hepatic SOD activity and GSH	LPS/ d-GalN	Ahmedy et al. (2022)
20-50 mg/kg/b.w./rat	Inhibit hepatic fibrosis by reducing excess collagen accumulation in the liver. Also, inhibited the elevation of serum ALT, AST, ALP, and bilirubin levels	Dimethylnitrosam	ine Lee et al. (2004)

Paracetamol: PCM; Lipopolysaccharides: LPS; STZ; Acetaminophen: APAP; Sodium fluoride: NaF; Arylamide: ACR; 5-flourouracil: 5-FU; Methotrexate: MTX; Arylamide: ACR; d-galactosamine: d-GalN.



Catechin possesses anti-hyperlipidemic properties that can be used to treat the clinical symptoms of non-alcoholic fatty liver disorders, where abnormalities in protein and lipid metabolism play a significant role in the pathophysiology of the liver (Pezeshki et al., 2016). Epicatechin is crucial for the metabolism of lipids in hypercholesteremia and fatty liver disease. It can suppress the activity of crucial liver enzymes including SGPT and SGOT, which exacerbates the abnormalities in the liver (Shanmugam et al., 2017). Ferulic acid has strong antioxidant properties that can stop the free radical activity. By preventing extracellular matrix-related gene expression, it has a dominant anti-cholestatic effect on liver cholestasis. By improving lipid metabolism, it can frequently activate the MAPK or AMPK signalling pathway. According to numerous reports, ferulic acid regulates the expression of key physiological variables, including PPAR- and CPT-1, which is crucial for treating fatty liver illnesses (Saha et al., 2019). Naringenin, a natural flavonoid, possesses antioxidant, anticancer and anti-inflammatory activity. A naturally occurring flavonoid known as naringenin has antioxidant, anticancer, and anti-inflammatory properties. Although naringenin has very low direct antioxidant activity as a scavenger, it aids in the upregulation of Nrf2 pathways and maintains the normal redox of the cell even in clinical situations where prooxidants and reactive oxygen species are generated as a damage mechanism in hepatocytes (Esmaeili & Alilou, 2014; Saha et al., 2019). Resveratol produced an improvement in cellular antioxidant defence and markedly reversed LPO, enzymatic leakage, and Cyt P450 inactivation. increased levels of SOD mRNA and protein expression in hepatic cells increases the activities of SOD, CAT, and GPx while improving the level of AST and ALT, MDA, and protects against significant congestion and haemorrhage associated with the injury to the hepatic tissues. Additionally, resveratrol is linked to a significant decrease in the number of liver enzymes, cytokines, and transcriptional factors like NF-B. Following the activation of its inhibitor IB, it reduces NF-B expression to prevent liver injury (Kapil et al., 1995; Y. Zhang et al., 2015).

Considering the aforementioned fact, it is apparent that gape contains a huge number of hepatoprotective constituents that may protect the liver as a result of their antioxidant, antiapoptotic, and anti-inflammatory activities described in Table 2 and their hepatoprotective mechanism was shown in (Figure 2).

#### 3.5. Clinical studies of grape

A double-blind randomized clinical trial revealed that grape consumption increases the total antioxidant capacity in the body which will help to fight oxidative stress and inflammation (Sarkhosh-Khorasani et al., 2021). In another study, it is found that grape extract significantly attenuates the body weight and mass index, waist circumference, as well as waist-to-hip ratio and also lower TNF- $\alpha$ , and high sensitivity C-reactive protein levels (Parandoosh et al., 2020)

. Grape juice decreased LDL cholesterol, oxLDL and



apoB concentrations and raised HDL cholesterol and apoA-1 concentrations in healthy as well as hemodialysis-receiving adults (Castilla et al., 2006). In human male smokers, a polyphenol extract from grape seeds decreased the sensitivity of LDL to oxidation (Vigna et al., 2003). Furthermore, a reported study suggested that the intake of grapes or foods rich in polyphenols boosts vascular health, thereby significantly reducing the risk of cardiovascular diseases as well as diabetes mellitus (Rasines-Perea & Teissedre, 2017).

#### Table 3

D I		<pre>/</pre>	
Drug-d	lrug (	grape	) interactions

Drug	Effect of grapefruit/seeds/peels
Benzodiazepines	Substantial impact on the neurological system
Ciclosporin	Inhibits metabolism and elevate blood concentration
Coumarins	Interferes with the analysis of coumarins
Diazepam	Enhanced concentration
Felodipine	Enhances the effects and can cause increased blood pressure and heart rate, headache, flushing, and light-headedness
Lovastatin	Significantly increases the serum concentration
Nifedipine	Enhanced plasma concentration
Nisoldipine	Enhanced plasma concentration
Quinidine	Delays absorption and maximal effect on QT interval
Saquinavir	Enhanced AUC and Cmax
Terfenadine	Increases systemic availability, with cardiotoxicity at higher doses of grapefruit juice concentration
Triazolam	Enhanced plasma concentrations

#### 3.6. Drug-drug (grape interaction)

For millennia, medical care has undergone a global transformation for the development of synthetic drugs throughout the globe. Nevertheless, large portions of the populace in poor nations still rely on herbal medicine and its active constituents for their primary care because they thought that herbal medicine is safe and free from any type of side effects. Although therapies involving herbal products are established most of them have not been studied well, and their usage is either not monitored at all or is monitored very poorly. As a result, there is insufficient understanding of their mechanism of action, potential side effects, contraindications, and interactions with currently available conventional medications and functional foods to recommend the safe and sensible use of these medicines (Ekor, 2014).

Grapefruits, including the seeds and peels, are rare in that it is not used therapeutically, but because of the numerous bioactive compounds in grapefruit, it inhibits CYP3A4 might result in drug interactions. Dihydropyridine calcium channel blockers and HMG-CoA reductase inhibitors are the medications most frequently involved. Of the several hundred chemical entities in the grape that are responsible for these interactions. Hence, the safety of any herbal medicine is important as like efficacy and it is needed to assure that all herbal drugs are safe and of suitable quality. Some examples of drug-drug interaction in the context of grapes are presented in Table 3 (Aronson, 2006).

# 4. CONCLUSION AND FUTURE PROSPECTIVE

Grapes, one of the most widely consumed fruits in the world and are used in the traditional system of medicine to treat or prevent a variety of diseases. The polyphenols along with vitamins and fibre found in grape skin and seeds can be used as a functional component to treating various health ailments. In the current study, through numerous research studies, it has been established that grapes and polyphenols are the potentials to protect against liver toxicity and they may replace or enhance currently prescribed synthetic drugs for the treatment of hepatotoxicity by adding them to other efficient pharmaceutical formulations.

Herbal medicine including grape or polyphenols, continue to gain recognition and application on a global scale. Contrary to the erroneous belief that herbal medicine is "safe" because they are produced from "natural" sources, issues linked to adverse responses have also recently become more apparent, more common, and unassailable. In order to safeguard the health of the public, it is necessary to ensure the safety and quality of all herbal medications. In the meanwhile, using herbal medicines correctly can have positive therapeutic effects on users with lower or minimal associated risk. However, it should be understood that there are some adverse effects associated with using herbal products. Additionally, the use of adulterated herbal materials and improper formulation must be prohibited because they might result in the production of inferior and lifethreatening herbal medications. Though, thorough scientific investigations (both preclinical and clinical), analysis, and suitable laws and regulations should be followed strictly to ensure the efficacy, safety and quality of herbal medicines.

Conclusively, it is plausible to conclude that grapes and polyphenols have a promising future in the healthcare system and may eventually replace or be used as an adjuvant to synthetic medicine for the management of hepatotoxicity.

## 5. CONFLICTS OF INTEREST

None

#### ORCID

Shabina Khan 0000-0	0002-4956-1998
---------------------	----------------

Mohammad Ibrahim 0000-0002-5780-4232

## AUTHOR CONTRIBUTIONS

S. K - Collected data formatted and drafted the manuscript. M. I -Design the study, analyse data and corrected the manuscript.

### REFERENCES

Abdel-Ghaffar, O., Mahmoud, S.T., Said, A.A., Sanad, F.A.A.Y., 2017. Hepatoprotective effect of rutin against oxidative stress of isoniazid in albino rats. International Journal of Pharmacology. 13(6), 516–528. https://dx.doi.org/10.3923/ijp.2017.516.528

- Abdelkader, N.F., Elyamany, M., Gad, A.M., Assaf, N., Fawzy, H.M., Elesawy, W.H., 2020. Ellagic acid attenuates liver toxicity induced by valproic acid in rats. Journal of Pharmacological Sciences. 143(1), 23– 29. https://doi.org/10.1016/j.jphs.2020.01.007
- Abo-Salem, O.M., Abd-Ellah, M.F., Ghonaim, M.M., 2011. Hepatoprotective activity of quercetin against acrylonitrile-induced hepatotoxicity in rats. Journal of Biochemical and Molecular Toxicology. 25(6), 386–392. https://doi.org/10.1002/jbt.20406
- Afifi, N.A., Ibrahim, M.A., Galal, M.K., 2018. Hepatoprotective influence of quercetin and ellagic acid on thioacetamide-induced hepatotoxicity in rats. Canadian Journal of Physiology and Pharmacology. 96(6), 624–629. https://doi.org/10.1139/cjpp-2017-0651
- Ahmedy, O.A., Salem, H.H., Sayed, N.H., Ibrahim, S.M., 2022. Naringenin affords protection against lipopolysaccharide/D-galactosamineinduced acute liver failure: role of autophagy. Archives of Biochemistry and Biophysics. 717, 109121. https://doi.org/10.1016/j .abb.2022.109121
- Alam, N., Ikram, R., 2018. Methylphenidate-induced hepatotoxicity in rats and its reduction by buspirone. Pakistan Journal of Pharmaceutical Sciences. 31(3), 29716850–29716850.
- Al-Amoudi, W.M., 2017. Effect of grapefruit juice on aluminuminduced hepatotoxicity in albino rats: histological, ultrastructural and histochemical assessment. Advances in Bioscience and Biotechnology. 8(12), 463–477. https://doi.org/10.4236/abb.2017.812034
- Al-Baqami, N.M., Hamza, R.Z., 2021. Protective effect of resveratrol against hepatotoxicity of cadmium in male rats: Antioxidant and histopathological approaches. Coatings. 11(5), 594. https://doi.org/ 10.3390/coatings11050594
- Alkinani, K.B., Ali, E.M., Al-Shaikh, T.M., Khan, J.A.A., Al-Naomasi, T.M., Ali, S.S., Abduljawad, A.A., Mosa, O.F., Zafar, T.A., 2021. Hepatoprotective Effects of (-) Epicatechin in CCl4-Induced Toxicity Model Are Mediated via Modulation of Oxidative Stress Markers in Rats. Evidence-based Complementary and Alternative Medicine, 4655150. https://doi.org/10.1155/2021/4655150
- Amedei, A., Niccolai, E., 2014. Plant and Marine Sources: Biological activity of natural products and therapeutic use, V. Havlíček, J. Spížek, et al., (Eds.), Natural Products Analysis: Instrumentation, Methods, and Applications. John Wiley & Sons,, pp. 43–113. https://doi.org/ 10.1002/9781118876015.ch4
- Ansar, S., Siddiqi, N.J., Zargar, S., Ganaie, M.A., Abudawood, M., 2016. Hepatoprotective effect of Quercetin supplementation against Acrylamide-induced DNA damage in wistar rats. BMC Complementary and Alternative Medicine. 16(1), 327. https://doi.org/10.1186/ s12906-016-1322-7
- Aronson, J.K., 2006. Meyler's Side Effects of Drugs: The International Encyclopedia of Adverse Drug Reactions and Interactions.
- Azadnasab, R., Kalantar, H., Khorsandi, L., Kalantari, H., Khodayar, M.J., 2021. Epicatechin ameliorative effects on methotrexate-induced hepatotoxicity in mice. Human and Experimental Toxicology. 40(12), S603–S610. https://doi.org/ 10.1177/09603271211047924
- Barros, P.P., Silva, G.H.D., Gonçalves, G.M.S., Oliveira, J.C., Pagnan, L.G., Arco-E-Flexa, L., 2017. Hepatoprotective effect of quercetin pretreatment against paracetamol-induced liver damage and partial hepatectomy in rats. Brazilian Archives of Biology and Technology. 60, e17160138. https://doi.org/10.1590/1678-4324 -2016160138
- Bashar, S.M., Elhadidy, M.G., Mostafa, A.F., Hamed, B., Helmy, S., Abd-Elmoniem, H.A., 2021. Hepatoprotective effect of gallic acid against type 2-induced diabetic liver injury in male rats through modulation of fetuin-A and GLP-1 with involvement of ERK1/2/NF-



 $\kappa$ B and Wnt1/ $\beta$ -catenin signaling pathways. General Physiology and Biophysics. 40(3), 221–234. https://doi.org/10.4149/gpb\_2021005

- Bessone, F., Hernandez, N., Tagle, M., Arrese, M., Parana, R., Mendez-Sánchez, N., Ridruejo, E., Mendizabal, M., Dagher, L., Contreras, F., Fassio, E., Pesoa, M., Brahm, J., Silva, M., 2021. Drug-induced liver injury: A management position paper from the Latin American Association for Study of the liver. Annals of Hepatology. 24, 100321. https://doi.org/10.1016/j.aohep.2021.100321
- Bilgic, S., Dogan, Z., Azirak, S., Erdemli, M.E., Onderci, M., Turk, A., Ozer, M.K., 2018. Hepatoprotective effect of royal jelly, grape seed extract, and *Lycium barbarum* against diethylnitrosamine-induced liver toxicity in rats. Annals of Medical Research. 25(4), 342–248. https://doi.org/10.5455/annalsmedres.2018.03.040
- Binmowyna, M.N., Alfaris, N.A., 2021. Kaempferol suppresses acetaminophen-induced liver damage by upregulation/activation of SIRT1. Pharmaceutical Biology. 59(1), 146–156. https://doi.org/10 .1080/13880209.2021.1877734
- Brunetti, L., Song, J.H., Suh, D., Kim, H.J., Seong, Y.H., Lee, D.S., Lee, S.M., Suh, D.C., 2020. The risk of vancomycin toxicity in patients with liver impairment. Annals of Clinical Microbiology and Antimicrobials. 19(1), 13. https://doi.org/10.1186/s12941-020 -00354-2
- Buchner, I., Medeiros, N., Lacerda, D.D.S., Normann, C.A.B., Gemelli, T., Rigon, P., Wannmacher, C.M.D., Henriques, J.A.P., Dani, C., Funchal, C., 2014. Hepatoprotective and antioxidant potential of organic and conventional grape juices in rats fed a high-fat diet. Antioxidants. 3(2), 323–338. https://doi.org/10.3390/ antiox3020323
- Castilla, P., Echarri, R., Dávalos, A., Cerrato, F., Ortega, H., Teruel, J.L., Lucas, M.F., Gómez-Coronado, D., Ortuño, J., Lasunción, M.A., 2006. Concentrated red grape juice exerts antioxidant, hypolipidemic, and antiinflammatory effects in both hemodialysis patients and healthy subjects. American Journal of Clinical Nutrition. 84(1), 252– 262. https://doi.org/10.1093/ajcn/84.1.252
- Chen, W.M., Shaw, L.H., Chang, P.J., Tung, S.Y., Chang, T.S., Shen, C.H., Hsieh, Y.Y., Wei, K.L., 2016. Hepatoprotective effect of resveratrol against ethanol-induced oxidative stress through induction of superoxide dismutase *in vivo* and *in vitro*. Experimental and Therapeutic Medicine. 11(4), 1231–1238. https://doi.org/10.3892/ etm.2016.3077
- Chen, Z., Yang, Y., Mi, S., Fan, Q., Sun, X., Deng, B., Wu, G., Li, Y., Zhou, Q., Ruan, Z., 2019. Hepatoprotective effect of chlorogenic acid against chronic liver injury in inflammatory rats. Journal of Functional Foods. 62, 103540–103540. https://doi.org/10.1016/j.jff .2019.103540
- Conway, R., Carey, J.J., 2017. Risk of liver disease in methotrexate treated patients. World Journal of Hepatology. 9(26), 1092–1100. https://doi.org/10.4254/wjh.v9.i26.1092
- Devarbhavi, H., 2012. An Update on Drug-induced Liver Injury. Journal of Clinical and Experimental Hepatology. 2(3), 247–259. https:// doi.org/10.1016/j.jceh.2012.05.002
- Dogan, A., Celik, I., 2012. Hepatoprotective and antioxidant activities of grapeseeds against ethanol-induced oxidative stress in rats. British Journal of Nutrition. 107(1), 45–51. https://doi.org/10.1017/ s0007114511002650
- Domitrović, R., Jakovac, H., Marchesi, V.V., Vladimir-Knežević, S., Cvijanović, O., Ž Tadić, Ž Romić, Rahelić, D., 2012. Differential hepatoprotective mechanisms of rutin and quercetin in CCl4intoxicated BALB/cN mice. Acta Pharmacologica Sinica. 33(10), 1260–1270. https://doi.org/10.1038/aps.2012.62
- Dong, D., Xu, L., Yin, L., Qi, Y., Peng, J., 2015. Naringin prevents carbon tetrachloride-induced acute liver injury in mice. Journal of Functional

Foods. 12, 179–191. https://doi.org/10.1016/j.jff.2014.11.020

- Dönmez, Y.E., Özcan, Ö., Soylu, N., Sarıoğlu, F.K., Selimoğlu, A., 2017. Management of hepatotoxicity induced by the use of olanzapine. Journal of Child and Adolescent Psychopharmacology. 27(3), 293– 294. https://doi.org/10.1089/cap.2016.0178
- Douglas, I.J., Langham, J., Bhaskaran, K., Brauer, R., Smeeth, L., 2013. Orlistat and the risk of acute liver injury: self-controlled case series study in UK Clinical Practice Research Datalink. British Medical Journal. 346, f1936. https://doi.org/10.1136/bmj.f1936
- Ekor, M., 2014. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. Frontiers in Neurology. 10(4), 177–177. https://doi.org/10.3389/fphar.2013 .00177
- Esmaeili, M.A., Alilou, M., 2014. Naringenin attenuates CC l4-induced hepatic inflammation by the activation of an Nrf2-mediated pathway in rats. Clinical and Experimental Pharmacology and Physiology. 41(6), 416–422. https://doi.org/10.1111/1440-1681.12230
- Faras, A.A.E., Elsawaf, A.L., 2017. Hepatoprotective activity of quercetin against paracetamol-induced liver toxicity in rats. Tanta Medical Journal. 45(2), 92–92.
- Farzaei, M.H., Zobeiri, M., Parvizi, F., El-Senduny, F.F., Marmouzi, I., Coy-Barrera, E., Naseri, R., Nabavi, S.M., Rahimi, R., Abdollahi, M., 2018. Curcumin in liver diseases: a systematic review of the cellular mechanisms of oxidative stress and clinical perspective. Nutrients. 10(7), 855–855. https://doi.org/10.3390/nu10070855
- Fernandez, S., Lenoir, C., Samer, C.F., Rollason, V., 2021. Drug-drug interactions leading to adverse drug reactions with rivaroxaban: A systematic review of the literature and analysis of vigibase. Journal of Personalized Medicine. 11(4), 250–250. https://doi.org/10.3390/ jpm11040250
- Fountain, F.F., Tolley, E.A., Jacobs, A.R., Self, T.H., 2009. Rifampin hepatotoxicity associated with treatment of latent tuberculosis infection. American Journal of the Medical Sciences. 337(5), 317– 320. https://doi.org/10.1097/maj.0b013e31818c0134
- Gad, R.A., El-Latif, A., Shoka, A.A.A., 2013. Hepatoprotective Activity of Citrus Fruits Peel Extracts on Lipopolysaccharide: D-Galactosamine induced Liver Damage in Rats. Egyptian Journal of Zoology. 60(60), 245–266. https://doi.org/10.12816/0003293
- Ganesan, R.S., Eswaran, M., Thakur, S., Jayaraman, A., 2015. Reversal of phenytoin induced hepatotoxicity by alpha lipoic acid in rats. African Journal of Pharmacy and Pharmacology. 9(7), 198–204.
- Gao, F.F., Lv, J.W., Wang, Y., Fan, R., Li, Q., Zhang, Z., Wei, L., 2016. Tamoxifen induces hepatotoxicity and changes to hepatocyte morphology at the early stage of endocrinotherapy in mice. cal Reports. 4(1), 102–106. https://doi.org/10.3892/br.2015.536
- Gayam, V., Mandal, A.K., Khalid, M., Shrestha, B., Garlapati, P., Khalid, M., 2018. Valproic acid induced acute liver injury resulting in hepatic encephalopathy-a case report and literature review. Journal of Community Hospital Internal Medicine Perspectives. 8(5), 311–314. https://doi.org/10.1080/20009666.2018.1514933
- Gelen, V., Şengül, E., Gedikli, S., Atila, G., Uslu, H., Makav, M., 2017. The protective effect of rutin and quercetin on 5-FU-induced hepatotoxicity in rats. Asian Pacific Journal of Tropical Biomedicine. 7(7), 647–653. https://doi.org/10.1016/j.apjtb.2017.06.013
- Giribabu, N., Karim, K., Kilari, E.K., Kassim, N.M., Salleh, N., 2018. Anti-inflammatory, antiapoptotic and proproliferative effects of *Vitis vinifera* seed ethanolic extract in the liver of streptozotocinnicotinamide-induced type 2 diabetes in male rats. Canadian Journal of Diabetes. 42(2), 138–149. https://doi.org/10.1016/j.jcjd.2017.04 .005
- Girish, C., Koner, B.C., Jayanthi, S., Rao, K.R., Rajesh, B., Pradhan, S.C., 2009. Hepatoprotective activity of picroliv, curcumin and ellagic acid



**View Article Online** 

compared to silymarin on paracetamol induced liver toxicity in mice. Fundamental and Clinical Pharmacology. 23(6), 735–745. https:// doi.org/10.1111/j.1472-8206.2009.00722.x

- Gomez-Peralta, F., Velasco-Martínez, P., Abreu, C., Cepeda, M., Fernández-Puente, M., 2018. Hepatotoxicity in hyperthyroid patient after consecutive methimazole and propylthiouracil therapies. Diabetes & Metabolism Case Reports. 2018, 1–4. https://doi.org/10 .1530/edm-17-0173
- Guicciardi, M.E., Malhi, H., Mott, J.L., Gores, G.J., 2013. Apoptosis and necrosis in the liver. Comprehensive Physiology. 3(2), 977–1010. https://doi.org/10.1002/cphy.c120020
- Gupta, A., Kumar, R., Ganguly, R., Singh, A.K., Rana, H.K., Pandey, A.K., 2021. Antioxidant, anti-inflammatory and hepatoprotective activities of *Terminalia bellirica* and its bioactive component ellagic acid against diclofenac induced oxidative stress and hepatotoxicity. Toxicology Reports. 8, 44–52. https://doi.org/ 10.1016/j.toxrep.2020.12.010
- Hafez, M.M., Al-Harbi, N.O., Al-Hoshani, A.R., Al-Hosaini, K.A., Shrari, S.D., Rejaie, S.S.A., Sayed-Ahmed, M.M., Al-Shabanah, O.A., 2015. Hepato-protective effect of rutin via IL-6/STAT3 pathway in CCl4-induced hepatotoxicity in rats. Biological Research. 48, 30. https://doi.org/10.1186/s40659-015-0022-y
- Haghighatdoost, F., Gholami, A., Hariri, M., 2020. Effect of grape polyphenols on selected inflammatory mediators: A systematic review and meta-analysis randomized clinical trials. EXCLI Journal. 19, 251– 267. https://doi.org/10.17179/excli2020-1011
- Hajdarevic, B., Vehabovic, I., Catic, T., Masic, I., 2020. The Role of Diet Therapy in the Treatment of Liver Disease. Materia Socio Medica. 32(3), 200–206. https://doi.org/10.5455/msm.2020.32.200-206
- Hermenean, A., Ardelean, A., Stan, M., Hadaruga, N., Mihali, C.V., Costache, M., Dinischiotu, A., 2014. Antioxidant and hepatoprotective effects of naringenin and its β-cyclodextrin formulation in mice intoxicated with carbon tetrachloride: a comparative study. Journal of Medicinal Food. 17(6), 670–677. https://doi.org/10.1089/jmf.2013 .0007
- Hsu, Y.W., Chen, Y.Y., Tsai, C.F., 2022. Protective Effects of Chlorogenic Acid against Carbon Tetrachloride-Induced Hepatotoxicity in Mice. Processes. 10(1), 31. https://doi.org/10.3390/pr10010031
- Huang, Z.Q., Chen, P., Su, W.W., Wang, Y.G., Wu, H., Peng, W., Li, P.B., 2018. Antioxidant activity and hepatoprotective potential of quercetin 7-rhamnoside *in vitro* and *in vivo*. Molecules. 23(5), 1188. https:// doi.org/10.3390/molecules23051188
- Ibrahim, M., Parveen, B., Zahiruddin, S., Gautam, G., Parveen, R., Khan, M.A., Gupta, A., Ahmad, S., 2022. Analysis of polyphenols in *Aegle marmelos* leaf and ameliorative efficacy against diabetic mice through restoration of antioxidant and anti-inflammatory status. Journal of Food Biochemistry. 46(4), e13852. https://doi.org/10 .1111/jfbc.13852
- Ismail, A.F., Salem, A.A., Eassawy, M.M., 2016. Hepatoprotective effect of grape seed oil against carbon tetrachloride induced oxidative stress in liver of γ-irradiated rat. Journal of Photochemistry and Photobiology B: Biology. 160, 1–10. https://doi.org/10.1016/j.jphotobiol.2016.03 .027
- Itoh, A., Isoda, K., Kondoh, M., Kawase, M., Watari, A., Kobayashi, M., Tamesada, M., Yagi, K., 2010. Hepatoprotective effect of syringic acid and vanillic acid on CCl4-induced liver injury. Biological and Pharmaceutical Bulletin. 33(6), 983–987. https://doi.org/10.1248/ bpb.33.983
- Kamal, P., Bhabhra, R., 2019. SUN-185 Glipizide Induced Hepatotoxicity. Diabetes & Metabolic Disorders. 6(1), SUN–185. https:// doi.org/10.1210/js.2019-SUN-185

Kapil, A., Koul, I.B., Suri, O.P., 1995. Antihepatotoxic effects

of chlorogenic acid from *Anthocephalus cadamba*. Phytotherapy Research. 9(3), 189–193. https://doi.org/10.1002/ptr.2650090307

- Ko, M.S., Choi, Y.H., Jung, S.H., Lee, J.S., Kim, H.S., Lee, C.H., Kim, S.G., 2015. Tacrolimus therapy causes hepatotoxicity in patients with a history of liver disease. International Journal of Clinical Pharmacology and Therapeutics. 53(5), 363–371. https://doi.org/10 .5414/cp202226
- Koriem, K.M., Soliman, R.E., 2014. Chlorogenic and caftaric acids in liver toxicity and oxidative stress induced by methamphetamine. Journal of Toxicology, 583494. https://doi.org/10.1155/2014/583494
- Laster, J., Satoskar, R., 2014. Aspirin-induced acute liver injury. ACG Case Reports Journal. 2(1), 48–49. https://doi.org/10.14309/crj .2014.81
- Latief, U., Husain, H., Mukherjee, D., Ahmad, R., 2016. Hepatoprotective efficacy of gallic acid during Nitrosodiethylamine-induced liver inflammation in Wistar rats. The Journal of Basic & Applied Zoology. 76, 31–41. https://doi.org/10.1016/j.jobaz.2016.07.002
- Lee, M.H., Yoon, S., Moon, J.O., 2004. The flavonoid naringenin inhibits dimethylnitrosamine-induced liver damage in rats. Biological and Pharmaceutical Bulletin. 27(1), 72–76. https://doi.org/10.1248/ bpb.27.72
- Li, M., Wang, X.F., Shi, J.J., Li, Y.P., Yang, N., Zhai, S., Dang, S.S., 2015. Caffeic acid phenethyl ester inhibits liver fibrosis in rats. World Journal of Gastroenterology. 21(13), 3893–3903. https://doi.org/10 .3748/wjg.v21.i13.3893
- Li, S., Tan, H.Y., Wang, N., Cheung, F., Hong, M., Feng, Y., 2018. The Potential and Action Mechanism of Polyphenols in the Treatment of Liver Diseases. Oxidative Medicine and Cellular Longevity. 4, 8394818. https://doi.org/10.1155/2018/8394818
- Liao, M.F., Yen, S.C., Lin, C.Y., Lyu, R.K., 2013. Delayed liver function impairment secondary to interferon β-1a use in multiple sclerosis. Case Reports in Neurology. 5(2), 130–134. https://doi.org/10.1159/ 000354042
- Liu, J., Lu, J.F., Wen, X.Y., Kan, J., Jin, C.H., 2015. Antioxidant and protective effect of inulin and catechin grafted inulin against CCl4-induced liver injury. International Journal of Biological Macromolecules. 72, 1479–1484. https://doi.org/10.1016/j.ijbiomac.2014 .09.066
- Madrigal-Santillán, E., Madrigal-Bujaidar, E., Álvarez González, I., Sumaya-Martínez, M.T., Gutiérrez-Salinas, J., Bautista, M., Morales-González, A., González-Rubio, M.G.L., Aguilar-Faisal, J.L., Morales-González, J.A., 2014. Review of natural products with hepatoprotective effects. World Journal of Gastroenterology. 20(40), 14787– 14804. https://doi.org/10.3748/wjg.v20.i40.14787
- Mahmoud, M., Hamdan, D., Wink, M., El-Shazly, A., 2013. Naringin and rutin prevent d-galactosamine-induced hepatic injury in rats via attenuation of the inflammatory cascade and oxidative stress. European Scientific Journal. 9(30), 1857–7881.
- Mani, G.D.M., Balamuralikrishnan, B., Manikandan, R., Sampathkumar, P., Arun, M., Lakshminarayanan, R.R., Liu, W.C., Anand, V.A., 2022. Effects of *Euphorbia thymifolia* and *Euphorbia hirta* leaf extracts on membrane-bound, mitochondrial enzymes and lipid profile of carbon tetrachloride-induced hepatotoxicity in rats. Natural Resources for Human Health. 2(4), 443–449. https://doi.org/10.53365/nrfhh/146770
- Manju, M., Jagadeesan, G., 2019. *In vivo* hepatoprotective effect of caffeic acid on mercuric chloride-induced biochemical changes in albino wistar rats. Asian Journal of Pharmaceutical and Clinical Research. 12(4), 119–119. https://doi.org/10.22159/ajpcr.2019.v12i4.31609
- Morales, M., Vélez, L., L, Muñoz, N., M, G, O., 2016. Hepatotoxicity: a drug-induced cholestatic pattern. Revista Colombiana de Gastroenterologia. 31(1), 36–47.



- Nabavi, S.F., Nabavi, S.M., Habtemariam, S., Moghaddam, A.H., Sureda, A., Jafari, M., Latifi, A.M., 2013. Hepatoprotective effect of gallic acid isolated from *Peltiphyllum peltatum*- against sodium fluoride-induced oxidative stress. Industrial Crops and Products. 44, 50–55. https://doi.org/10.1016/j.indcrop.2012.10.024
- Nada, S.A., Gowifel, A.M., El-Denshary, E.E.S., Salama, A.A., Khalil, M.G., Ahmed, K.A., 2015. Protective effect of grape seed extract and/or silymarin against thioacetamide-induced hepatic fibrosis in rats. Journal of Liver. 4(178), 2167–0889.
- Ojeaburu, S.I., Oriakhi, K., 2021. Hepatoprotective, antioxidant and, anti-inflammatory potentials of gallic acid in carbon tetrachlorideinduced hepatic damage in Wistar rats. Toxicology Reports. 8, 177– 185. https://doi.org/10.1016/j.toxrep.2021.01.001
- Okkay, I.F., Okkay, U., Gundogdu, O.L., Bayram, C., Mendil, A.S., Ertugrul, M.S., Hacimuftuoglu, A., 2022. Syringic acid protects against thioacetamide-induced hepatic encephalopathy: Behavioral, biochemical, and molecular evidence. Neuroscience Letters, 769–769. https://doi.org/10.1016/j.neulet.2021.136385
- Parandoosh, M., Yousefi, R., Khorsandi, H., Nikpayam, O., Saidpour, A., Babaei, H., 2020. The effects of grape seed extract (*Vitis vinifera*) supplement on inflammatory markers, neuropeptide Y, anthropometric measures, and appetite in obese or overweight individuals: A randomized clinical trial. Phytotherapy Research. 34(2), 379–387. https://doi.org/10.1002/ptr.6529
- Pari, L., Amudha, K., 2011. Hepatoprotective role of naringin on nickel-induced toxicity in male Wistar rats. European Journal of Pharmacology. 650(1), 364–370. https://doi.org/10.1016/j.ejphar .2010.09.068
- Perdices, E.V., Medina-Cáliz, I., Hernando, S., Ortega, A., Martín-Ocaña, F., Navarro, J.M., Peláez, G., Castiella, A., Hallal, H., Romero-Gómez, M., Gonzalez-Jimenez, A., 2014. Hepatotoxicity associated with statin use: analysis of the cases included in the Spanish Hepatotoxicity Registry. Revista Espanola de Enfermedades Digestivas. 106(4), 25075655–25075655.
- Pezeshki, A., Safi, S., Feizi, A., Askari, G., Karami, F., 2016. The effect of green tea extract supplementation on liver enzymes in patients with nonalcoholic fatty liver disease. International Journal of Preventive Medicine. 7, 28–28. https://doi.org/10.4103/2008-7802.173051
- Raj, P.V., Nitesh, K., Gang, S.S., Jagani, V.H., Chandrashekhar, H.R., Rao, J.V., Rao, C.M., Udupa, N., 2010. Protective role of catechin on d-galactosamine induced hepatotoxicity through a p53 dependent pathway. Indian Journal of Clinical Biochemistry. 25(4), 349–356. https://doi.org/10.1007/s12291-010-0073-3
- Raschi, E., De Ponti, F., 2015. Drug-and herb-induced liver injury: progress, current challenges and emerging signals of post-marketing risk. World Journal of Hepatology. 7(13), 1761–1771. https://doi .org/10.4254/wjh.v7.i13.1761
- Rasines-Perea, Z., Teissedre, P.L., 2017. Grape polyphenols' effects in human cardiovascular diseases and diabetes. Molecules. 22(1), 68. https://doi.org/10.3390/molecules22010068
- Rasool, M.K., Sabina, E.P., Ramya, S.R., Preety, P., Patel, S., Mandal, N., Mishra, P.P., Samuel, J., 2010. Hepatoprotective and antioxidant effects of gallic acid in paracetamol-induced liver damage in mice. Journal of Pharmacy and Pharmacology. 62(5), 638–643. https:// doi.org/10.1211/jpp.62.05.0012
- Reddy, M.K., Reddy, A.G., Kumar, B.K., Madhuri, D., Boobalan, G., Reddy, M.A., 2017. Protective effect of rutin in comparison to silymarin against induced hepatotoxicity in rats. Veterinary World. 10(1), 74–80. https://doi.org/10.14202/vetworld.2017.74-80
- Rochwerg, B., Xenodemetropoulos, T., Crowther, M., Spyropoulos, A., 2012. Dabigatran-induced acute hepatitis. Clinical and Applied Thrombosis/Hemostasis. 18(5), 549–50. https://doi.org/10.1177/

1076029611435840

- Saha, P., Talukdar, A.D., Nath, R., Sarker, S.D., Nahar, L., Sahu, J., Choudhury, M.D., 2019. Role of natural phenolics in hepatoprotection: a mechanistic review and analysis of regulatory network of associated genes. Frontiers in Pharmacology. 24, 509. https://doi.org/ 10.3389/fphar.2019.00509
- Sarkhosh-Khorasani, S., Sangsefidi, Z.S., Hosseinzadeh, M., 2021. The effect of grape products containing polyphenols on oxidative stress: a systematic review and meta-analysis of randomized clinical trials. Nutrition Journal. 20(1), 25. https://doi.org/10.1186/s12937-021 -00686-5
- Shanmugam, B., Shanmugam, K.R., Ravi, S., Subbaiah, G.V., Ramakrishana, C., Mallikarjuna, K., Reddy, K.S., 2017. Exploratory studies of (-)-Epicatechin, a bioactive compound of *Phyllanthus niruri*, on the antioxidant enzymes and oxidative stress markers in D-galactosamine-induced hepatitis in rats: A study with reference to clinical prospective. Pharmaconosy Magazine. 13, S56–S62. https://doi.org/10.4103/0973-1296.203973
- Singh, D., Cho, W.C., Upadhyay, G., 2016. Drug-induced liver toxicity and prevention by herbal antioxidants: an overview. Frontiers in Physiology. 6, 363–363. https://doi.org/10.3389/fphys.2015.00363
- Udom, G.J., Yemitan, O.K., Umoh, E.E., Mbagwu, H.O., Ukpe, E.E., Thomas, P.S., 2018. Hepatoprotective properties of ethanol seed extract of *Citrus paradisi* Macfad (grape fruit) against paracetamolinduced hepatotoxicity in Wistar rats. Journal of Herbal Drugs. 8(4), 219–225.
- Vigna, G.B., Costantini, F., Aldini, G., Carini, M., Catapano, A., Schena, F., Tangerini, A., Zanca, R., Bombardelli, E., Morazzoni, P., Mezzetti, A., 2003. Effect of a standardized grape seed extract on low-density lipoprotein susceptibility to oxidation in heavy smokers. Metabolism: Clinical and Experimental. 52(10), 1250–1257. https:// doi.org/10.1016/s0026-0495(03)00192-6
- Wang, M., Sun, J., Jiang, Z., Xie, W., Zhang, X., 2015. Hepatoprotective effect of kaempferol against alcoholic liver injury in mice. American Journal of Chinese Medicine. 43(2), 241–254. https://doi.org/10 .1142/s0192415x15500160
- Wiegand, J., Berg, T., 2013. The etiology, diagnosis and prevention of liver cirrhosis: part 1 of a series on liver cirrhosis. Deutsches Ärzteblatt international. 110(6), 85–91. https://doi.org/10.3238/arztebl.2013 .0085
- Xu, Y., Chen, J., Yu, X., Tao, W., Jiang, F., Yin, Z., Liu, C., 2010. Protective effects of chlorogenic acid on acute hepatotoxicity induced by lipopolysaccharide in mice. Inflammation Research. 59(10), 871– 877. https://doi.org/10.1007/s00011-010-0199-z
- Xu, Y., Guo, W., Zhang, C., Chen, F., Tan, H.Y., Li, S., Wang, N., Feng, Y., 2020. Herbal medicine in the treatment of non-alcoholic fatty liver diseases-efficacy, action mechanism, and clinical application. Frontiers in Pharmacology. 12(11), 601–601. https://doi.org/ 10.3389/fphar.2020.00601
- Yan, J.Y., Nie, X.L., Tao, Q.M., Zhan, S.Y., De Zhang, Y., 2013. Ketoconazole associated hepatotoxicity: a systematic review and metaanalysis. Biomedical and Environmental Sciences. 26(7), 605–610. https://doi.org/10.3967/0895-3988.2013.07.013
- Yang, N., Dang, S., Shi, J., Wu, F., Li, M., Zhang, X., Li, Y., Jia, X., Zhai, S., 2017. Caffeic acid phenethyl ester attenuates liver fibrosis via inhibition of TGF-β1/Smad3 pathway and induction of autophagy pathway. Biochemical and Biophysical Research Communications. 486(1), 22–28. https://doi.org/10.1016/j.bbrc.2017.02.057
- Yew, W.W., Leung, C.C., 2006. Antituberculosis drugs and hepatotoxicity. Respirology. 11(6), 699–707. https://doi.org/10.1111/j.1440-1843 .2006.00941.x
- Yoon, E., Babar, A., Choudhary, M., Kutner, M., Pyrsopoulos, N., 2016.



Acetaminophen-induced hepatotoxicity: a comprehensive update. Journal of Clinical and Translational Hepatology. 4(2), 131–142. https://doi.org/10.14218/jcth.2015.00052

- Zang, Y., Zhang, D., Yu, C., Jin, C., Igarashi, K., 2017. Antioxidant and hepatoprotective activity of kaempferol 3-O-β-D-(2, 6-di-O-α-L-rhamnopyranosyl) galactopyronoside against carbon tetrachlorideinduced liver injury in mice. Food Science and Biotechnology. 26(4), 1071–1076. https://doi.org/10.1007/s10068-017-0170-7
- Zhang, X., Shi, L., Chen, R., Zhao, Y., Ren, D., Yang, X., 2021. Chlorogenic acid inhibits trimethylamine-N-oxide formation and remodels intestinal microbiota to alleviate liver dysfunction in high

L-carnitine feeding mice. Food and Function. 12(21), 10500–10511. https://doi.org/10.1039/D1FO01778K

- Zhang, Y., Chen, M.L., Zhou, Y., Yi, L., Gao, Y.X., Ran, L., Chen, S.H., Zhang, T., Zhou, X., Zou, D., Wu, B., 2015. https://doi.org/10. 1002/mnfr.201500016. Molecular Nutrition & Food Research. 59, 1443–1457. https://doi.org/10.1002/mnfr.201500016
- Zhao, L., Mehmood, A., Soliman, M.M., Iftikhar, A., Iftikhar, M., Aboelenin, S.M., Wang, C., 2021. Protective effects of ellagic acid against alcoholic liver disease in mice. Frontiers in Nutrition. 8, 744520. https://doi.org/10.3389/fnut.2021.744520

